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Alpha7 Nicotinic Acetylcholine Receptor Mediates Nicotine-induced Apoptosis and Cell Cycle Arrest of Hepatocellular Carcinoma HepG2 Cells

Running title: $\alpha 7$ nAChR and apoptosis of HepG2 cells

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Abstract

Purpose: The cytotoxic properties upon treatment with nicotine have been reported in several studies, but the underlying mechanisms remain not fully defined. The alpha7 nicotinic acetylcholine receptor ($\alpha 7$ nAChR) is one of the important nicotinic receptors, which nicotine partly by binding to this receptor exerts its effects. The current study aimed to investigate the influences of nicotine on cellular proliferative and apoptotic activities and tried to determine the involvement of $\alpha 7$ nAChR in these functions.

Methods: Human hepatocellular carcinoma (HepG2) cell line was used to determine the individual or combined effects of treatments with nicotine (10 μ M) and specific siRNA (100 nM) targeting $\alpha 7$ nAChR expression. The MTT assay, DAPI staining assay, and flow cytometry assay were applied to measure the cell viability, apoptosis and cell cycle progression of the cells, respectively. In addition, the changes in the mRNA level of the genes were assessed by qRT-PCR.

Results: Compared to control groups, the cells treated with nicotine exhibited significant dose-dependent decreases in cell viability (log IC₅₀ = -5.12 \pm 0.15). Furthermore, nicotine induced

apoptosis and cell cycle arrest especially at G2/M Phase. The qRT-PCR revealed that nicotine increased the mRNA levels of $\alpha 7$ nAChR as well as caspase-3 and suppressed the expression of cyclin B1. Treatment with $\alpha 7$ -siRNA abolished these effects of nicotine.

Conclusion: These experiments determined that upregulation of $\alpha 7$ nAChR by nicotine inhibits HepG2 cells proliferation and induces their apoptosis. These effects blocked by treatment with $\alpha 7$ -siRNA, which indicates the involvement of $\alpha 7$ nAChR pathways in these processes.

Keywords: Alpha7 nicotinic acetylcholine receptor; Small interfering RNA; Nicotine; HepG2; Apoptosis

Introduction

The smoking habit is a health disparity and a well-known risk factor for different diseases including cancers, but further researches are needed to be conducted on elucidating the mechanisms underlying the effects of cigarette use.¹ Nicotine as a well-documented component of cigarette smoking is an addictive substance of tobacco and poses several health hazards.² This compound exerts its consequences by binding to nicotinic acetylcholine receptors (nAChRs) that are ligand-activated ion channels.³ Among different subtypes of nAChRs, alpha7-subtype of nAChR ($\alpha 7$ nAChR) appears to be of special significance in cellular functions. This receptor functionally expressed by a variety of human normal and cancer cell and tissues such as liver cancer.⁴ It modulates numerous cancer-related properties in most of the cancers and additionally plays a crucial role in the regulation of inflammation through the cholinergic anti-inflammatory pathway in various pathological conditions.^{5,6} Such diverse impacts that initiate from this receptor may have significant influences in determining the consequences of different cancers.

The list of malignancies that are related to nicotine exposure is increasing and involves many different types of cancers.⁷ Among them, hepatocellular carcinoma (HCC), as a common type of liver cancer, is one of the most dangerous and difficult-to-cure malignancies in the world.⁸ Several epidemiologic studies have reported that tobacco smoking is causally associated with HCC development.⁹ The consumed nicotine is broadly metabolized in the liver to produce various types of metabolites.¹⁰ In a recent study, it has been shown that in humans, the greatest amount of $\alpha 7$ nAChR recognized in the liver and this finding demonstrated the importance of $\alpha 7$ receptor-related functions in this organ.¹¹ Nicotine as a double-edged sword exerts diverse physiological and pathological effects in the liver and it may modify the HCC initiation and progression. However, our knowledge about the role of nicotine and $\alpha 7$ nAChR in the development of HCC is very limited and the role and mechanisms by which nicotine influences this cancer remain unclear. There will be opportunities for the identification of novel treatments and prevention options in the future, if follow-up research on the impacts of nicotine through $\alpha 7$ nAChRs continues. In this study, we determined the effects of treatment with nicotine on human hepatocellular carcinoma (HepG2) cell line and aimed to investigate the involvement of $\alpha 7$ nAChR in its effects.

Materials and Methods

Main material and reagents

Human hepatocellular carcinoma cell line (HepG2) was obtained from National Cell Bank of Iran (Pasteur Institute of Iran, Tehran, Iran). Cell culture substances, Roswell Park Memorial

Institute (RPMI) 1640 medium, fetal bovine serum (FBS), trypsin/EDTA and penicillin/streptomycin mixtures were purchased from Gibco Co. (Gibco, Carlsbad, CA, USA). The rest of the materials were bought from Santa Cruz Biotechnology (Santa Cruz, CA, USA), unless otherwise specified in the text. The cells were sustained in RPMI medium with 10% FBS and routinely cultured at 37°C with 5% CO₂ and were used in the logarithmic phase of growth in all tests according to our previous study.¹² Each experiment was repeated three times.

MTT assay

HepG2 cells were cultured at a density of 15×10^3 cells per well in 96-well culture plates. Nicotine (SC-203161) in various concentrations (10^{-8} M to 10^{-2} M) was added to the culture and incubated for 72 h. To determine the concentration-dependent cytotoxicity of nicotine, the MTT (3-(4,5-Dimethylthiazol-2-yl)-2,5-Diphenyltetrazolium Bromide) assay was used as previously described.¹³ Briefly, the MTT at a concentration of 2mg/ml was added to the wells and after that incubated for 4 h at 37°C. After removal of the media, 200µl dimethyl sulfoxide (DMSO) was added to the wells. The values of optical density (OD) of the cells were evaluated at 570 nm with an ELISA Reader (Sunrise RC, Tecan, Switzerland). The results were shown as percentages of the control groups.

siRNA transfection

The siRNA targeting human $\alpha 7$ nAChR (sense: 5'- CCAGACAUCUCCUCUAUA- 3') and the negative control siRNA were purchased from Microcynth (AG, Switzerland). Cells were transfected with these siRNAs following the manufacturer's guidelines. In brief, 2×10^5 cells were seeded at 6-well plates in RPMI-10% FBS medium. The siRNAs at a final concentration of 100 nM in all experiments were transfected into the cells by using nanoparticles according to our previous studies.¹⁴ The siRNAs and nanoparticles were diluted in dilution buffer and incubated for 20 min at room temperature. Afterward, the siRNAs were added to the wells with Opti-MEM solution. The plates were then incubated for additional 6 h at 37°C in a CO₂ incubator. Following that, RPMI-20% FBS medium was added to the wells containing transfected cells. After 48 h of incubation, the suppression of $\alpha 7$ nAChR gene expression was determined by quantitative real-time PCR (qRT-PCR) method.

RNA isolation, cDNA synthesis, and qRT-PCR

The gene expression of $\alpha 7$ nAChR and other apoptosis and cell cycle-related genes were analyzed by qRT-PCR. Briefly, 2×10^5 cells were seeded into 6-well plates one day before the start of the nicotine and siRNA treatments. Total RNA was isolated from the cells using TRIzol (Riboex, Gene All Biotechnology, Seoul, Korea). The total RNA purity and integrity was confirmed by using a NanoDrop (Thermo Scientific, USA). Then 1µg total RNA was reverse transcribed into cDNA (Biofact, South Korea). The qRT-PCR was performed using light cycler 96 (Roche Diagnostics, Mannheim, Germany) and reported by the $2^{-\Delta\Delta CT}$ method. Glyceraldehyde 3- phosphate dehydrogenase (GAPDH) was used as an internal control. The primers sequences for $\alpha 7$ nAChR and other genes were obtained from Sinaclon (Tehran, Iran) and listed in table 1.

Apoptosis assays

Annexin/ PI assay

The apoptosis of HepG2 cells was assessed by flow cytometry (FCM) assay using propidium iodide (PI) fluorescence staining. To estimate the percentage of apoptosis of the cells, they were seeded in the 6-well plates at a density of 2×10^5 cells per well. After 72 h of the treatment with nicotine and 48 h of the siRNA transfection, the cells were stained with an Annexin V-FITC/PI staining assay kit (EXBIO, Vestec, Czech Republic). By using flow cytometry instrument (MACS Quant 10; Miltenyi Biotech, GmbH, Germany) the rate of apoptotic cells

was measured and obtained data were analyzed using the package of FlowJo software (Treestar, Inc., San Carlos, CA).

DAPI staining assay

To evaluate the effect of chronic nicotine exposure and gene silencing of $\alpha 7$ nAChR on chromatin fragmentation, DAPI (4',6- diamidino- 2- phenylindole) staining was performed. For this aim, approximately 15×10^3 of the cells were seeded into 96-well plates. After that, the cells were silenced alone or in a combination with pretreatment with nicotine for 24 h. After the fixation of the cells with 5% paraformaldehyde for 4 h, In the next step, the cells were incubated with Triton X-100 (0.1%) for 5 min and then were stained with DAPI (0.1%) for an additional 10 min. Ultimately, the cells were observed by using an imaging fluorescence microscope system (Cytation 5, Biotek, USA).

Cell cycle analysis

For determining the cell cycle arrest properties, HepG2 cells were seeded at 6-well plate and then treated with nicotine for 24 h and $\alpha 7$ -siRNA for 48 h, respectively. The cells were collected by centrifugation and were incubated with PI using flow cytometric kits (EXBIO, Vestec, Czech Republic) according to the manufacturer's recommendations.¹⁵ In brief, the cell plates were dissolved in a mixture of PBS and RNase A solution and incubated for 30 min. Then 1 mL of the Tris buffer solution was blended with 100 mL of PI solution and added to each well. Ultimately, after 10 min incubation time the cell cycle analysis was carried out by a flow cytometry system (MACS Quant 10; Miltenyi Biotec, GmbH, Germany).

Statistical analyses

All data are shown as the mean \pm SEM. Statistical significance of differences between variables with normal distribution was assessed via one-way ANOVA followed by Tukey post hoc test by using GraphPad Prism 6 software (San Diego, CA, USA). Two-way ANOVA was used when the effect of the two variables was assessed. The P values smaller than 0.05 were considered statistically significant.

Results

Analysis of cell viability

The MTT results indicate that 72 h nicotine treatment significantly reduced the viability of HepG2 cells in a concentration-dependent fashion ($\log IC_{50} = -5.12 \pm 0.15$) (Figure 1A). Increased toxicity was observed at a higher concentration of nicotine. Based on this result, we selected IC_{50} doses of nicotine (10 μM) for subsequent experiments. In this concentration of nicotine, the cells underwent a significant decrease in cellular density (Figure 1B).

Nicotine upregulates $\alpha 7$ nAChR expression, an effect that blocked by $\alpha 7$ -siRNA transfection

The HepG2 cells were examined for $\alpha 7$ nAChR gene expression using RT-PCR. We achieved that after 72 h treatment with nicotine (10 μM) the mRNA level of $\alpha 7$ nAChR was upregulated as compared with non-treated cells (Figure 2). The efficiency of siRNA in down-regulation of $\alpha 7$ nAChR expression was examined in combined nicotine and $\alpha 7$ -siRNA treatment groups. The results indicated that the increased effect of nicotine on $\alpha 7$ nAChR expression was blocked by treatment with $\alpha 7$ -siRNA transfection (Figure 2). Negative control siRNA has no significant effect on $\alpha 7$ nAChR mRNA expression. The results were normalized with the GAPDH housekeeping gene mRNA level.

Effect of $\alpha 7$ -siRNA on apoptosis of HepG2 cells

The effects of nicotine and $\alpha 7$ -siRNA on apoptosis of HepG2 cells were determined by flow cytometry (Annexin V and PI staining) assay. By using this technique and fluorescence-activated cell sorting (FACS) analysis, the portion of apoptotic cells was analyzed and quantified in the cells that incubated with nicotine or $\alpha 7$ -siRNA or combination of them (Figure 3A). In this technique, viable cells are both annexin and PI negative (annexin V-/PI-), while

early apoptotic cells are annexin V+/PI-. Also, the cells in late apoptosis are both Annexin V and PI positive and necrotic cells stain with PI only (V-/PI+).¹⁶ The results indicated that nicotine strongly promoted both early and late apoptosis of the cells but $\alpha 7$ -siRNA inhibited these changes (Figure 3B). In addition to this, DAPI staining verified that the chromatin fragmentation in the processes of apoptosis is enhanced in nicotine-treated cells compared with non-treated control cells. Similar to flow cytometry results, these effects blocked in $\alpha 7$ -siRNA transfected cells (Figure 3C).

Effect of $\alpha 7$ -siRNA on cell cycle progression of HepG2 cells

We investigated whether nicotine influences the arrest of the cell cycle in HepG2 cells. For this purpose, flow cytometry analysis was performed to differentiate the diverse phases of the cell cycle (Figure 4A). The results showed that nicotine inhibited cell cycle progression by especially inducing G2/M phase arrest. This increased response in cell cycle arrest was blocked when the cells were incubated with $\alpha 7$ -siRNA, which indicated the pivotal role of the expression of this receptor in nicotine-induced cell cycle arrest of HepG2 cells (Figure 4B).

Expression analysis of caspase-3 and cyclin B1 mRNA levels

Assessment of apoptosis-related gene revealed that nicotine caused the greatest increase in the expression of caspase-3 in the cells (Figure 5A). In addition to this, cyclin B1 expression as a cell cycle progression-related gene was examined. This gene is a regulatory gene and playing the most pivotal role in the cell cycle progression and transition of the G2/M phase.¹⁷ The results indicated that incubation of nicotine induced down-regulation of cyclin B1 expression (Figure 5B). These observed effects of nicotine on caspase-3 and cyclin B1 mRNA levels were abolished by transfection with $\alpha 7$ -siRNA, which provided other evidence about the involvement of $\alpha 7$ nAChR in the control of various pathways inducing the mitochondrial type of apoptosis.

Discussion

Nicotine, which is an important component of cigarette smoking, may be responsible for various cancers initiation, progression, and therapy responses.¹⁸ This compound has been shown to influence an extensive variety of biological functions ranging from gene expression, oxidative stress, DNA damage, apoptosis, proliferation, angiogenesis, and regulation of hormone secretion.^{18,19} There is controversy in the literature about nicotine-induced toxicity in exposed subjects.²⁰ It is well established that nicotine treatment results in oxidative stress that is associated with multiple alterations of cell structure and function.²¹ Also, accumulating evidence indicates that exposure to nicotine can result in the activation of intracellular signaling pathways that are known to induce apoptosis.^{22–24} In this context, to defend against the harmful effects of this substance, the protective effects of many natural and pharmacologic agents on nicotine-induced toxicity have been investigated in numerous studies.^{25,26} Kim and colleagues revealed that $\alpha 7$ nAChR contributes to the proapoptotic effects of nicotine in periodontal cells.²⁷ In this study, the proapoptotic effects of nicotine were abolished by the pretreatment of α -bungarotoxin, a selective antagonist of $\alpha 7$ nAChR, which highlighted the key role of $\alpha 7$ nAChR in the modulation of nicotine-induced apoptosis.²⁷ Conversely to these findings, in some other studies nicotine induced the antiapoptotic and proliferative effects in different cell types.^{28–30} Therefore, the effect of nicotine on apoptosis may be complicated in various parts of the body. For instance, Jalili and colleagues have shown that in the heart and lung, nicotine caused a significant decrease in caspase-3 mRNA level compared to the control group.³¹ However, in the kidney and liver, the results were significant increases in caspase-3 mRNA level which indicates the different responses of organs to the nicotine exposure.³¹ It is well recognized that apoptosis plays important roles in an extensive variety of physiologic processes and is a major

feature of the biology of malignant diseases. It should be mentioned that, in the liver tissue, enhanced hepatocyte apoptosis and death could result in induced compensatory proliferation.³² In most cases, this event eventually leads to the appearance of HCC.³³ Furthermore, it is known that the growth and proliferation of the progenitor or stem cells are stimulated via signals that are released from apoptotic cells.³⁴ Li and colleagues have shown in an animal model that wound healing and liver regeneration after partial hepatectomy are dependent on Caspase-3 because targeted inactivation of this caspase inhibits these regenerative processes.³⁵ By considering these facts, we assumed that nicotine by dysregulation of the apoptotic processes may involve in the etiology of liver cancer.

The epidemiological positive correlation between cigarette smoking and liver cancer has been demonstrated in previous studies.⁹ Among different subtypes of nicotinic receptors, $\alpha 7$ nAChR is one of the highly expressed receptors in the liver.⁴ In the present study, the effect of nicotine on the viability of cultures of HepG2 cells was analyzed and we provide functional evidence that nicotine induced proapoptotic gene caspase-3 expression and increased apoptosis through the $\alpha 7$ nAChR in HepG2 cells (Figure 6). It has been shown that nicotinic receptors activation can induce hepatocyte proliferation, but only in the presence of hepatic nonparenchymal cells.³⁶ Therefore, the role of other liver resident cells such as Kupffer cells and hepatic stellate cells in the diverse functions of $\alpha 7$ nAChR in this organ should be carefully considered when investigating the toxic or proliferative effects of nicotine in the liver. In addition to apoptosis, it is clear that cell cycle arrest mediates apoptosis in a variety of diseases. Cell cycle regulation ensures the correctness of DNA replication and division. These checkpoints permit progression of the cell cycle or arrest of it in response to DNA damage to provide an opportunity for DNA repair. We also observed that nicotine induces cell cycle arrest in association with increased activation of $\alpha 7$ nAChR pathways (Figure 6). Although this study suggests that $\alpha 7$ nAChR is probably a target for the effect of nicotine on liver cancer progression, it is in controversy with our working hypothesis and provides evidence against a direct role of nicotine in the development of liver cancer. In this study, we could not obtain more detailed mechanisms regarding other cytotoxic responses of HepG2 cells to nicotine exposure, thus further studies are needed to determine the exact intracellular signaling pathways downstream of $\alpha 7$ nAChR in the pathogenesis of liver cancer.

Conclusion

Altogether, it can be concluded that nicotine could lead to disturbance of the crucial equilibrium in cell death and proliferation, which may result in compensatory proliferation in the liver and eventually dysregulated increase of the cells. The findings suggest caution in the use of nicotine, as they could have a potentially detrimental effect on patients at high-risk for liver tumor development.

Acknowledgments

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Ethical Issues

The study was approved by the ethical committee of National Institute for Medical Research Development, Iran (ethical code: 1397.494).

Conflict of Interest

The authors have no conflicts of interest to declare.

References

1. Li LF, Chan RLY, Lu L, et al. Cigarette smoking and gastrointestinal diseases: The causal relationship and underlying molecular mechanisms (review). *Int J Mol Med*. 2014;34(2):372-380. doi:10.3892/ijmm.2014.1786
2. Chaturvedi P, Mishra A, Datta S, Sinukumar S, Joshi P, Garg A. Harmful effects of nicotine. *Indian J Med Paediatr Oncol*. 2015;36(1):24. doi:10.4103/0971-5851.151771
3. Hurst R, Rollema H, Bertrand D. Nicotinic acetylcholine receptors: From basic science to therapeutics. *Pharmacol Ther*. 2013;137(1):22-54. doi:10.1016/j.pharmthera.2012.08.012
4. Zhang L, Wu LL, Huan HB, et al. Sympathetic and parasympathetic innervation in hepatocellular carcinoma. *Neoplasma*. 2017;64(06):840-846. doi:10.4149/neo_2017_605
5. Schuller HM. Regulatory role of the $\alpha 7$ nAChR in cancer. *Curr Drug Targets*. 2012;13(5):680-687.
6. Wang H, Yu M, Ochani M, et al. Nicotinic acetylcholine receptor $\alpha 7$ subunit is an essential regulator of inflammation. *Nature*. 2003;421(6921):384-388. doi:10.1038/nature01339
7. Grando SA. Connections of nicotine to cancer. *Nat Rev Cancer*. 2014;14(6):419-429. doi:10.1038/nrc3725
8. Hajiasgharzadeh K, Somi MH, Shانهbandi D, Mokhtarzadeh A, Baradaran B. Small interfering RNA-mediated gene suppression as a therapeutic intervention in hepatocellular carcinoma. *J Cell Physiol*. 2019;234(4):3263-3276. doi:10.1002/jcp.27015
9. Abdel-Rahman O, Helbling D, Schöb O, et al. Cigarette smoking as a risk factor for the development of and mortality from hepatocellular carcinoma: An updated systematic review of 81 epidemiological studies. *J Evid Based Med*. 2017;(March):1-10. doi:10.1111/jebm.12270
10. Yildiz D. Nicotine, its metabolism and an overview of its biological effects. *Toxicon*. 2004;43(6):619-632. doi:10.1016/j.toxicon.2004.01.017
11. Sakata M, Wu J, Toyohara J, et al. Biodistribution and radiation dosimetry of the (α)7 nicotinic acetylcholine receptor ligand [^{11}C]CHIBA-1001 in humans. *Nucl Med Biol*. 2011;38(3):443-448. doi:10.1016/j.nucmedbio.2010.09.007
12. Yousefi B, Darabi M, Baradaran B, et al. Inhibition of MEK/ERK1/2 signaling affects the fatty acid composition of HepG2 human hepatic cell line. *BioImpacts*. 2012;2(3):145-150. doi:10.5681/bi.2012.019
13. Mohammadzadeh R, Baradaran B, Valizadeh H, Yousefi B, Zakeri-Milani P. Reduced ABCB1 Expression and Activity in the Presence of Acrylic Copolymers. *Adv Pharm Bull*. 2014;4(3):219-224. doi:10.5681/apb.2014.032
14. Aletaha M, Mansoori B, Mohammadi A, Fazeli M, Baradaran B. Therapeutic effects of bach1 siRNA on human breast adenocarcinoma cell line. *Biomed Pharmacother*. 2017;88:34-42. doi:10.1016/j.biopha.2017.01.030
15. Mohammadi A, Mansoori B, Aghapour M, Shirjang S, Nami S, Baradaran B. The *Urtica dioica* extract enhances sensitivity of paclitaxel drug to MDA-MB-468 breast cancer cells. *Biomed Pharmacother*. 2016;83:835-842. doi:10.1016/j.biopha.2016.07.056

16. Wlodkowic D, Skommer J, Darzynkiewicz Z. Flow Cytometry-Based Apoptosis Detection. In: *Calcium Signaling, Second Edition*. Vol 44. ; 2009:19-32. doi:10.1007/978-1-60327-017-5_2
17. Eyvani H, Moghaddaskho F, Kabuli M, et al. Arsenic trioxide induces cell cycle arrest and alters DNA methylation patterns of cell cycle regulatory genes in colorectal cancer cells. *Life Sci*. 2016;167:67-77. doi:10.1016/j.lfs.2016.10.020
18. Zeidler R, Albermann K, Lang S. Nicotine and apoptosis. *Apoptosis*. 2007;12(11):1927-1943. doi:10.1007/s10495-007-0102-8
19. Lee J, Cooke JP. Nicotine and pathological angiogenesis. *Life Sci*. 2012;91(21-22):1058-1064. doi:10.1016/j.lfs.2012.06.032
20. Berkowitz L, Schultz BM, Salazar GA, et al. Impact of cigarette smoking on the gastrointestinal tract inflammation: Opposing effects in Crohn's disease and ulcerative colitis. *Front Immunol*. 2018;9(JAN):1-10. doi:10.3389/fimmu.2018.00074
21. Lan X, Lederman R, Eng JM, et al. Nicotine induces podocyte apoptosis through increasing oxidative stress. *PLoS One*. 2016;11(12):1-17. doi:10.1371/journal.pone.0167071
22. Kim CS, Choi JS, Joo SY, et al. Nicotine-induced apoptosis in human renal proximal tubular epithelial cells. *PLoS One*. 2016;11(3):1-17. doi:10.1371/journal.pone.0152591
23. Marinucci L, Balloni S, Fettucciari K, Bodo M, Talesa VN, Antognelli C. Nicotine induces apoptosis in human osteoblasts via a novel mechanism driven by H₂O₂ and entailing Glyoxalase 1-dependent MG-H1 accumulation leading to TG2-mediated NF- κ B desensitization: Implication for smokers-related osteoporosis. *Free Radic Biol Med*. 2018;117(May 2017):6-17. doi:10.1016/j.freeradbiomed.2018.01.017
24. Huang C, Guo X, Zhao H, et al. Nicotine induces H9C2 cell apoptosis via Akt protein degradation. *Mol Med Rep*. 2017;16(5):6269-6275. doi:10.3892/mmr.2017.7331
25. Kim H-J, Park K-K, Chung W-Y, Lee SK, Kim K-R. Protective Effect of White-fleshed Peach (*Prunus persica* (L.) Batsch) on Chronic Nicotine-induced Toxicity. *J Cancer Prev*. 2017;22(1):22-32. doi:10.15430/jcp.2017.22.1.22
26. Yarahmadi A, Zal F, Bolouki A. Protective effects of quercetin on nicotine induced oxidative stress in 'HepG2 cells.' *Toxicol Mech Methods*. 2017;27(8):609-614. doi:10.1080/15376516.2017.1344338
27. Kim SY, Kang KL, Lee JC, Heo JS. Nicotinic acetylcholine receptor α 7 and β 4 subunits contribute to nicotine-induced apoptosis in periodontal ligament stem cells. *Mol Cells*. 2012;33(4):343-350. doi:10.1007/s10059-012-2172-x
28. Lee H-J, Guo H-Y, Lee S-K, et al. Effects of nicotine on proliferation, cell cycle, and differentiation in immortalized and malignant oral keratinocytes. *J Oral Pathol Med*. 2005;34(7):436-443. doi:10.1111/j.1600-0714.2005.00342.x
29. Marrero MB, Bencherif M. Convergence of alpha 7 nicotinic acetylcholine receptor-activated pathways for anti-apoptosis and anti-inflammation: Central role for JAK2 activation of STAT3 and NF- κ B. *Brain Res*. 2009;1256:1-7. doi:10.1016/j.brainres.2008.11.053
30. Wang C, Niu W, Chen H, et al. Nicotine suppresses apoptosis by regulating α 7nAChR/Prx1 axis in oral precancerous lesions. *Oncotarget*. 2017;8(43):75065-75075. doi:10.18632/oncotarget.20506
31. Jalili C, Salahshoor MR, Moradi MT, Ahookhash M, Taghadosi M, Sohrabi M. Expression Changes of Apoptotic Genes in Tissues from Mice Exposed to Nicotine. *Asian Pac J Cancer Prev*. 2017;18(1):239-244. doi:10.22034/APJCP.2017.18.1.239
32. Maeda S, Kamata H, Luo JL, Leffert H, Karin M. IKK β couples hepatocyte death to

- cytokine-driven compensatory proliferation that promotes chemical hepatocarcinogenesis. *Cell*. 2005;121(7):977-990. doi:10.1016/j.cell.2005.04.014
33. Zhou Z, Park S, Kim JW, et al. Detrimental effects of nicotine on thioacetamide-induced liver injury in mice. *Toxicol Mech Methods*. 2017;27(7):501-510. doi:10.1080/15376516.2017.1323256
 34. Ryoo HD, Bergmann A. The role of apoptosis-induced proliferation for regeneration and cancer. *Cold Spring Harb Perspect Biol*. 2012;4(8):1-18. doi:10.1101/cshperspect.a008797
 35. Li F, Huang Q, Chen J, et al. Apoptotic Cells Activate the “Phoenix Rising” Pathway to Promote Wound Healing and Tissue Regeneration. *Sci Signal*. 2010;3(110):ra13-ra13. doi:10.1126/scisignal.2000634
 36. Yoshimura R, Somekawa S, Omori H, Endo Y. Carbachol Induces Hepatocyte Proliferation, but Only in the Presence of Hepatic Nonparenchymal Cells. *J Physiol Sci*. 2007;57(3):139-145. doi:10.2170/physiolsci.rp003707

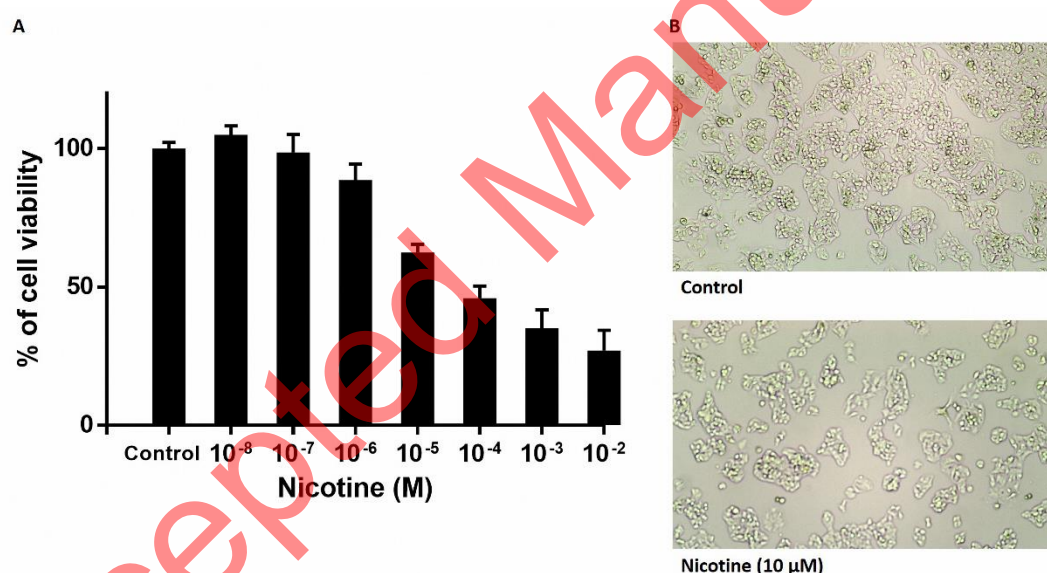


Figure 1. (A) Nicotine leads to the inhibition of the proliferation of HepG2 cells in dose-dependent patterns ($\log IC_{50} = -5.12 \pm 0.15$). Data are shown as Mean \pm SEM and all of the experiments were repeated three times. (B) In IC_{50} concentration of nicotine, the cells underwent a significant decrease in cellular density and increased toxicity was observed at a higher concentration of nicotine.

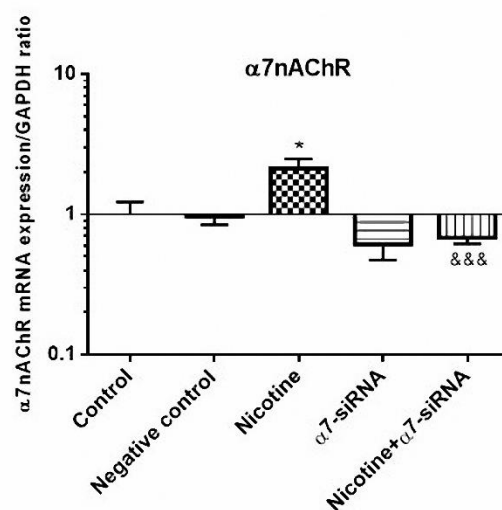


Figure 2. The mRNA level of $\alpha 7$ nAChR was upregulated after treatment with nicotine (10 μ M) as compared with non-treated cells. This effect was blocked by $\alpha 7$ -siRNA transfection. To determine the differences between the statistical significance of the groups, Student's T and ANOVA tests were performed. Negative control siRNA has no significant effect on $\alpha 7$ nAChR mRNA expression. The results were normalized with the GAPDH housekeeping gene mRNA level. * $P < 0.05$ in comparison with the control group. &&& $P < 0.001$ in comparison with nicotine-treated group.

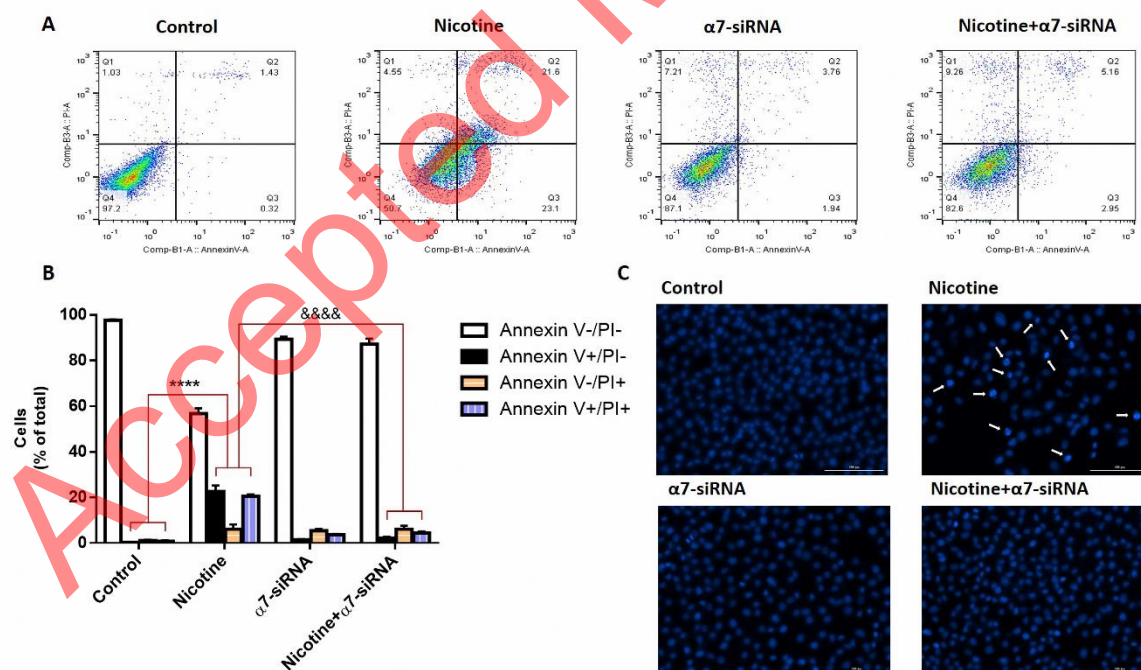


Figure 3. (A) By utilizing the FACS analysis and annexin V/PI assay, the portion of apoptotic cells were separated in HepG2 cells that incubated with nicotine, $\alpha 7$ -siRNA or combination of them. (B) The analysis indicated that nicotine strongly promotes apoptosis of HepG2 cells. **** $P < 0.0001$ in comparison with control group. The siRNA-mediated knockdown of $\alpha 7$ nAChR in these cells inhibits the proapoptotic effects of nicotine. &&&& $P < 0.0001$ in

comparison with nicotine group. (C) DAPI staining also validated the fragmentation of chromatin which occurs in the processes of apoptosis and increased in nicotine-treated cells compared with non-treated cells. This effect blocked in $\alpha 7$ -siRNA transfected cells.

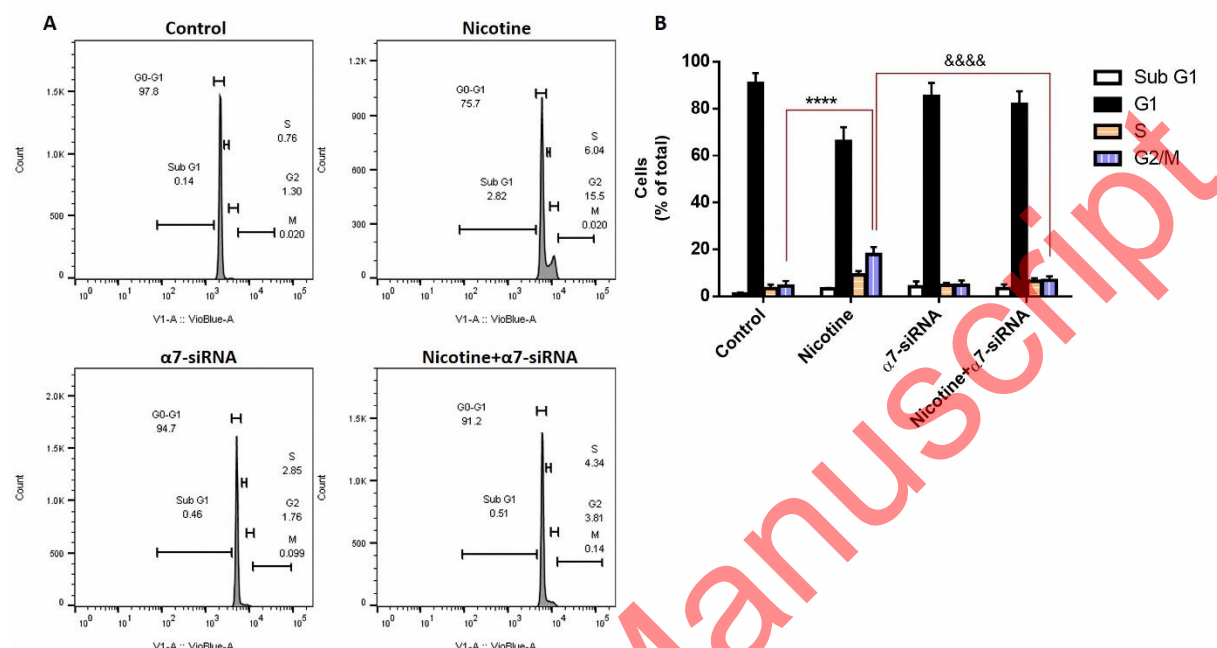


Figure 4. (A) The cell cycle distribution was determined after treatment with nicotine, $\alpha 7$ -siRNA or combination of them. (B) Flow cytometry analysis showed that nicotine inhibited cell cycle progression by inducing G2/M phase arrest. These increases were reduced when the cells incubated with $\alpha 7$ -siRNA. **** $P < 0.0001$ in comparison with control group. &&&& $P < 0.0001$ in comparison with nicotine group.

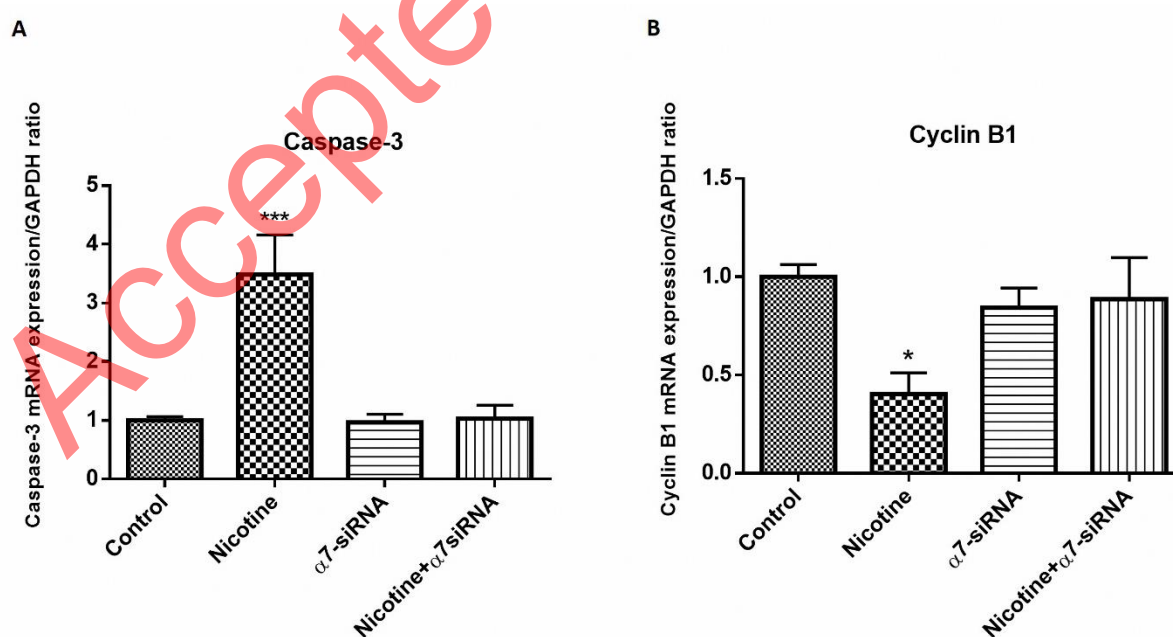


Figure 5. (A) Nicotine caused the greatest increase in the gene expression of caspase-3 in the HepG2 cells. *** $P < 0.001$ in comparison with the control group. (B) Incubation of nicotine

induces down-regulation of cyclin B1 expression. *P <0.05 in comparison with the control group. These observed effects of nicotine on caspase-3 and cyclin B1 mRNA levels were abolished by transfection with $\alpha 7$ -siRNA.

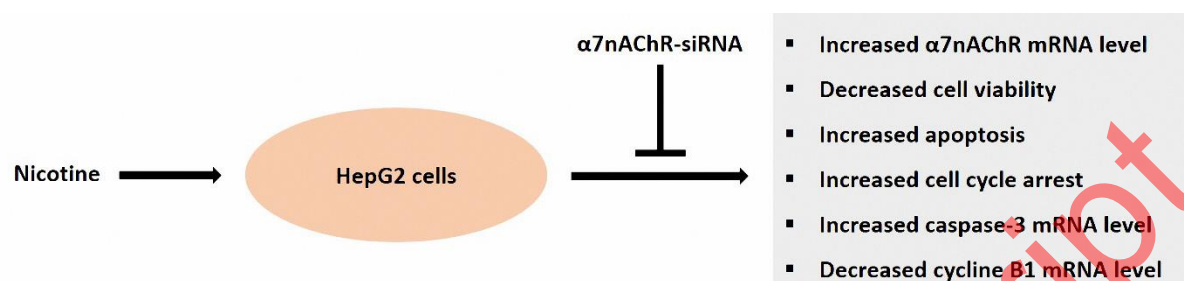


Figure 6. Treatment of the HepG2 cells with the $\alpha 7$ nAChR specific siRNA blocks nicotine-induced proliferative and apoptotic effects.

Table 1. The sequence of primers for alpha7 nicotinic acetylcholine receptor ($\alpha 7$ nAChR), caspase-3, cyclin B1, and GAPDH genes.

Genes		Sequences
$\alpha 7$ nAChR	Forward	5' CGCCACATTCCACACTAACG 3'
	Reverse	5' AGACCAGGACCCAAACTTCAG 3'
Caspase-3	Forward	5' TGTCATCTCGCTCTGGTACG 3'
	Reverse	5' AAATGACCCCTTCATCACCA 3'
Cyclin B1	Forward	5' GGTGG GTCGGCCTCTACCT 3'
	Reverse	5' AGCCAGGTGCTGCATAACTGGAA 3'
GAPDH	Forward	5' CAAGATCATCAGCAATGCCTCC 3'
	Reverse	5' GCCATCACGCCACAGTTTCC 3'