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# **Review Article**

Lipid-based nanoparticles as oral drug delivery systems: overcoming poor gastrointestinal absorption and enhancing bioavailability of peptide/protein-based drugs

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#### **Abstract**

Delivery and formulation of oral therapeutic peptide/protein-based biotechnological drugs have always been a challenge for the pharmaceutical industry. The bioavailability of oral biopharmaceuticals mainly relies on their gastrointestinal solubility and permeability which are affected by their poor membrane penetration, high molecular weight and proteolytic (chemical and enzymatic) degradation resulting in limited delivery and therapeutic efficacy. The present review article highlights the challenges and limitations of oral delivery of therapeutic peptide/protein-based drugs focusing on the application, potential and importance of solid lipid nanoparticles (SLNs) and nanostructure

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lipid carriers (NLCs) as lipid-based drug delivery systems (LBDDSs) and their advantages and drawbacks.

LBDDSs, due to their lipid-based matrix can encapsulate both lipophilic and hydrophilic drugs, and by reducing the first-pass effect and avoiding proteolytic degradation offer improved drug stability, dissolution rate, absorption, bioavailability and controlled drug release. Furthermore, their small size, high surface area and surface modification increase their mucosal adhesion, tissuetargeted distribution, physiological function and half-life.

Properties such as simple preparation, high-scale manufacturing, biodegradability, biocompatibility, prolonged half-life, lower toxicity, lower adverse effects, lipid-based structure, higher drug encapsulation rate and various drug release profile compared to other similar carrier systems makes LBDDSs a promising drug delivery system. Nevertheless, undesired physicochemical features of peptide/protein drug development and discovery such as plasma stability, membrane permeability and circulation half-life remain a serious challenge which should be addressed in future.

**Keywords:** peptide/protein-based drugs, biopharmaceuticals, lipid-based drug delivery systems, Solid lipid nanoparticles, nanostructured lipid carriers, oral drug delivery

#### Introduction

In recent decades, high demands for cost-benefit healthcare expenses, efficient therapeutics (e.g. safety and efficacy) and non-stop generic substitution has urged pharmaceutical companies,<sup>1</sup> industry and market to shift to biotechnological, peptide/protein-based drugs and biopharmaceuticals. These novel categories of drugs, unlike Active Pharmaceutical Ingredients (APIs, drugs), offer better feedback due to higher potency, selectivity and specificity for their extracellular target.<sup>2</sup>

Peptides and proteins, as cell products, have various physiological functions in body such as hormones, enzyme substrates and inhibitors, antibiotics, biological regulators, structural components, signaling factors and catalyzers which all implies their importance in body; hence, any abnormality in their amino acid sequence or structural disfunction leads to sever diseases and pathological conditions; diabetes,<sup>3</sup> dwarfism,<sup>4</sup> cystic fibrosis,<sup>5</sup> thalassemia <sup>6</sup> or impaired blood clotting,<sup>7</sup> among many others.<sup>8</sup> as such, due to their biological specificity and efficient affinity and efficacy, peptides and proteins have been exploited as drugs for treatment of diseases (Figure 1).<sup>10</sup>

Stability in proteins is the result of balance among four destabilizing and stabilizing forces: electrostatic interactions, hydrogen bonding, van der Waals

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forces and hydrophobic interactions, which form the secondary, tertiary and quaternary structures of proteins and any disruption will influence the structural balance and destabilizes that particular protein. Different environmental factors can influence the chemical and physical stability of proteins such as pH, ionic strength, temperature, high pressure, non-aqueous solvents, metal ions, detergents, adsorption, agitation and shearing which all are inevitably part of the manufacturing, sterilization and lyophilization process, and consequently might damage the developing protein resulting in biological inactivation, aggregation, immunogenicity and precipitation. The secondary tertiary and precipitation.

Peptide/protein-based drugs are highly dependent on the production process, yet are biocompatible, cost-benefit, with modifiable in-vivo bioactivity, specific targeting, chemical diversity, and easily synthesized by using solid-phase peptide synthesis methodologies (e.g. Merrifield's method) in which the amino acid sequence can be precisely chosen and inserted at the molecular level by modifying the basic units. 16 nevertheless, undesired physicochemical features of protein drug development and discovery, remains a serious challenge for formulation scientists as well as pharmaceutical and biotechnology companies. Only recently peptides have been considered as therapeutic agents while they were never considered as a potential therapeutic agent;<sup>17</sup> mostly due to their protease degradation, metabolic instability, short half-life, manufacturing complications and high expenses, which in long-term administration renders them unfavorable in terms of patient costs and compliance especially with regard to parenteral administration as the majority of peptides (10%) have a very low oral bioavailability. 18 On the other hand, peptides' biodegradability into low-toxicity metabolites,<sup>2</sup> low drug-drug interactions immunogenicity, higher tissue penetration (owing to their small size), higher invivo activity (per unit mass), stability and lower expenses favors them over large therapeutic proteins and antibodies for regulatory approval (higher than 20%) which is twice the rate of small molecules.<sup>17</sup>

As well as peptides, proteins also gained importance and their application in pharmaceutical science and industry was emphasized due to more advanced analytical methods resulting in the recognition of various peptides and hormones as therapeutic biopharmaceuticals, novel genetic and molecular engineering methods to produce large-scale proteins and recently-defined roles of proteins as regulatory components of numerous diseases.<sup>19720</sup> Since then, pharmaceutical industry has developed various large-scale oral delivery technologies for peptides/proteins as active ingredients.<sup>21</sup>

Low oral bioavailability of peptide/protein-based drugs renders them being formulated as parenteral preparations; large molecular size,<sup>22</sup> susceptibility to

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enzymatic degradation (local or mostly GI tract), poor stability in the gastric acidic environment,<sup>23</sup> poor intestinal penetration, short plasma half-life, immunogenicity and the propensity to aggregation, adsorption, and denaturation.<sup>24/25</sup> Enzymatic degradation and poor intestinal penetration, among all, have been mainly mentioned for low oral bioavailability and short half-life of protein drugs (<1%) which is claimed to be increased to 30-50% by pharmaceutical enterprises.<sup>26/27</sup> Pharmaceutical therapeutic proteins, due to their large molecular size which leads to low blood absorption and diffusion, requires specific epithelial transporters otherwise they cannot enter the general circulation by the ordinary routes of drugs ansorption. Furthermore, low pH and protease enzymes of GI aggravate this condition even more.<sup>28</sup>

Among different administartion routes, the most common route for pharmaceutical/bitechnologicalproteins is intravenous (I.V.) injections (Table 1), which is not favorable in terms of patient compliance, clearance varies from a few minutes to several days, and might result in undesired deposition and distribution which require repeated injections with higher therapeutic doses to achieve efficacy, <sup>29</sup> subsequently causing severe adverse effects.

The subcutaneous (S.C.) and intramuscular (I.M.) injections are other administration routes (Table 1), the former is the most common one, especially for vaccines. Different factors such as molecular weight, site of injection, local injection site activity and pathological conditions can influence the fate of protein drugs following S.C. injection leading to a bioavailability as high as 100% or much lower.<sup>31</sup> upon injection, proteins with high molecular weight (<16,000 Da) are absorbed either from vessels' endothelial cells to capillaries or reach the local lymphatic system and through thoracic duct join the blood circulation, while proteins with small molecular weight are mainly diffused through the local capillaries.<sup>31</sup> however, protein transportation through lymphatic way is undesired due to slow time of circulation which might result in protein enzymatic degradation.<sup>31/32</sup>

Non-invasive routes have increasingly been investigated as well as an alternative to the conventional invasive injectable routes (Table 1). Numerous studies have investigated nasal, ophthalmic, buccal, rectal, vaginal, transdermal and pulmonary routes for peptide/protein delivery.<sup>28/33-40</sup>

Based on studies, mucosae which so far have been neglected for drug delivery seem to be a promising approach for drug absorption, especially efficient for biomolecules of large size and molecular weight.<sup>28</sup> The advantages of mucosal surfaces (mouth, eye, nose, rectum and vagina) for drug delivery over skin and GI tract can be named as: fewer biological barriers to pass for systemic diffusion, rapid absorption and evading hepatic first-pass effect. However, one practical

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challenge of mucosae is related to the preparations that are formulated for local long-term treatment.

Table 1: various peptides and proteins' administration routes

Method	Delivery routes	Formulation/Device requirement	Reference
	Intravenous (I.V.),	Liquid or reconstituted solid (syringe),	
	subcutaneous (S.C.),	i.v. injected liposomes	[42-46]
	intramuscular (I.M.),		
Invasive	Intracerebral vein (I.C.V.)		
	Depot system (S.C. or I.M.)	Biodegradable polymers, liposomes,	[4 <mark>2</mark> -
		permeable polymers (not degradable)	44,47,48]
		microspheres, implants	
	Pulmonary	Liquid or powder formulations,	[42-45,49]
		nebulizers, metered dose inhalers,	
		dry powder inhalers	
	Oral	Solids, emulsions, microparticles,	[42-
		nanoparticles, with or without	44,49,50]
		absorption enhancers	
Non-invasive	Nasal	Liquid, usually requires permeation	[42-
		enhancers, nanoparticles	44,51,52]
	Transdermal	Iontophoresis, electroporation, chemical	[42-
		permeation enhancers, prodrugs,	44,53,54]
		sonophoresis, transfersomes	
	Buccal, rectal, vaginal,	Gels, suppositories, bioadhesives,	[42-
	ocular	microparticles	44,55,56]

Despite alternate routes of drug delivery, oral delivery (P.O.) is still the most preferred one; non-invasive, painlessness, easy self-administration, low risk of cross-infection, high patient convenience/compliance, outpatient feasibility,<sup>57</sup> cost-benefit (no need for sterile manufacturing)<sup>57</sup>. Oral route does not offer the drawbacks of I.V. route; drug extravasation from blood, catheter-related infectious complications, thrombosis and being expensive and invasive, especially for chronic conditions.

However, biological barriers of GI tract with their associated enzymatic and chemical processes hamper the efficiency of oral route for drug delivery. Furthermore, the epithelial cell monolayer membrane of the GI tract even more aggravate the condition of low permeability for many protein drugs with low gastrointestinal solubility which finally results in low bioavailability.<sup>58</sup>

Some active moieties cannot be delivered through oral route. According to the Biopharmaceutic Classification System (BCS), oral bioavailability of each drug is determined by its solubility along the GI tract and cellular penetration. Most of the potential drug candidates developed with high-throughput screening methods generally have higher molecular weights and tend to be

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lipophilic in nature.<sup>62</sup> Other factors contributing to low oral bioavailability of drugs are low stability in the gastrointestinal environment and poor membrane permeability. Most of drugs are substrates to intestinal efflux transporters like p-glycoprotein resulting in poor oral bioavailability.<sup>63</sup>

Regardless of the administration route, majority of peptide/protein-based biopharmaceuticals, lacking necessary physicochemical requirements, fail to diffuse and be absorbed to their target tissue which implies the importance of drug delivery and tissue-targeting systems for achieving as maximum therapeutic effects as possible. The carrier systems diffuse and distribute the intended therapeutic molecules to their targeted site with as maximum concentration as possible in the affected area and as minimum concentration as possible in the intact tissues to lower the general adverse effects.<sup>64</sup>

#### Peptide/protein drug delivery

Introduction of novel biotechnological molecules as potential therapeutics, advent of chemical synthesis methods and recombinant DNA technology have all rendered protein synthesis and delivery an important area of research which resulted in the production of numerous large-scales drugs of peptide/protein origin such as monoclonal antibodies, hormones and vaccines. According to The 2018 and 2019 PhRMA reports,<sup>65</sup> there have been respectively 4751 and 5422 novel biotechnological medicines in research and development (R&D) phase for more than 100 diseases such as cancer, infectious diseases, autoimmune diseases, AIDS/HIV, antiparasitic and related conditions (Figure 1), which have been either in human clinical trials or under review by the Food and Drug Administration (FDA).<sup>65</sup> nevertheless, abovementioned challenges for their delivery through GI tract <sup>66-70</sup> and the blood brain barrier (BBB)(in the case of central nervous system diseases),<sup>71/72</sup> makes their therapeutic potential and clinical application questionable.

In the last decades, numerous drugs of peptide/protein origin have been in preclinical studies and clinical trials,<sup>73</sup> more than 400 recombinant peptides and proteins and 1300 under clinical trials.<sup>74</sup> The reason could be attributed mostly to the larger size of peptides and proteins comparing to conventional drugs, which provides drug-target interaction with binding pockets that are not normally available to small molecular drugs. These targets could be part of intracellular protein-protein interaction network which have been recognized in numerous diseases. The peptide/protein-based drugs in order to interact with such targets must penetrate cells, however, most of them are known to have extracellular targets, <sup>73</sup> and are parenterally administered so cellular penetration is not their ordinary route as it is for mucosal surfaces. Currently, the main obstacle of the oral administration of these novel categories of drugs for their

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maximum therapeutic effects could be addressed as the penetration through intestinal cellular membranes and target cellular membranes.

#### Transport mechanisms in the GI tract

To formulate and synthesize drug delivery systems for oral peptide/protein-based drugs, and biopharmaceuticals a throughput understanding of the biological pathways involved for their absorption and diffusion in the GI tract is necessary and worthwhile. Various physicochemical features govern the pathway through which the molecules will be penetrating the intestinal cells; molecular weight, hydrophobicity/hydrophilicity, ionization constants, and pH stability, among all.

Paracellular transport. It has the following features as, space dimension of 10 Å, aqueous pores (epithelial tight junctions) 7–9, 3–4 and 8–9 Å for the jejunum, the ileum and the colon respectively,<sup>75</sup> to allow the passage of solutes with a specific molecular radius and tight junctions building 0.01% of the total absorption surface area of the intestine.<sup>76</sup> These data prove the restriction of the paracellular transport toward the passing molecules (Table 2). however, there is an electrical resistance diversity and consequently ionic selectivity. In the latter case, also transcellular pathway's collaboration adjusts rate and selectivity of export of ions and solutes and overall tissue-specific transport. The tight junction along with ion channels are involved in size and charge selectivity, ion concentration-dependent penetration, competitive-based penetration among different molecules, unordinary mole-fraction effects and pH-sensitivity.<sup>77</sup> hydrogen bonding capacity and lipophilicity do not influence much the paracellular pathway.

Transcellular transport. It is an endocytic process at apical membrane and the absorbed molecules are released at the basolateral membrane, glucose is also transported with this mechanism (Table 2). The protein-lipid ratio is very insignificant in the basolateral membrane due to its thinner and more permeable structure than the apical membrane. This transport mechanism is governed by various factors: different physicochemical properties of molecules, size, lipophilicity/hydrophobicity, hydrogen bond formation, surface charge, superficial ligands; the physiological condition of the GI system and the animal models studied for transport mechanism. There are mainly two types of primary intestinal epithelial cells for molecules transportation; Enterocytes and M cells, the former lining about 99% of the GI tract and the latter mainly the area of Peyer's patches and the human follicle-associated epithelium (FAE)(antigen-specialized). M cells function as presenting and transporting peptides and proteins to the local lymphoid tissues for immune reactions and a vulnerable and available way for pathogenic organisms. Due to their great

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endocytosis and transcytosis capacity for transporting diverse molecules and biomaterials (e.g. nanoparticles),82'83 M cells could be used for oral delivery of peptides and proteins and finally through phagocytosis, adsorptive (through clathrin-coated pits and vesicles) and fluid phase endocytosis they adsorb microorganisms.84 and Some studies nanoparticles transportation through intestinal villi and contradicting the recent debates over the rate of particle absorption.85'86 There is a consensus on the transportation of the majority of particles in FAE,85'87'88 for which there have been studies on the Peyer's patches and M cells involvement on various biomaterials absorbency. The transcellular mechanism however is not a desirable route for low molecular-weight lipophilic drugs. Overall, absorption by this mechanism is reduced in a great extent in the colon part of the large intestines in comparison to the paracellular mechanisms.89

Carrier-mediated transport. It is an active and energy-dependent transportation of specific molecules against their concentration gradient through specific membrane receptors, such as  $\beta$ -lactam antibiotics and angiotensin converting enzyme (ACE) inhibitors, monosaccharides and amino acids (Table 2). In one study using Caco-2 cell monolayers, it was proved that the conjugated insulin is transported 5 to 15 times more through the transferrin receptor than then insulin receptor itself. 90

Receptor-mediated transport. It has been investigated to evaluate the oral bioavailability of peptide and protein drugs by modifying receptor specific ligands-drug interaction. This mechanism has functions in different processes such as endocytosis (clathrin-mediated), phagocytosis, pinocytosis and potocytosis (nonclathrin-mediated)(Table 2). The absorption starts with the binding of molecules to their specific receptors and their internalization into endosomes with low acidic pH which might dissociate receptor-ligand bound and accordingly degrades endosomes. The absorbed peptide and protein access into systemic blood circulation with two distinct pathways: hepatic portal vein and intestinal lymphatic vessels, the amount of peptides and proteins absorbed through either of these two pathways depends greatly on the physicochemical features of the formulation, portal vein is the main pathway for the majority of orally administered peptide and protein drugs and through which hydrophilic molecules are absorbed and transported to the blood systemic circulation first through the hepatic portal vein and then by the hepatic artery, and finally they are delivered to their sites of action. But lipophilic molecules penetrating through the same intestinal barriers are transported to the intestinal lymphatic vessels, bypassing the first-pass effect, and directly delivered to the vena cava for blood systemic circulation.

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Feature of molecules **Transport mechanisms Process** Note Paracellular Passive diffusion in Ions, large substances Restricted protein delivery and solutes <15 Å due to tight junctions [94] intercellular spaces between epithelial (3.5kDa)[93] cells, tissue-specific transport [91,92] Transcellular Intestinal Various physicochemical Limited transport of relatively properties low molecular-weight transcytosis, Enterocytes and M lipophilic drugs cells [95] Small di-/tripeptides Carrier-mediated Across the cell Utilized by small membrane or entire hydrophilic molecules monosaccharides, and amino cell [96] [97] acids are transported transcellularly [98] Direct delivery of hydrophilic Receptor-mediated Receptor specific The physicochemical and ligand [99], metabolic features ligands to liver, direct delivery endocytosis [100] of lipophilic ligands to the vena cava [101]

Table 2: Transport mechanisms in the GI tract

#### Absorption of oral drugs

Drugs in order to be absorbed through GI tract are required to have high solubility and permeability, however, this is not the case for numerous drugs with low aqueous solubility and consequently low and diverse bioavailability.<sup>102</sup> for such drugs simultaneous presence of high amount of fat through meals can increase their oral bioavailability,<sup>103-105</sup> via prolonging GI tract passage time, exocrine pancreas secretion stimulation, reduced metabolism, lymphatic-associated absorption, increased intestinal penetration, reduced cellular efflux and liver- and mesenteric-related blood alteration.<sup>106/107</sup>

Introduction of lipid-based drug delivery systems in 1990s provided scaffolds which increase dissolution rate of poor aqueous soluble drugs (i.e. hydrophobic drugs) by providing a phase in which the drugs can disintegrate and be absorbed and diffused toward its site of action. 108 after the degradation of lipids in the intestine, active mono- and diglycerides are formed on the surface of lipids which later disassociate and transform into micelles and simultaneously drug is also solubilized inside micelles. These micelle-drug mixtures are finally absorbed. 109-112

The absorbed components through intestinal segment of GI system follow two distinct pathways according to their features: blood vessels and lymphatic vessels. The former is the preferred route for most of the oral drugs by which

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they are absorbed into the systemic circulation via portal vain, and the latter for highly lipophilic drugs (log P >5) by which drugs are absorbed into systemic circulation via lymphatic vessels.

Overall, the presence of lipid increases absorption of numerous drugs more through lymphatic vessels, especially lipophilic ones and macromolecules of high molecular weight, mostly due to their higher permeability to nanoparticles than blood vessels, and most importantly overpassing hepatic first-pass effect. Nevertheless, it has been demonstrated that the absorption through lymphatic pathway is affected by the length of fatty acid chains; long-chain triglycerides (14-18 chains) are more favored for absorption than low-chain ones. 111/117

# Physiological barriers of peptide/protein-based drugs absorption Gastrointestinal barriers

In-depth studies of molecular pharmacology have provided a better understanding of the biological and molecular processes of the GI system, while its target sites has introduced new insights for targeted delivery of oral peptide/protein-based drugs and biopaharmaceuticals. There are two types of proteolytic enzymes with their specific site of action which are responsible partly for the physiological processes of GI system toward peptides and proteins: *endoproteases*, including trypsin, chymotrypsin, and elastase, which hydrolyze the internal bonds of the peptide chain to the amino- and carboxy-terminus, and *exopeptidases*, including carboxypeptidase A and aminopeptidase, which hydrolyze the amino- and carboxy-terminus bonds to the peptide chain (Table 3). Enzymatic degradation happens at the lumen, brush border, the cytosol of the enterocytes, in the lysosomes and other cell organelles. 121

The secretions of stomach (hydrochloric acid (HCl), potassium chloride (KCl) and sodium chloride (NaCl)) provide an acidic pH of 1.5-3.5 for the proteolysis of peptides and proteins by breaking them down into aminoacids, dipeptides and tripeptides for absorption. The digestion of peptides and proteins starts with pepsin in the acidic environment of stomach (pH 2), however, the alkaline environment of the intestines (pH 6) inactivates pepsin. The instantaneous wide change of pH from acidic stomach to alkaline intestines influences the degradation of ingested peptides and proteins and might contribute to their precipitation that redissolve following pH change. 122-124

The small intestine is the major site of absorption along the GI tract due to the higher enzymatic activity of proteases (mostly in duodenum and jejunum). The brush borders of the intestinal epithelial cells secrete numerous specific enzymes (e.g. sucrose) leading to peptide/protein absorption and

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degradation.<sup>120</sup> furthermore, the digestive secretions of the exocrine part of the pancreas also contain endo-/exopeptidases which are released into duodenum to increase pH for the activity of intestinal enzymes (e.g. trypsin). However, in the terminal parts of jejunum and ileum the enzymatic activity of aminopeptidases is decreased to 20–30% where Peyer's patches are located and these areas could be a potential site for peptide/protein drug delivery.<sup>98</sup>/125

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Types	Enzymes	Major site of action	
Gastric proteases	Pepsins (aspartic proteases)	Broad activity, hydrolyzes	
		numerous peptide bonds	
Brush border proteases	Aminopeptidase A	Aminopeptidases are N-	
	Aminopeptidase N	terminopeptidases, degrading	
	Aminooligopeptidase	most <mark>ly 3–10</mark> amino acid	
	Dipeptidylaminopeptidase IV	residue-dipeptides and amino	
	Carboxypeptidase	acids	
Cystosolic proteases	Di- and tripeptidase	2-3 Aminopeptide amino acids	
Intestinal pancreatic	Trypsin (endopeptidase)	Peptide bonds of basic amino	
proteases	α-chymotrypsin (endopeptidase)	acids/peptides Peptide bonds	
	Elastase (endopeptidase)	of hydrophobic amino	
	Carboxypeptidases	acids/peptides Peptide bonds	
	(exopeptidase)	of smaller and nonaromatic	
		amino acids/peptides	
		A: C-terminal amino acid	
		B: C-terminal basic amino acid	
Brush border proteases	Aminopeptidase A	Aminopeptidases are N-	
	Aminopeptidase N	terminopeptidases, degrading	
	Aminooligopeptidase	mostly 3–10 amino acid	
	Dipeptidylaminopeptidase IV	residue-dipeptides and amino	
	Carboxypeptidase	acids	

#### Mucosal barriers

Mucus has a major importance and function by determining the absorption and bioavailability of administered drugs especially through oral route. The mucosal surface of stomach has three components which further hamper the drug diffusion and absorption as an exogenous component: the first one, which is lined by surface epithelial cells and tight junctions, has a role against irritant and unsuitable fluids; the second one has a very unique insoluble protective mucus (the mixture of surface epithelial cells and neck cell) which creates a jelly-like layer throughout the entire surface mucosae of the stomach; and the third one is composed of bicarbonate ions which are secreted by the surface epithelial cells. 126/127

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There are however other components which further disturb peptide/protein absorption through oral administration, such as glycocalyx located on the surface layer of the stomach epithelial cells with an acidic nature and containing sulfated mucopolysaccharides. Goblet cells of the stomach wall secrete mucus, which covers upper layer of glycocalyx, 128 and contains mucin glycoproteins, enzymes, electrolytes and water. 129 the mucin glycoprotein gives glycocalix an adhesive feature and functions more as a physical barrier than a chemical one, 130 · 131 and in the stomach and colon parts of GI tract has a thick layer while in the small intestine part is thinner and this fact can be justified according to the digestive functions of each segment. 132 Overall, the abovementioned components altogether are protected by viscoelastic layers. Peptide and proteins first of all are required to pass the outermost layers, mucus and glycocalayx, to reach the cellular membranes which present a viscous barrier to absorption and diffusion.

# Nanomedicines – novel drug delivery systems

After administrating drugs, their plasma concentration increases and they affect their site of action effectively followed by a gradual decrease to a point when their plasma concentration is not sufficient enough anymore to elicit their intended therapeutic effect. Hence, Drugs are required to be re-administrated based on the dose and frequency of administration to provide the same concentration which must be neither higher nor lower than their therapeutic concentration level; a concept widely known as "therapeutic window"; higher doses will result in general toxic effects and lower ones cannot elicit any therapeutic effects. Their "therapeutic window" and this issue implies the importance of novel drug delivery systems to be introduced.

A novel interdisciplinary branch of biomedical science, "nanomedicine", has been investigating the potential application of biomaterials and nanostructures as drug delivery systems (DDSs) for sustained, controlled and tissue-targeting delivery of drugs and active agents, which due to their pharmaceutical features overcome some of the limitations of conventional drugs.

Studies have proved that "drug discovery" alone does not offer practical solutions to therapeutics as most of the successful in-vitro experiments result in failure in in-vivo experiments mostly due to: poor drug concentration owing to low absorption, quick metabolism and elimination (peptides and proteins); general blood distribution which results in drug-related adverse effects and toxicity (anti-cancer drugs); low drug solubility of aqueous solutions when administered intravenously; unpredictable bioavailability and high plasma-level

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variations with oral administration and physiological processes involving a drug's plasma level (e.g. food on cyclosporine).

In DDSs the in-vivo destiny of drugs is directly influenced by a series of factors which can be modified for in-site delivery with the desired therapeutic concentration. Peptide/protein-based drugs are encapsulated in nanomedicines for transportation through GI tract offering benefits such as high stability for storage and administration, and large-scale sterile manufacturing for oral preparations. Nanomedicines can be formulated and modified with the desired criteria such as size, surface properties and release profile to have tissue-targeted delivery within drug's unique therapeutic window.

Nanomedicines have a size ranging from 1 to 100 nm where the therapeutic molecules can be incorporated in the core, matrix or attached on the surface (in the case of high surface/volume ratio), which the latter results in longer half-life and systemic circulation and increased mean residence time (MRT). Since 1990s lipid-based drug delivery systems (LPDDSs) have been under investigation owing to their advantages; biocompatibility, higher penetration capacity, lipophilicity with no need for surface modification, simple fabrication, cost benefit and industrial-scale production compared to their counterpart DDSs such as polymeric and inorganic nanomedicines. 1377138

Based on the fabrication methods and physicochemical properties lipid-based drug delivery systems (LPDDSs) are classified into the following:

1) Liposomes in the form of spherical vesicles which are composed of one or multiple lipid bilayer (phospholipid or natural phospholipids) enclosing an agueous core. 137/138 Their size varies from 10 to 1000 nm and they are the first generation of LPDDSs that were employed mainly for parenteral route of drug delivery. They possess low antigenicity and toxicity, high drug encapsulation and loading efficiency and sustained and controlled drug release as their benefits. However, their synthesis is complex and has low stability and rapid reticuloendothelial system clearance and scale-up problems. Despite introducing various surface functionalization (antibodies and peptides) and modification (e.g. PEG coating) which enhances their blood half-time, structural stability and therapeutic efficiency, they are still undesirable in terms of production. 139-141 Currently, numerous liposome-based industrial scale formulations are approved for a variety of diseases: Doxorubicin for cancers (Doxil®, Myocet® and Lipodox ®), Amphotericin B for fungal infections (Ambisome®), Cytarabine for lymphomatous meningitis (Depocyt®), Morphine sulfate for pain management (DepoDur®), inactivated hepatitis A virus (strain RG-SB) for hepatitis A (Epaxal®) and inactivated hemagglutinin of Influenza virus strains A and B for influenza (Inflexal®). 142

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Lipoplexes are fabricated from liposomes and are designed in multi-lamellar lipoplexes with positive lipid bilayer and distinct negative nucleic acids. The electrostatic process between self-assembly liposomes and nucleic acids produces such scaffolds. Given their similarity to liposomes they possess similar benefits and drawbacks with a few of their own; multiple cations tendency to bind negative nucleic acids which decreases the internal cell transfection process. They have been investigated for brain-focused studies.<sup>143</sup>

Natural body lipoproteins are another lipid-based system which are very similar to liposomes hence sharing the same advantages and disadvantages and carry lipids (mainly cholesterol), proteins, enzymes, and miRNAs. They have been investigated with other nanoparticles (e.g., albumin, PEG-PLGA) for numerous central nervous system diseases.<sup>144</sup>

- 2) Niosomes are vesicles with a lamellar self-assembled structure which are composed of non-ionic surfactants and cholesterol or its derivatives. They can be encapsulated by lipophilic and hydrophilic substances. They are cheaper in terms of production and more stable than liposomes. 146
- 3) Transferosomes which are composed of a lipid bilayer fabricated by a lipid matrix stabilized by various surfactants, and are similar to niosomes and to liposomes.
- 4) Solid lipid nanoparticles (SLNs) which are composed of a solid lipid core. 148
- 5) Nanostructured lipid carriers (NLCs) which are composed of a liquid lipid phase core within the solid lipid phase.<sup>149</sup>

Among all the lipid-based DDSs, this review will focus further on SLNs and NLCs and their unique characteristics and properties. SLNs, comparing to NLCs which are formulated with both solid and liquid oils, are formulated only with solid ones which gives them more controlled drug release due to limited drug mobility in solid lipids and are designed as oral pellets and retard capsule (e.g. Mucosolvan®), as microparticles by spray drying, 150 and oral nanopellets. 151

#### SLNs & NLCs

SLNs and NLCs are lipid-based colloidal drug carriers, synthesized from lipids (solid or liquid), surfactants, co-surfactants and active pharmaceutical ingredients (API, drugs). SLNs are composed of solid lipids and surfactants but NLCs are also composed of liquid lipids and oils. Lipids have solid form both at room and body temperature and could be chosen as purified triglycerides, glyceride mixtures and waxes. Surface surfactants increase and improve the stability and cellular permeability and consequently absorption.<sup>152</sup>

They collectively offer the benefits of other colloidal DDSs (e.g. liposomes and polymeric NPs) and avoid their drawbacks;<sup>154</sup> enhanced dissolution rate, bioavailability, tissue distribution, encapsulation rate, absorption, stability of

drug in body fluids, no unpleasant taste (exists with oral preparations), lower toxicity, no organic solvents usage, large-scale production possibility, sterility, reduced first-pass effect and controlled and sustained tissue-targeted delivery with various routes of administration: oral, parenteral, nasal, rectal, ophthalmic, etc.<sup>155</sup>

Due to some drawbacks of SLNs such as low drug loading capacity, unpredictable gelation tendency, and drug expulsion after polymorphic transition during storage, NLCs were introduced and synthesized as the improved version of lipid-based nanocarriers with numerous methods exploited to formulate and prepare them. Hence, the final nanoparticle characterization must comply with the dynamic processes and such a characterization is the real challenge to represent the highest quality for the product, storage drug expulsion, unpredictable gelation and their co-encapsulation with nano- and micro-sized structures owing to high lipid concentration and surfactants might influence the in-vivo fate of nanoparticles.

The major criteria for the characterization of nanoparticles as efficient and safe DDSs could be addressed as: size, encapsulation efficiency, structure, co-existence of material, surface morphology and functionalization, and minimum drug dose, which arguably influence directly the bioavailability, absorption and distribution of the encapsulated drug (Table 4).

Model	Drug loading site	Drug release pattern
Homogenous matrix of solid	Homogeneous drug dispersion	Diffusion from the solid lipid
solution [158, 159]	in the lipid matrix of the	matrix and/or by degradation
	particles	of lipid matrix in GI
Drug-enriched shell [158, 159]	Drug concentration on the	Burst release (160) modified by
	outer shell of the	varying the formulation
	nanoparticles	conditions: production
		temperature (preferably cold
		homogenization) and
		surfactant concentration (161)
Drug-enriched core [158, 159]	Drug concentration in the core	Prolonged drug release (161)
	of the nanoparticles	

Table 4: Models of drug incorporation for the lipid nanoparticles

# Types of SLNs

Type 1. Drug molecules/APIs are dispersed either in the lipid core or as amorphous clusters in "the homogenous matrix model", which offers controlled release features. In order to design and fabricate this type, appropriate concentration of API/lipid ratio must be adjusted using either above-melting point of the lipid or cold methods of High-Pressure Homogenization (HPH).<sup>162</sup>

Type 2. This type, known as "Drug enriched shell model", is designed and fabricated with low concentration of API in the melted lipid. Using the hot method of HPH, the lipid phase is precipitated during the cooling phase which leaves a higher concentration of API in the residue of melted lipid leading to the formation of a free-API lipid core being surrounded by an outer shell composed of the saturated API and lipid. This type doesn't function for sustained release, however, it does for burst release of API.<sup>162</sup>

Type 3. Hence "Drug enriched core model", it is designed by solubilizing the drug in the melted lipid up to its saturation solubility. Following cooling of the lipid the drug is super-saturated in the melted lipid and recrystallizes before the lipid does. Further cooling process renders also lipid recrystallization surrounding the prior formed drug-enriched core. This type offers sustained and prolonged drug release.<sup>162</sup>

#### Types of NLCs

Imperfect. They are named "imperfect" due to the tiny pores in the solid matrix core which are loaded with API. They are designed by adding and blending solid and liquid lipids (oil) which the co-presence of fatty acids with different chain length (mono-, di- and triacylglycerols) confers an imperfect structure for encapsulating the API. 163

Amorphous. They are designed blending lipids which don't crystallize after homogenization and cooling process,<sup>164</sup> such as hydroxyl octacosanyl hydroxy stearate, isopropyl myristate and dibutyl adipate, that give them an unorganized amorphous matrix which reduces API repelling of storage and shelf time.<sup>163</sup>

Multiple. The advantage of the higher solubility of lipophilic drugs in liquid lipids than solid lipids can be used for formulating multiple types of NLC. Solid lipids are mixed with oils which are gradually added in higher amounts exceeding their solubility which results in phase separation of tiny particles with the surrounding solid lipid matrix. This type offers controlled drug release without expelling it out of the lipid matrix. <sup>163</sup>

Nevertheless, even the lipid-based drug delivery sustems have their own advantages and disadvantages (Table 5).

Table 5: Advantages and disadvantages of lipid-based drug delivery systems

Characteristics	Biological/Technological aspects	Perspectives	Reference
	Various administration routes	Broad-spectrum drug application, therapy optimization	165
	Biodegradability	Sustained drug release	166
	Controlled drug release	Patients safety, prolonged drug release, insite drug concentration	167
	Site-specific targeting	Decreased systemic toxicity, targeted therapy	168
	Biocompatibility	No allergic reactions	169

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	Increased bioavailability of encapsulated	Decreased dose	170
	drug		
	Decreased adverse effects of toxic drugs	Improved patient safety	171
	Biological barriers penetration	Various administration routes	172
	Reduced dosing frequency	Patients compliance	167
Advantages	Chemical and enzymatic degradation	Possibility of broad-spectrum administration	168
	drug protection	routes	
	Physical stability	Improved drug formulation stability	173
	Capacity to encapsulate hydrophilic and	Versatility for different drug groups	174
	hydrophobic drugs		
	Scaled up production	Industrial production possibility	165
	Simple manufacturing	Easy fabrication in labs, low cost	166
	No organic solvents	No toxicity concerns, green chemistry	169
	Co-delivery	Offering combined therapy	166
	Increased drug loading capacity	Decreased formulation dose	173
	Sterilization possibility Parenteral administration optimization		175
	Small size distribution	Potential alternative for drug delivery	176
	Initial burst effect of encapsulated drug	Patient overdose risk	177
	Low plasma circulation time	Fast reticuloendothelial clearance before in-	178
		site deposition	
	Drug expulsion during storage	Storage and administration stability	179
		challenges, industrial-scale limitation	
Disadvantages	Low drug loading capacity	High requirement of formulation doses	180
	Polydispersity	Undesirable for intravenous administration	181
	Agglomeration	Storage issues	182
	Storage in refrigerated conditions	Transportation issues, expensive storage	183
		costs	
	High operative temperature	Susceptibility of thermolabile drugs	184

#### Mechanisms of drug release by SLNs/NLCs

To improve the benefits and avoid the drawbacks, different criteria have been considered for formulation, design and encapsulation rate of nanomedicines. The encapsulated drug undergoes surface dissolution and degradation of the lipid matrix which results in diffusion of molecules from the matrix into the surrounding tissue. Drug release from SLNs/NLCs is affected by the localization of the drug, which can be loaded both in the core matrix and on the surface, the former results in prolonged and sustain release while the latter burst release (quick early-phase release), subsequently conferring a biphasic release profile starting with an quick release owing to the surface-loaded drug continuing by sustained release of more loaded-drug from the lipid matrix.

The burst release is a feature determined by modifying drug aqueous solubility which is influenced mainly by the surfactant concentration and the temperature via a direct proportion; the higher the last two the higher the burst release. Preparation of SLN/NLC nanoparticles at room temperature has demonstrated no burst release owing to no drug partitioning into water phase and following re-partitioning into lipid phase. Therefore, to decrease the burst release SLNs/NLCs have been prepared without surfactants or surfactants not solubilizing the drug.<sup>185</sup>/187

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#### In-vivo fate of SLNs/NLCs after administration

There are different criteria which determine the in-vivo fate of nanoparticles: administration route, biological interactions with their environment including distribution, surface adsorption, nanoparticle disaggregation and enzymatic degradation.

Due to the presence of lipids and waxes in the structure of lipid-based nanoparticles, their fate is highly affected by different pathways and enzymes for their biological interactions, namely as lipases which exist ubiquitously in body and mostly activate by oil/water interface, 188-190 and actively confer various degradation rates to nanoparticles due to their formulation material 191-194 the free fatty acids of degradation have been studied by enzymatic test, 195 which demonstrated lesser degradation with long-chain fatty acids of triglycerides and surfactants contained in nanoparticles. Surfactants (e.g. poloxamer 407, poloxamer 188) function either to fasten or postpone the degradation process of nanoparticles, as different surfactants (e.g. cetyl palmitate) have different chain lengths, and this feature could be used in nanoparticle preparation to render a more controlled drug release profile.

So far, there has been few studies,<sup>196</sup> approving whether the presence of food in stomach would affect nanoparticles' in-vivo function or not, and this still remains a dilemma to be solved. In one animal study increased bioavailability and blood half-time was reported with oral administration of lipid nanodispersions,<sup>197</sup> and in another study the increased absorption of nanoparticles into lymph through intraduodenal route was reported.<sup>198</sup>

#### In-vivo toxicity evaluation of SLNs/NLCs

Along with their different therapeutic application as DDSs, SLNs and NLCs have been investigated for their in-vivo toxicity/safety profile. Due to their composition of lipids which are physiological components they are generally recognized as safe (GRAS) and better-tolerated nanoparticles than polymeric ones showing lower toxicity, 199'200 as they are degraded by normal physiological pathways. Nevertheless, the type of lipids and surfactants (emulsifiers) used for their preparation might increase or decrease cell toxicity and even influence encapsulated drug toxicity. 201'202 Therefore, the toxicity evaluation must include bulk materials, SLNs/NLCs, drug/API itself and drug-encapsulated nanoparticles to analyze thoroughly each component and materials contribution to toxicity. The excipients role for drug encapsulation must be assessed according to the route of administration. 203

Different in-vitro tests, among all cell viability (MTT assay) and oxidative stress, have been exploited to assess the cytotoxicity of nanoparticles; the former functions as the color-change of tetrazolium which is the indication of cell death,

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and the latter demonstrates DNA damages, elevated amounts of reactive oxygen species, lipid peroxidases and alterations in oxidation/reduction glutathione reactions.<sup>204</sup> MTT assay is the most common method used with different dyes such as Neutral red, Trypan blue,<sup>204–206</sup> and there are in-vitro and in-vivo experiments on cells proving their low toxicity.<sup>207/208</sup>

#### SLNs and NLCs co-delivery strategies

Through numerous cancer-related studies, it has been proved that broad-spectrum anti-cancer agents will have many benefits in terms of efficacy over monotherapy. SLNs and NLCs could be a promising carrier for co-delivery of anti-cancer, therapeutic nucleic acids and antibiotics, <sup>209</sup> as they are significantly able to enhance the in-vitro and in-vivo therapeutic efficacy of such drugs. Besides such advantage, co-encapsulation of different drugs in one LPDDS might decrease toxicity of the respective anti-cancer drugs and other adverse effects coming with them separately. <sup>210</sup>

Another alternative being offered by co-delivery is RNA interference,<sup>211</sup> especially siRNAs have been exploited for cancer cells to silence the oncogenes expression.<sup>209</sup> In this context, miRNAs have been investigated and proved to be efficient to regulate genes associated with tumorigenesis.<sup>210</sup>

In one study,<sup>209</sup> cationic SLNs for co-delivery of paclitaxel and human myeloid cell leukemia (MCL1) specific siRNA have been investigated and the final result demonstrated enhanced in-vitro and in-vivo efficacy than administering them separately.

In another similar study,<sup>210</sup> SLNs were encapsulated for co-delivery of the same active substance with miRNA-34a. The final result was significant in terms of eliminating lung cancer relapse mostly owing to the synergic efficacy and higher inhibition of specific receptors.

In another study,<sup>212</sup> the efficacy of Paclitaxel and Verapamil co-loaded SLNs toward breast cancer were investigated to prove the efficiency of verapamil for inhibiting drug efflux transporters (e.g. p-glycoprotein) on multidrug resistance cancer cells. The study demonstrated higher expression downregulation of g-glycoproteins in the specific cancer cells, as well as higher cellular drug uptake and toxicity comparing to Paclitaxel and sole anti-cancer administration.

In another study on antibiotics,<sup>213</sup> increased antibacterial activity of Vancomycin was demonstrated. Ion pairing with linoleic acid was exploited for co-delivery and significant effects against Staphylococcus aureus infections which could be interpreted owing to the increased lipophilicity, sustained release of antibiotic and synergistic effect.

So far different pharmaceutical/biotechnological products have been marketed using lipid-based drug delivery systems (Table 6).

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Table 6: List of marketed lipid-based oral pharmaceutical products

Product/Tra	able 6: List of marketed lipid-based oral pharmaceutical products    Drug   Drug   Malagula   Nanotachnology   Doca   Thereposition   Company   Allian					
de name	Drug/Molecule	Nanotechnology/Dosa ge form	Therapeutic use/Indication	Company/Allian ce		
	Amphatariain D	-	-			
Abelcet	Amphotericin B	Nanoliposome/solutio	Fungal infections	The Liposome		
A t	la atuatia aia	n Faculais a /a oft a platin a	A ±:	Company Inc		
Accutane	Isotretinoin	Emulsion/soft gelatine	Anti-	Roche		
		capsule	comedogenic			
Agenerases	Amprenavir	Soft gelatine capsule	HIV antiviral	GlaxoSmithKline		
ALEC	Dry protein free	Liposome	Lung diseases in	<ul><li>Britannia</li></ul>		
	powder of DPPC-		infants	Pharmaceuticals		
	PG			Ltd		
Ambisome	Amphotericin B	Powder	Fungal infections	NeXstar		
				Pharmaceutical Pharmaceutical		
				Inc		
Amphocil	Amphotericin B	Colloidal dispersion	Fungal infections	Sequus		
				Pharmaceutical		
				Inc		
Amphotec	Amphotericin B	Nanoliposome/Solutio	Fungal infections,	Sequus		
		n	leishmaniasis	Pharmaceutical		
		<b>A</b> ( /		Inc		
Aptivus	tipranavir	Emulsion/soft gelatine	AIDS	Boehringer		
	·	capsule		Ingelheim		
Atragen	Tretinoin	Liposome	Acute myeloid	Aronex		
			leukemia	Pharmaceuticals		
				Inc		
Avodart	Dutasteride	Emulsion	Benign Prostatic	GSK		
			Hyperplasia			
Avian	Killed avian	Suspension	Chickenpox	Vineland		
retrovirus	retrovirus			Laboratories		
vaccine						
Cipro	Ciprofloxacin	Oral suspension	Antibiotic	Bayer		
Convulex	Valproic acid	Soft gelatine capsule	Antiepileptic	Pharmacia		
DaunoXome	Daunorubicin	Solution	Kaposi sarcoma in	NeXstar		
Budiloxionic	citrate	Solution	AIDS	Pharmaceutical		
	Citiate		71105	Inc/ Galen Ltd		
Depakene	Valproic acid	Emulsion	Epilepsy	Abbott		
Depocyt	Cytarabin	Nanoliposome/Solutio	Lymphomatous	Pacira		
Веросус	Cytarabiii	n	meningitis	Pharmaceuticals		
		"	inclingitis	Inc		
DepoDur	Morphine	Cusponsion	Doct curgical pain	Pacira		
Беробиг	Morphine	Suspension	Post-surgical pain reliever	Pharmaceuticals		
			reliever			
Devil	Dovorubisis	Colution	Motostatia	Inc		
Doxil	Doxorubicin	Solution	Metastatic	Sequus		
			ovarian, Kaposi	Pharmaceutical		
	A	No. 10 /0	sarcoma in AIDS	Inc		
Emend	Aprepitant	Nanosuspensions/Cap	Antiemetic	Merck-Elan		
		sule		Drug Delivery		

- 15		Ι .		
Epaxal Berna	Inactivated	Suspension	Hepatitis A	Swiss serum &
Vaccine	hepatitis-A			vaccine institute
Estus es ale	Virions	Taminal amazdaina	N/anananal	Navasas
Estrasorb	estradiol	Topical emulsion	Menopausal	Novavax
	D 1.1.1.	12	therapy	Th
Evacet	Doxorubicin	Liposome	Metastatic breast	The liposome
F	E Ch i .	T. I. I	cancer	company
Fenogal	Fenofibrate	Tablet	Anti	Genus
			hyperlipproteino	
Fortovoso		Cooptonoously	mic	A Do ele e
Fortovase	saquinavir	Spontaneously	HIV antiviral	Roche
		emulsifying		
		systems/soft gelatine		
Fungizono	Amphotericin B	capsule Solution	Fungal infections	Bristol-Myers
Fungizone	Amphotericin B	301011011	Fullgal IIIIections	Squibb
Gengraf	Cyclosporin A/III	Spontaneously	Immuno-	Abott
Gengrai	Cyclosporiii Ayiii	emulsifying	suppressant	Abott
		systems/hard gelatine	Suppressure	
		capsule		
Hectoral	Doxercalciferol	Emulsion	Calcium regulator	Bone care
Juvela	Tocopherol	Capsule	Hypertension,	Eisai Co.
	nicotinate		hyperlipidemic	
Kaletra	Lopinavir &	Emulsion/oral solution	HIV antiviral	Abott
	Ritonavir			
Lamprene	Clofazamine	Emulsion	Leprosy	Alliance
·				laboratories/
				Geigy
Lipirex	fenofibrate	Hard gelatine capsule	hyperlipidemia or	Sanofi-Aventis
			mixed	
			dyslipidemia	
Marinol	Dronabionol	Emulsion	Anoxeria	Roxane
Megace ES	Megestrol	Nanosuspension	anorexia,	Par
	acetate		cachexia, weight	Pharmaceuticals
			loss in HIV	- Elan Drug
			patients	Delivery
MiKasome	Amikacin	Liposome	Bacterial	NeXstar
			infection	Pharmaceutical
				Inc
Neoral	Cyclosporin A/I	Emulsion	Immunosuppress	Novartis
▼			ant	
Norvir	Ritonavir	Spontaneously	HIV antiviral	Abott
		emulsifying		
		systems/soft gelatine		
		capsule		-
Nyotran	Nystatin	Liposome	Fungal infections	Aronex
				pharmaceuticals
				Inc

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Panzem NCD	2-Methoxy estradiol	Nanosuspension	anti-proliferative and anti- angiogenic effect	EntreMed Inc.
Prometrium	Progesterone	Emulsion	Endometrial hyperplasia	Solvay
Rapamune	Sirolimus	Nanosuspensions/Tabl et	Immunosuppress ant	Wyeth Pharmaceuticals – Elan Drug Delivery
Restandol	Testosterone undecanoate	Capsules	Hormone replacement therapy	Organon laboratories
Rocaltrol	Calcitriol	Emulsion/soft gelatine capsule	Calcium regulator	Roche
Sandimmun e Neoral	cyclosporine A/I	Spontaneously emulsifying systems/soft gelatine capsule	Immunosuppress ant	Novartis
Sustiva	Efavirenz	Capsules	HIV antiviral	Bristol-Meyers
Targretin	bexarotene	Soft gelatine capsule	liver cancer	Novartis
Topex-Br	Terbutalinesulph ate	Syrup	Asthma	Ozone Pharmaceuticals Ltd
Tricor	Fenofibrate	Nanosusp <mark>ensions/</mark> Tabl et	Antihyperlipidemi c agent	Abbott Laboratories
Triglide	Fenofibrate	Nanosuspensions/Tabl et	Antihyperlipidemi c agent	Skye Pharma- First Horizon
Ventus	Prostaglandin-E1	Liposome	Systemic inflammatory disease	The liposome company
Vesanoid	tretinoine	Emulsion/soft gelatine capsule	Acne	Roche
VincaXome	Vincristine	Liposome	Solid tumors	NeXstar Pharmaceutical Inc
Zemplar	Paricalcitol	Emulsion	Calcium regulator	Abbott

# LBDDSs formulations to enhance oral delivery of hydrophobic peptide/protein-based drugs

Although there have been promising achievements with LBDDSs for oral delivery of hydrophobic peptide-based drugs, the hydrophilic peptide-based drugs delivery still remains a challenge and limited to the in-vitro and in-vivo experiments with no product in the pharmaceutical market.

There have been many studies using these lipid-based scaffolds to prove their potential to be exploited in future studies. there have been numerous studies of Insulin as a hydrophilic peptide encapsulated in micelles, microemulsion NPs

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and nanocapsules with in-situ and in-vivo experiments on rat with promising results as enhanced permeability, bioavailability and efficacy.<sup>214–220</sup> In one study SK&F 106760 (a hydrophilic RGD peptide) in the form of microemulsion exploited in in-vivo experiments and demonstrated 50-fold elevated bioavailability.<sup>221</sup> In another study Vasopressin was encapsulated microemulsion in-situ experiments and resulted in enhanced bioavailability.<sup>222</sup> In one study EGF (a single-chain polypeptide) was encapsulated in microemulsions for in-vivo experiments of gastric ulcer in rats and showed increase efficacy.<sup>223</sup> In one study on ß-lactamase in-vivo bioavailability.<sup>224</sup> N-2.5-fold experiments resulted in enhanced acetylglucosaminyl and N-acetylmuramyl dipeptide were exploited in one study which demonstrated 10-fold increased bioavailability.<sup>225</sup> In another study Leuprolide acetate was encapsulated in microemulsion for in-vivo experiments and proved increased efficacy. <sup>226</sup> Two experiments of lipid mixtures with Hexarelin and DMP 728 (Cyclic peptide fibrinogen antagonist) as encapsulated drugs with in-situ experiments showed 20-fold and 3-fold intestinal permeability and bioavailability, respectively. 227/228 In the latter study in dog, DuP 532 (an Angiotensin II antagonist) was encapsulated in microemulsion in in-vivo experiments which resulted in 3-fold bioavailability.<sup>228</sup> In three studies in rat and pig, calcitonin was encapsulated in mixed micelles and emulsion for in-situ and in-vivo experiments which demonstrated increased permeability and efficacy and 4-fold hypocalcemia response. 229-231 Human growth hormone was encapsulated in in-vivo studies on rabbit and showed 3.3% increased bioavailability.<sup>232</sup>

#### Regulatory status, commercialization plan and safety information

The status of excipients should be assessed with the regulatory authorities before any pharmaceutical product's introduction into the market <sup>233</sup> but the expenses of in-vivo toxicity studies are prohibitive for the companies. Such a challenge is happening mainly with the polymeric NPs as there are few of them in the market but lipid NPs owing to their various applications of oils, fats, stabilizer and surfactants have introduced oral and dermal products. The majority of the introduced excipients so far for lipid NPs synthesis are biodegradable, biocompatible and are approved as safe, but some are toxic at high concentrations. <sup>234</sup> In this context the FDA has published guide lists of safe materials and substances (GRAS) and Inactive Ingredient Guide (IIG) for excipients that are approved for exploiting in the pharmaceutical products in the market. <sup>235</sup> These lists explain and provide insights regarding the appropriate excipient concentration for each administration route and the approved inactive ingredients used for a specific route can be used in all the new formulations. This

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facilitates the process of synthesizing new formulation as the necessary information can be extracted from the GRAS and IIG. Therefore, such excipients are assessed as substances of a drug not individually as from a "scientific point of view" excipients are a major part of the drug formulation.

Nevertheless, there are other challenges to consider from a "regulatory point of view"; preclinical and clinical studies addressing safety issues and in-vivo manifestations of the LBDDSs in terms of clinical therapeutic efficacy. In-vivo immunological and stability findings toward oils and lipid excipients must be reported to provide in-depth information for the regulatory authorities.<sup>236</sup>

Besides, factors coming from the biopharmaceuticals are required to be evaluated toward the drug or excipients and this might have paradoxical in-vitro results with in-vivo results due to the physiology of GI tract. Various experiments must be designed and conducted to characterize and recognize the interactions happening among excipients, in-vivo physiological conditions and the drug.<sup>237</sup>

In order to understand and characterize the in-vivo fate of drugs encapsulated in LBDDSs, a consortium of academic and industrial scientists has been established (<a href="http://www.lfcsconsortium.org">http://www.lfcsconsortium.org</a>) which designs experiments to evaluate the function of LBDDSs dispersion and digestion as vital criteria.

#### **Conclusion**

Nanotechnology offers promising strategies for enhancing oral bioavailability and therapeutic efficacy of a vast range of drugs; conventional chemical drugs with poor water solubility and biotechnological, peptide/protein-based drugs and biopharmaceuticals. Regarding the latter, their unique physicochemical and biopharmaceutical features pose challenge for their oral delivery. Hence, their success in site delivery highly depends on technologies and methods to modify these two features not influencing their biological function. In the recent decades numerous DDSs have been introduced and offered by nanotechnology to achieve as high successful delivery as possible and LBDDSs among all has been under investigation owing to their potential for oral delivery of hydrophilic, hydrophobic and lipophilic peptide- and protein-based drugs.

LBDDSs enhance solubility and bioavailability of drugs offering strategies such as gastrointestinal lymphatic transport, altering physiological and biochemical properties of gastrointestinal barriers, elevated solubilization and prolonged gastrointestinal retention. Although, such improvements rely on the encapsulation/loading rate and intrinsic composition of the material used during the fabrication process. Obviously, the choice of materials, such as excipients, will influence the success of delivery route which is determined both by lipid formulation design and peptide/protein molecule emphasizing that each

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peptide/protein-loaded LBDDS must be designed uniquely. Such material must be in correlation with the drug of choice to achieve the maximum therapeutic efficacy and in-site dose.

Most of the scaffolds described in this review article suggest promising alternatives to overcome gastrointestinal enzymatic degradation and poor membrane penetration. Further systematic studies are required to evaluate their in-vivo efficacy in terms of peptide-/protein-based oral drug delivery. Besides "pharmaco-biotechnological" challenges mentioned in this review such as membrane permeability, protease stability, delivery strategies and increased circulation half-life, there are inevitably several "industrial" challenges as well which finally hamper their industrial scale production and consequently their biomedical translation from lab to pharmaceutical market. "Oral bioavailability" still remains the main challenge of peptide/protein-based drug delivery. These factors could be addressed as materials cost, drug potential market feedback, regulatory status, simple industrial-scale fabrication, financial schemes for required instruments, patient compliance administration and high adaptability to human diverse pharmacokinetics.

#### Ethical approval

There is none to be disclosed.

# **Conflict of Interests**

The author declares no conflict of interest.

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#### Reference

- 1. Vlieghe P, Lisowski V, Martinez J, Khrestchatisky M. Synthetic therapeutic peptides: science and market. Drug Discovery Today. 2010 Jan;15(1-2):40–56. DOI: 10.1016/j.drudis.2009.10.009
- 2. Lalatsa A, Schatzlein AG, Uchegbu IF. Strategies To Deliver Peptide Drugs to the Brain. Molecular Pharmaceutics. 2014 Mar 21;11(4):1081–93. DOI: 10.1021/mp400680d
- 3. Vajo Z, Fawcett J, Duckworth WC. Recombinant DNA Technology in the Treatment of Diabetes: Insulin Analogs. Endocrine Reviews [Internet]. 2001 Oct;22(5):706–17. DOI: 10.1210/edrv.22.5.0442
- 4. Takeda A, Cooper K, Bird A, Baxter L, Frampton G, Gospodarevskaya E. Recombinant human growth hormone for the treatment of growth disorders in children: a systematic review and economic evaluation. Health Technology Assessment. 2010 Sep;14(42). DOI: 10.3310/hta14420
- 5. Cutting GR. MODIFIER GENETICS: Cystic Fibrosis. Annual Review of Genomics and Human Genetics. 2005 Sep;6(1):237–60. https://doi.org/10.1146/annurev.genom.6.080604.162254
- 6. Weatherall DJ. Phenotype—genotype relationships in monogenic disease: lessons from the thalassaemias. Nature Reviews Genetics. 2001 Apr;2(4):245–55. DOI: 10.1038/35066048

- 7. Powell JS. Lasting power of new clotting proteins. Hematology. 2014 Dec 5;2014(1):355–63. DOI: 10.1182/asheducation-2014.1.355
- 8. Hirschhorn JN, Lohmueller K, Byrne E, Hirschhorn K. A comprehensive review of genetic association studies. Genetics in Medicine. 2002 Apr;4(2):45–61. DOI: 10.1097/00125817-200203000-00002
- 9. Savic S, McDermott MF. New monogenic diseases span the immunological disease continuum. Nature Reviews Rheumatology. 2014 Dec 23;11(2):67–8. DOI:10.1038/nrrheum.2014.215
- 10. Hussain N. Recent advances in the understanding of uptake of microparticulates across the gastrointestinal lymphatics. Advanced Drug Delivery Reviews. 2001 Aug 23;50(1-2):107–42. DOI: 10.1016/s0169-409x(01)00152-1
- 11. Banga AK. Drug delivery today, Pharma Tech 2002, Business Briefing World Market Series, London, 2002, 150–154.
- 12. Wang W. Instability, stabilization, and formulation of liquid protein pharmaceuticals. International Journal of Pharmaceutics. 1999 Aug;185(2):129–88. DOI: 10.1016/s0378-5173(99)00152-0
- 13. Hillery AM. Drug Delivery and Targeting. 1st Edition. Lloyd AW, Swarbrick J, editors. London: CRC Press; 2001. https://doi.org/10.1201/b12801
- 14. Frokjaer S, Otzen DE. Protein drug stability: a formulation challenge. Nature Reviews Drug Discovery. 2005 Apr;4(4):298–306. DOI: 10.1038/nrd1695
- 15. Wang W. Protein aggregation and its inhibition in biopharmaceutics. International Journal of Pharmaceutics. 2005 Jan;289(1-2):1–30. DOI: 10.1016/j.ijpharm.2004.11.014
- 16. Rubert Pérez CM, Stephanopoulos N, Sur S, Lee SS, Newcomb C, Stupp SI. The Powerful Functions of Peptide-Based Bioactive Matrices for Regenerative Medicine. Annals of Biomedical Engineering. 2014 Nov 4;43(3):501–14. DOI: 10.1007/s10439-014-1166-6
- 17. Lax RT. The future of peptide development in the pharmaceutical industry.

Pharmanufacturing: The International Peptide Review, World Business Journals, Pharmaceutical Division, London. 2010.

- 18. Serrano Lopez DR, Lalatsa A. Peptide pills for brain diseases? Reality and future perspectives. Therapeutic Delivery. 2013 Apr;4(4):479–501. DOI: 10.4155/tde.13.5
- 19. Adessi C, Soto C. Converting a Peptide into a Drug: Strategies to Improve Stability and Bioavailability. Current Medicinal Chemistry. 2002 May 1;9(9):963–78. DOI: 10.2174/0929867024606731
- 20. Adessi C, Soto C. Strategies to Improve Stability and Bioavailability of Peptide Drugs. Frontiers in Medicinal Chemistry Online. 2004 Jan 1;1(1):513–28. Doi: 10.2174/978160805204210401010513
- 21. Park K, Kwon IC, Park K. Oral protein delivery: Current status and future prospect. Reactive and Functional Polymers. 2011 Mar;71(3):280–7. https://doi.org/10.1016/j.reactfunctpolym.2010.10.002
- 22. Donovan MD, Flynn GL, Amidon GL. Absorption of polyethylene glycols 600 through 2000: the molecular weight dependence of gastrointestinal and nasal absorption. Pharmaceutical Research. 1990;07(8):863–8. DOI: 10.1023/a:1015921101465
- 23. Ikesue K, Kopečkovà P, Kopeček J. Degradation of proteins by guinea pig intestinal enzymes. International Journal of Pharmaceutics. 1993 Jun;95(1-3):171–9. https://doi.org/10.1016/0378-5173(93)90404-4
- 24. Saffran M, Kumar G, Savariar C, Burnham J, Williams F, Neckers D. A new approach to the oral administration of insulin and other peptide drugs. Science. 1986 Sep 5;233(4768):1081–4. DOI: 10.1126/science.3526553

- 25. Fix JA. Oral controlled release technology for peptides: status and future prospects. Pharmaceutical Research. 1996;13(12):1760–4. DOI: 10.1023/a:1016008419367
- 26. Lee HJ. Protein drug oral delivery: The recent progress. Archives of Pharmacal Research. 2002 Oct;25(5):572–84. DOI: 10.1007/BF02976925
- 27. Vincent HL, Satish DK, George MG, et al. Oral route of protein and peptide drug delivery, in: H.L. Vincent (Ed.), Peptide and protein drug delivery, Marcel Dekker, New York, 1991: 691–738.
- 28. Pettit DK, Gombotz WR. The development of site-specific drug-delivery systems for protein and peptide biopharmaceuticals. Trends in Biotechnology. 1998 Dec;16(8):343–9. DOI: 10.1016/s0167-7799(98)01186-x
- 29. Tauzin B, Report: Biotechnology Medicines in Development, Pharmaceutical Research and Manufacturers Association, Washington DC, 2006.
- 30. Crommelin D, van Winden E, Mekking A. Delivery of pharmaceutical proteins, in: M.E. Aulton (Ed.), Pharmaceutics: The Science of Dosage Forms Design, Churchill Livingstone, Edinburgh, 2001, pp. 544–553.
- 31. Crommelin DJA, Storm G, Verrijk R, et al. Shifting paradigms: biopharmaceuticals versus low molecular weight drugs. Int. J. Pharm. 2003;266:3–16. DOI: 10.1016/s0378-5173(03)00376-4
- 32. Saltzman M. Drug Delivery: Engineering Principles for Drug Therapy, Oxford University Press, New York, 2001. https://doi.org/10.1093/oso/9780195085891.003.0005
- 33. Ugwoke MI, Agu RU, Verbeke N, et al. Nasal mucoadhesive drug delivery: background, applications, trends and future perspectives. Adv. Drug Deliv. Rev. 2005;57:1640–1665. DOI: 10.1016/j.addr.2005.07.009
- 34. Myles ME, Neumann DM, Hill JM. Recent progress in ocular drug delivery for posterior segment disease: emphasis on transscleral iontophoresis. Adv. Drug Deliv. Rev. 2005;57:2063–2079. DOI: 10.1016/j.addr.2005.08.006
- 35. Smart JD. Buccal drug delivery. Expert Opin Drug Deliv. 2005;2:507–517. DOI: 10.1517/17425247.2.3.507
- 36. Mackay M, Phillips J, Hastewell J. Peptide drug delivery: colonic and rectal absorption. Adv. Drug Deliv. Rev. 1997;28:253–273. doi.org/10.1016/S0169-409X(97)00076-8
- 37. Hussain A, Ahsan F. The vagina as a route for systemic drug delivery. J. Control. Release. 2005;103:301–313. DOI: 10.1016/j.jconrel.2004.11.034
- 38. Schuetz YB, Naik A, Guy RH, et al. Emerging strategies for the transdermal delivery of peptide and protein drugs. Expert Opin Drug Deliv. 2005;2:533–548. DOI: 10.1517/17425247.2.3.533
- 39. Agu RU, Ugwoke MI, Armand M, et al. The lung as a route for systemic delivery of therapeutic proteins and peptides. Respir. Res. 2001;2:198–209. DOI: 10.1186/rr58
- 40. Bosquillon C, Préat V, Vanbever R. Pulmonary delivery of growth hormone using dry powders and visualization of its local fate in rats. J. Control. Release. 2004;96:233–244. DOI: 10.1016/j.jconrel.2004.01.027
- 41. Ghilzai NMK, Desai A. Facing the challenges of transmucosal absorption—buccal, nasal and rectal routes, Pharma Tech 2004, Business Briefing World Market Series, London, 2004, pp. 104–106.
- 42. Florence AT, Attwood D. Physicochemical Principles of Pharmacy, Pharmaceutical Press, London, 2006.
- 43. Cleland JL, Langer R. Formulation and delivery of proteins and peptides: design and development strategies, in: J.L. Cleland, R. Langer (Eds.), Formulation and Delivery of Proteins and Peptides, American Chemical Society, Washington DC, 1994, pp. 1–19.

- 44. Almeida AJ, Souto E. Solid lipid nanoparticles as a drug delivery system for peptides and proteins. Adv Drug Deliv Rev. 2007;59(6):478-90. DOI: 10.1016/j.addr.2007.04.007
- 45. Wang W. Instability, stabilization, and formulation of liquid protein pharmaceuticals. Int. J. Pharm. 1999;185:129–188. DOI: 10.1016/s0378-5173(99)00152-0
- 46. Metselaar JM, Mastrobattista E, Storm G. Liposomes for intravenous drug targeting: design and applications. Mini Rev. Med. Chem. 2002;4:319–329. DOI: 10.2174/1389557023405873
- 47. Gombotz WR, Pettit DK. Biodegradable polymers for protein and peptide drug delivery. Bioconjug. Chem. 1995;6:332–351. DOI: 10.1021/bc00034a002
- 48. Packhaeuser CB, Schnieders J, Oster CG, et al. In situ forming parenteral drug delivery systems: an overview. Eur. J. Pharm. Biopharm. 2004;58:445–455. DOI: 10.1016/j.ejpb.2004.03.003
- 49. Kompella UB, Lee VHL. Delivery systems for penetration enhancement of peptide and protein drugs: design considerations, Adv. Drug Deliv. Rev. 2001;46:211–245. DOI: 10.1016/s0169-409x(00)00137-x
- 50. Prego C, Torres D, Alonso MJ. The potential of chitosan for the oral administration of peptides. Expert Opin Drug Deliv. 2005;2:843–854. DOI: 10.1517/17425247.2.5.843
- 51. Ugwoke MI, Agu RU, Verbeke N, et al. Nasal mucoadhesive drug delivery: background, applications, trends and future perspectives. Adv Drug Deliv Rev. 2005;57:1640–1665. DOI: 10.1016/j.addr.2005.07.009
- 52. Alpar HO, Somavarapu S, Atuah KN, et al. Biodegradable mucoadhesive particulates for nasal and pulmonary antigen and DNA delivery. Adv Drug Deliv Rev. 2005;57:411–430. DOI: 10.1016/j.addr.2004.09.004
- 53. Schuetz YB, Naik A, Guy RH, et al. Emerging strategies for the transdermal delivery of peptide and protein drugs. Expert Opin Drug Deliv. 2005;2:533–548. DOI: 10.1517/17425247.2.3.533
- 54. Langer R. Where a pill won't reach. Sci Am. 2003;288:50–57. DOI: 10.1038/scientificamerican0403-50
- 55. Myles ME, Neumann DM, Hill JM. Recent progress in ocular drug delivery for posterior segment disease: emphasis on transscleral iontophoresis. Adv Drug Deliv Rev. 2005;57:2063–2079. DOI: 10.1016/j.addr.2005.08.006
- 56. Smart JD. Buccal drug delivery. Expert Opin Drug Deliv. 2005;2:507–517. DOI: 10.1517/17425247.2.3.507
- 57. Choonara BF, Choonara YE, Kumar P, et al. A review of advanced oral drug delivery technologies facilitating the protection and absorption of protein and peptide molecules. Biotechnol Adv. 2014;32:1269-82. DOI: 10.1016/j.biotechadv.2014.07.006
- 58. Ensign LM, Cone R, Hanes J. Oral drug delivery with polymeric nanoparticles: the gastrointestinal mucus barriers. Adv Drug Deliv Rev. 2012;64:557-70. DOI: 10.1016/j.addr.2011.12.009
- 59. des Rieux A, Fievez V, Garinot M, et al. Nanoparticles as potential oral delivery systems of proteins and vaccines: a mechanistic approach. J Control Release. 2006;116(1):1-27. DOI: 10.1016/j.jconrel.2006.08.013
- 60. Prego C, Torres D, Fernandez-Megia E, et al. Chitosan–PEG nanocapsules as new carriers for oral peptide delivery: effect of chitosan pegylation degree. J Control Release. 2006;111(3):299-308. DOI: 10.1016/j.jconrel.2005.12.015
- 61. Amidon GL, Lennernaes H, Shah VP, et al. A theoretical basis for a biopharmaceutic drug classification: the correlation of in vitro drug product dissolution and in vivo bioavailability. Pharm Res. 1995;12:413–20. DOI: 10.1023/a:1016212804288
- 62. Lipinski CA. Avoiding investment in doomed drugs—is solubility an industry wide

- problem? Curr Drug Discov. 2001;1:17–9.
- 63. Murakami T, Takano M. Intestinal efflux transporters and drug absorption. Exp Op Drug Met Tox. 2008;4:923–39. https://doi.org/10.1517/17425255.4.7.923.
- 64. Uekama K, Hirayama F, Irie T. Cyclodextrin drug carrier systems. Chem Rev. 1998;98:2045–2076. DOI: 10.1021/cr970025p
- 65. Report: Biotechnology Medicines in Development, Pharmaceutical Research and Manufacturers Association, Washington DC, 2019.
- 66. Hillery AM. Drug delivery, the basic concepts, in: A.M. Hillery, A.W. Lloyd, J. Swarbrick (Eds.), Drug Delivery and Targeting for Pharmacists and Pharmaceutical Scientists, Taylor & Francis, London, 2001, pp. 1–48.
- 67. Frokjaer S, Otzen DE. Protein drug stability: a formulation challenge. Nat Rev Drug Discov. 2005;4(4):298-306. DOI: 10.1038/nrd1695
- 68. Banga AK. Drug delivery today, Pharma Tech 2002, Business Briefing World Market Series, London, 2002, pp. 150–154.
- 69. Crommelin D, van Winden E, Mekking A. Delivery of pharmaceutical proteins, in: M.E. Aulton (Ed.), Pharmaceutics: The Science of Dosage Forms Design, Churchill Livingstone, Edinburgh, 2001, pp. 544–553.
- 70. Wang W. Instability, stabilization, and formulation of liquid protein pharmaceuticals. Int J Pharm. 1999;185(2):129-88. DOI: 10.1016/s0378-5173(99)00152-0
- 71. Mehrdadi S. Acute Bacterial Meningitis: Diagnosis, Treatment and Prevention. J Arch Mil Med. Online ahead of Print; 6(4):e84749. doi: 10.5812/jamm.84749.
- 72. Mehrdadi S. Drug delivery of solid lipid nanoparticles (SLNs) and nanostructured lipid carriers (NLCs) to target brain tumors. Advanced Pharmaceutical Bulletin, doi: 10.34172/apb.2023.062
- 73. Tsomaia N. Peptide therapeutics: targeting the undruggable space. Eur J Med Chem. 2015;94:459-70. DOI: 10.1016/j.ejmech.2015.01.014
- 74. Global Data 2015. http://www.globaldata.com. 2015.
- 75. Tomita M, Shiga M, Hayashi M, et al. Enhancement of colonic drug absorption by the paracellular permeation route. Pharm Res. 1988;5(12):786-9. DOI: 10.1023/a:1015992819290 76. Pappenheimer JR. Physiological regulation of transepithelial impedance in the intestinal mucosa of rats and hamsters. J Membr Biol. 1987;100:137–148. https://doi.org/10.1007/BF02209146
- 77. Tang VW, Goodenough DA. Paracellular ion channel at the tight junction. Biophys J. 2003;84(3): 1660–1673. doi: 10.1016/S0006-3495(03)74975-3.
- 78. Florence AT. Issues in oral nanoparticle drug carrier uptake and targeting. J Drug Target. 2004;12:65–70. DOI: 10.1080/10611860410001693706
- 79. Burton PS, Conradi RA, Hilgers AR. (B) Mechanisms of peptide and protein absorption: (2) transcellular mechanism of peptide and protein absorption: passive aspects. Adv Drug Deliv Rev. 1991;7:365–385. doi.org/10.1016/0169-409X(91)90014-4
- 80. Giannasca PJ, Giannasca KT, Leichtner AM, et al. Human intestinal M cells display the sialyl Lewis A antigen. Infect. Immun. 1999;67:946–953. doi: 10.1128/iai.67.2.946-953.1999 81. Gebert A, Rothkötter HJ, Pabst R. M cells in Peyer's patches of the intestine, in: W.J. Kwang (Ed.), Int. Rev. Cytol, Academic Press, 1996, pp. 91–159.
- 82. Frey A, Neutra MR. Targeting of mucosal vaccines to Peyer's patch M cells. Behring Inst. Mitt. 1997;98:376–389.
- 83. Clark MA, Hirst BH, Jepson MA. Lectin-mediated mucosal delivery of drugs and microparticles. Adv Drug Deliv Rev. 2000;43:207–223. DOI: 10.1016/s0169-409x(00)00070-3

- 84. Buda A, Sands C, Jepson MA. Use of fluorescence imaging to investigate the structure and function of intestinal M cells. Adv Drug Deliv Rev. 2005;57:123–134. DOI: 10.1016/j.addr.2004.07.014
- 85. Jani PU, Florence AT, McCarthy DE. Further histological evidence of the gastrointestinal absorption of polystyrene nanospheres in the rat. Int J Pharm. 1992;84:245–252. doi.org/10.1016/0378-5173(92)90162-U
- 86. Kataoka K, Tabata J, Yamamoto M, et al. The association of gap junctions with large particles in the crypt epithelium of the rat small intestine. Arch Histol Cytol. 1989;52:81–86. DOI: 10.1679/aohc.52.81
- 87. Lavelle EC, Sharif S, Thomas NW, et al. The importance of gastrointestinal uptake of particles in the design of oral delivery systems. Adv Drug Deliv Rev. 1995;18:5–22. doi.org/10.1016/0169-409X(95)00048-C
- 88. O'Hagan DT. Intestinal translocation of particulates implications for drug and antigen delivery. Adv Drug Deliv Rev. 1990;5:265–285. doi.org/10.1016/0169-409X(90)90020-S
- 89. Hebden JM, Wilson CG, Spiller RC, et al. Regional differences in quinine absorption from the undisturbed human colon assessed using a timed release delivery system, Pharm Res. 1999;16:1087–1092. DOI: 10.1023/a:1018948102778
- 90. Shah D, Shen WC. Transcellular delivery of an insulin-transferrin conjugate in enterocytelike Caco-2 cells. J Pharm Sci. 1996;85:1306–1311. DOI: 10.1021/js9601400
- 91. Barry PH. Ionic permeation mechanisms in epithelia: biionic potentials, dilution potentials, conductances, and streaming potentials. Methods Enzymol. 1989;171:678-715. DOI: 10.1016/s0076-6879(89)71038-7
- 92. Powell DW. Barrier function of epithelia. Am J Physiol. 1981;241:275–288. DOI: 10.1152/ajpgi.1981.241.4.G275
- 93. Rubas W, Cromwell ME, Shahrokh Z, et al. Flux measurements across Caco-2 monolayers may predict transport in human large intestinal tissue. J Pharm Sci. 1996;85:165–169. DOI: 10.1021/js950267+
- 94. Madara JL. Regulation of the movement of solutes across tight junctions. Annu Rev Physiol. 1998;60:143-59. DOI: 10.1146/annurev.physiol.60.1.143
- 95. Shakweh M, Ponchel G, Fattal E. Particle uptake by Peyer's patches: a pathway for drug and vaccine delivery. Expert Opin Drug Deliv. 2004;1(1):141-63. DOI: 10.1517/17425247.1.1.141
- 96. Russell-Jones GJ, Carrier-mediated transport, oral drug delivery, in: E. Mathiowitz (Ed.), Encyclopedia of controlled drug delivery, 1, JohnWiley& Sons, New York, NY, 1999, pp. 173–184.
- 97. Barthe L, Woodley J, Houin G. Gastrointestinal absorption of drugs: methods and studies. Fundam Clin Pharmacol. 1999;13(2):154-68. DOI: 10.1111/j.1472-8206.1999.tb00334.x
- 98. Bai JPF, Amidon GL. Structural Specificity of Mucosal-Cell Transport and Metabolism of Peptide Drugs: Implication for Oral Peptide Drug Delivery. Pharm. Res. 1992;9:969–978. doi.org/10.1023/A:1015885823793
- 99. Russell-Jones GJ. The potential use of receptor-mediated endocytosis for oral drug delivery. Adv Drug Deliv Rev. 1996;20:83–97. doi.org/10.1016/0169-409X(95)00131-P
- 100. Swaan PW. Recent advances in intestinal macromolecular drug delivery via receptor-mediated transport pathways. Pharm Res. 1998;15(6):826-34. DOI: 10.1023/a:1011908128045 101. Charman WN, Porter CJH. Lipophilic prodrugs designed for intestinal lymphatic transport. Adv Drug Deliv Rev. 1996;19:149–169. doi.org/10.1016/0169-409X(95)00105-G
- 102. Chakraborty S, Shukla D, Mishra B, et al. Lipid—an emerging platform for oral delivery of drugs with poor bioavailability. Eur J Pharm Biopharm. 2009;73(1):1-15. doi:

- 10.1016/j.ejpb.2009.06.001.
- 103. Charman WN, Porter CJH, Mithani S, et al. Physicochemical and physiological mechanisms for the effects of food on drug absorption: the role of lipids and pH. J Pharm Sci. 1997;86(3):269–82. DOI: 10.1021/js960085v
- 104. Crounse RG. Human pharmacology of griseofulvin: the effect of fat intake on gastrointestinal absorption. J Invest Dermatol. 1961; 37:529–33. DOI: 10.1038/jid.1961.154
- 105. Horter D, Dressman JB. Influence of physicochemical properties on dissolution of drugs in the gastrointestinal tract. Adv Drug Deliv Rev. 2001;46(1–3):75–87. DOI: 10.1016/s0169-409x(00)00130-7
- 106. Wagner D, Spahn-Langguth H, Hanafy A, et al. Intestinal drug efflux: formulation and food effects. Adv Drug Deliv Rev. 2001;50(1):S13–31. DOI: 10.1016/s0169-409x(01)00183-1
- 107. Touitou E, Barry BW, editors. Enhancement in drug delivery. Florida: CRC Press; 2006. 108. Liversidge GG, Cundy KC. Particle size reduction for improvement of oral bioavailability of hydrophobic drugs: I. Absolute oral bioavailability of nanocrystalline danazol in beagle dogs. Int J Pharm. 1995;125(1):91–7. doi.org/10.1016/0378-5173(95)00122-Y
- 109. Charman WN. Lipids, lipophilic drugs, and oral drug delivery—some emerging concepts. J Pharm Sci. 2000;89(8):967–78. DOI: 10.1002/1520-6017(200008)89:8<967::aid-jps1>3.0.co;2-r
- 110. Charman WN, Porter CJH, Mithani S, et al. Physicochemical and physiological mechanisms for the effects of food on drug absorption: the role of lipids and pH. J Pharm Sci. 1997;86(3):269–82. DOI: 10.1021/js960085v
- 111. Porter CJH, Charman WN. Intestinal lymphatic drug transport: an update. Adv Drug Deliv Rev. 2001;50(1–2):61–80. DOI: 10.1016/s0169-409x(01)00151-x
- 112. Charman SA, Charman WN, Rogge MC, et al. Self-emulsifying drug delivery systems: formulation and biopharmaceutic evaluation of an investigational lipophilic compound. Pharm Res. 1992; 9(1):87–93. DOI: 10.1023/a:1018987928936
- 113. Li C, Fleisher D, Li L, et al. Regional-dependent intestinal absorption and meal composition effects on systemic availability of LY303366, a lipopeptide antifungal agent, in dogs. J Pharm Sci. 2001; 90(1):47–57. DOI: 10.1002/1520-6017(200101)90:1<47::aid-jps6>3.0.co;2-2
- 114. Martinez M, Amidon G, Clarke L, et al. Applying the biopharmaceutics classification system to veterinary pharmaceutical products. Part II. Physiological considerations. Adv Drug Deliv Rev. 2002;54(6):825–50. DOI: 10.1016/s0169-409x(02)00071-6
- 115. Sanjula B, Shah FM, Javed A, et al. Effect of poloxamer 188 on lymphatic uptake of carvedilol-loaded solid lipid nanoparticles for bioavailability enhancement. J Drug Target. 2009;17(3):249-56. doi: 10.1080/10611860902718672.
- 116. Trevaskis NL, Charman WN, Porter CJ. Lipid-based delivery systems and intestinal lymphatic drug transport: a mechanistic update. Adv Drug Deliv Rev. 2008; 60(6):702–16. DOI: 10.1016/j.addr.2007.09.007
- 117. Khoo SM, Shackleford DM, Porter CJH, et al. Intestinal lymphatic transport of halofantrine occurs after oral administration of a unit–dose lipid-based formulation to fasted dogs. Pharm Res. 2003;20(9):1460–5. DOI: 10.1023/a:1025718513246
- 118. Wang W. Oral protein drug delivery. J Drug Target. 1996; 4(4):195-232. DOI: 10.3109/10611869608995624
- 119. Patel G, Misra A. Oral delivery of proteins and peptides: concepts and applications, in: M. Ambikanandan (Ed.), Challenges in Delivery of Therapeutic Genomics and Proteomics, Elsevier, London, 2011, pp. 481–529.

- 120. Woodley JF. Enzymatic barriers for GI peptide and protein delivery. Crit Rev Ther Drug Carrier Syst. 1994;11(2-3):61-95.
- 121. Langguth P, Bohner V, Heizmann J, et al. The challenge of proteolytic enzymes in intestinal peptide delivery. J Control Release. 46 (1997) 39–57. doi.org/10.1016/S0168-3659(96)01586-6
- 122. Mrsny R. Challenges for the oral delivery of proteins and peptides: theoretical and practical approaches to their delivery, in: Capsugel Symposia Series, Greenwood, SC, 1991, p. 452.
- 123. Ganong WF, Regulation of gastrointestinal function, in:W.F. Ganong (Ed.), Review of medical physiology, 12th ed., Lange Medical Publications, 1983, pp. 394–420.
- 124. Mrsny RJ. Challenges for the oral delivery of proteins and peptides: theoretical and practical approaches to their delivery, in: Capsugel Library Symposia series, 1991, pp. 45–52.
- 125. Hayakawa E, Lee VH. Aminopeptidase activity in the jejunal and ileal Peyer's patches of the albino rabbit. Pharm Res.1992;9:535–540. DOI: 10.1023/a:1015800615674
- 126. Skillman JJ, Gould SA, Chung RS, et al. The gastric mucosal barrier: clinical and experimental studies in critically ill and normal man, and in the rabbit. Ann Surg. 1970;172:564–584. doi: 10.1097/00000658-197010000-00004
- 127. Meyer RA, McGinley D, Posalaky Z. The gastric mucosal barrier: structure of intercellular junctions in the dog. J Ultrastruct Res. 1984;86:192–201. DOI: 10.1016/s0022-5320(84)80058-1
- 128. MacAdam A. The effect of gastro-intestinal mucus on drug absorption, Adv Drug Deliv Rev. 1993;11:201–220. doi.org/10.1016/0169-409X(93)90010-2
- 129. Phelps CF. Biosynthesis of mucus glycoprotein. Br Med Bull.1978;34:43–48.
- 130. Allen A, Garner A. Mucus and bicarbonate secretion in the stomach and their possible role in mucosal protection. Gut. 1980;21:249–262. doi: 10.1136/gut.21.3.249
- 131. Kompella UB, Lee VH. Delivery systems for penetration enhancement of peptide and protein drugs: design considerations. Adv Drug Deliv Rev. 2001;46:211–245. DOI: 10.1016/s0169-409x(00)00137-x
- 132. Kerss S, Allen A, Garner A. A simple method for measuring thickness of the mucus gel layer adherent to rat, frog and human gastric mucosa: influence of feeding, prostaglandin, N-acetylcysteine and other agents, Clin Sci. 1982;63:187–195. DOI: 10.1042/cs0630187
- 133. Pankhurst QA, Connolly J, Jones S, et al. Applications of magnetic nanoparticles in biomedicine. J Phys D Appl Phys. 2003;36:R167. DOI:10.1088/0022-3727/36/13/201
- 134. Kreuter J. Nanoparticulate systems in drug delivery and targeting. J Drug Target. 1995;3:171–173. DOI: 10.3109/10611869509015940
- 135. Jabir NR, Tabrez S, Ashraf GM, et al. Nanotechnology-based approaches in anticancer research. Int J Nanomedicine. 2012;7:4391-408. doi: 10.2147/IJN.S33838.
- 136. Li SD, Huang L. Pharmacokinetics and biodistribution of nanoparticles. Mol Pharm. 2008;5(4):496-504. doi: 10.1021/mp800049w.
- 137. Pattni BS, Chupin VV, Torchilin VP. New developments in liposomal drug delivery. Chem. Rev. 2015;115:10938–10966, http://dx.doi.org/10.1021/acs. chemrev.5b00046.
- 138. Sawant RR, Torchilin VP. Challenges in development of targeted liposomal therapeutics. AAPS J. 2012;14:303–315, http://dx.doi.org/10.1208/s12248-012-9330-0.
- 139. Vieira DB, Gamarra LF. Getting into the brain: liposome-based strategies for effective drug delivery across the blood brain barrier. Int. J. Nanomedicine. 2016;5381–5414. doi: 10.2147/JJN.S117210
- 140. Craparo EF, Bondì ML, Pitarresi G, et al. Nanoparticulate Systems for Drug Delivery and Targeting to the central nervous system. CNS Neurosci. Ther. 2011;17:670–677,

- http://dx.doi.org/10.1111/j.1755-5949. 2010.00199.x.
- 141. H. Yang. Nanoparticle-mediated brain-specific drug delivery, imaging, and diagnosis. Pharm. Res. 2010;27:1759–1771, http://dx.doi.org/10.1007/s11095-010-0141-7.
- 142. Chang HI, Yeh MK. Clinical development of liposome-based drugs: formulation, characterization, and therapeutic efficacy. Int. J. Nanomed. 2012;7:49–60. DOI: 10.2147/IJN.S26766
- 143. Chen W, Li H, Liu Z, et al. Lipopolyplex for therapeutic gene delivery and its application for the treatment of Parkinson's disease. Front. Aging Neurosci. 2016;8:1–10, http://dx.doi.org/10.3389/fnagi.2016.00068.
- 144. Huang H, Cruz W, Chen J, et al. Learning from biology: synthetic lipoproteins for drug delivery. Wiley Interdiscip. Rev. Nanomed. Nanobiotechnol. 2015;7:298–314, http://dx.doi.org/10.1002/wnan.1308.
- 145. Ag Seleci D, Seleci M, Walter JG, et al. Niosomes as nanoparticular drug carriers: fundamentals and recent applications. J. Nanomater. 2016 (2016), http://dx.doi.org/10.1155/2016/7372306.
- 146. Moghassemi S, Hadjizadeh A. Nano-niosomes as nanoscale drug delivery systems: an illustrated review. J. Control. Release. 2014;185:22–36. DOI: 10.1016/j.jconrel.2014.04.015 147. Chaurasia S, Dogra SS. European journal of Ejpmr Transfersomes: novel approach for intranasal delivery. Eur. J. Pharm. Rev. Artic. Eur. J. Pharm. Med. Res. Med. Res. 2017;4:192–203.
- 148. Ezzati Nazhad Dolatabadi J, Omidi Y. Solid lipid-based nanocarriers as efficient targeted drug and gene delivery systems. TrAC Trends Anal. Chem. 2016;77:100–108, http://dx.doi.org/10.1016/j.trac.2015.12.016.
- 149. Beloqui A, Solinís MA, Rodríguez-Gascón A, et al. Nanostructured lipid carriers: promising drug delivery systems for future clinics. Nanomedicine. 2016;12:143–161, http://dx.doi.org/10.1016/j.nano.2015.09.004.
- 150. Eldem T, Speiser P, Hincal A. Optimization of spray-dried and congealed lipid micropellets and characterization of their surface morphology by scanning electron microscopy. Pharm Res. 1991;8:47–54. DOI: 10.1023/a:1015874121860
- 151. Speiser P. Lipidnanopellets als Trägersystem für Arzneimittel zur peroralen Anwendung. European Patent EP 0167825, 1990.
- 152. Wei L, Yang Y, Shi K, et al. Preparation and characterization of loperamide-loaded dynasan 114 solid lipid nanoparticles for increased oral absorption in the treatment of diarrhea. Front Pharmacol. 2016;7:332. DOI: 10.3389/fphar.2016.00332
- 153. Subedi RK, Kang KW, Choi HK. Preparation and characterization of solid lipid nanoparticles loaded with doxorubicin. Eur J Pharm Sci. 2009;37(3-4):508-13. doi: 10.1016/j.eips.2009.04.008.
- 154. Müller RH, Runge SA. Solid lipid nanoparticles (SLN®) for controlled drug delivery, in: S. Benita (Ed.), Submicron Emulsions in Drug Targeting and Delivery. Harwood Academic Publishers, Amsterdam, 1998, pp. 219–234.
- 155. Üner M, Yener G. Importance of solid lipid nanoparticles (SLN) in various administration routes and future perspectives. Int J Nanomedicine. 2007;2(3):289-00.
- 156. Mukherjee S, Ray S, Thakur RS. Solid Lipid Nanoparticles: A Modern Formulation Approach in Drug Delivery System. Indian J Pharm Sci. 2009;71(4):349-58. doi: 10.4103/0250-474X.57282.
- 157. Müller RH, Mäder K, Gohla S. Solid lipid nanoparticles (SLN) for controlled drug delivery: a review of the state of the art. Eur J Pharm Biopharm. 2000;50: 161-77. DOI: 10.1016/s0939-6411(00)00087-4

- 158. Mehnert W, Mader K. Solid lipid nanoparticles: production, characterization and applications. Adv Drug Deliv Rev. 2001;47(2–3):165–196. DOI: 10.1016/s0169-409x(01)00105-3
- 159. Muller RH, Mader K, Gohla S. Solid lipid nanoparticles (SLN) for controlled drug delivery—a review of the state of the art. Eur J Pharm Biopharm. 2000;50(1):161–77. DOI: 10.1016/s0939-6411(00)00087-4
- 160. Muller RH, Mehnert W, Lucks JS, et al. Solid lipid nanoparticles (SLN)—an alternative colloidal carrier system for controlled drug delivery. Eur J Pharm Biopharm. 1995;41(1):62–9.
- 161. Zur Muhlen A, Mehnert W. Drug release and release mechanism of prednisolone loaded solid lipid nanoparticles. Pharmazie. 1998;53(8):552–5.
- 162. Souto EB, Müller RH. Lipid nanoparticles: effect on bioavailability and pharmacokinetic changes. In: Schäfer-Korting, M. (Ed.), Drug Delivery. Springer Berlin Heidelberg, Berlin, Heidelberg, 2010 pp. 115–141.
- 163. Shah R, Eldridge D, Palombo E, et al. Lipid Nanoparticles: Production, Characterization and Stability. Springer International Publishing, 2015, USA.
- 164. Gaba B, Fazil M, Ali A, et al. Nanostructured lipid (NLCs) carriers as a bioavailability enhancement tool for oral administration. Drug Deliv. 2015; 22(6):691-700. doi: 10.3109/10717544.2014.898110.
- 165. Shegokar R, Singh KK, Müller RH. Production & stability of stavudine solid lipid nanoparticles fromlab scale to industrial scale. Int. J. Pharm. 2011;416:461–470. DOI: 10.1016/j.ijpharm.2010.08.014
- 166. Yu YH, Kim E, Park DE, et al. Cationic solid lipid nanoparticles for co-delivery of paclitaxel and siRNA. Eur. J. Pharm. Biopharm. 2012;80:268–273. DOI: 10.1016/j.ejpb.2011.11.002
- 167. Xie S, Zhu L, Dong Z, et al. Preparation, characterization and pharmacokinetics of enrofloxacin-loaded solid lipid nanoparticles: influences of fatty acids. Colloids Surf. 2011;83:382–387. DOI: 10.1016/j.colsurfb.2010.12.014
- 168. Gastaldi L, Battaglia L, Peira E, et al. Solid lipid nanoparticles as vehicles of drugs to the brain: current state of the art. Eur. J. Pharm. Biopharm. 2014;87:433–444. DOI: 10.1016/j.ejpb.2014.05.004
- 169. Silva AC, González-Mira E, García ML, et al. Preparation, characterization and biocompatibility studies on risperidone-loaded solid lipid nanoparticles (SLN): high pressure homogenization versus ultrasound. Colloids Surf., 2011;86:158–165. DOI: 10.1016/j.colsurfb.2011.03.035
- 170. Dwivedi P, Khatik R, Khandelwal K, et al. Pharmacokinetics study of arteether loaded solid lipid nanoparticles: an improved oral bioavailability in rats. Int. J. Pharm. 2014;466:321–327. DOI: 10.1016/j.ijpharm.2014.03.036
- 171. Raza K, Singh B, Singal P, et al. Systematically optimized biocompatible isotretinoin-loaded solid lipid nanoparticles (SLNs) for topical treatment of acne. Colloids Surf., 2013;105:67–74. DOI: 10.1016/j.colsurfb.2012.12.043
- 172. Ravi PR, Vats R, Dalal V, et al. A hybrid design to optimize preparation of lopinavir loaded solid lipid nanoparticles and comparative pharmacokinetic evaluation with marketed lopinavir/ritonavir coformulation. J. Pharm. Pharmacol. 2014;66:912–926. DOI: 10.1111/jphp.12217
- 173. Kheradmandnia S, Vasheghani-Farahani E, Nosrati M, et al. Preparation and characterization of ketoprofen-loaded solid lipid nanoparticles made from beeswax and carnauba wax. Nanomedicine. 2010;6:753–759. DOI: 10.1016/j.nano.2010.06.003

- 174. Madan J, Pandey RS, Jain V, et al. Poly (ethylene)-glycol conjugated solid lipid nanoparticles of noscapine improve biological half-life, brain delivery and efficacy in glioblastoma cells. Nanomedicine. 2013;9:492–503. DOI: 10.1016/j.nano.2012.10.003
- 175. Mehnert W, Mäder K. Solid lipid nanoparticles: production, characterization and applications, Adv. Drug Deliv. Rev. 2001;47:165–196. DOI: 10.1016/s0169-409x(01)00105-3 176. Wang S, Chen T, Chen R, et al. Emodin loaded solid lipid nanoparticles: preparation, characterization and antitumor activity studies. Int. J. Pharm. 2012;430:238–246. DOI: 10.1016/j.ijpharm.2012.03.027
- 177. Kuo YC, Wang CC. Cationic solid lipid nanoparticles with primary and quaternary amines for release of saquinavir and biocompatibility with endothelia. Colloids Surf., 2013;101:101–105. DOI: 10.1016/j.colsurfb.2012.06.002
- 178. Cai S, Yang Q, Bagby TR, et al. Lymphatic drug delivery using engineered liposomes and solid lipid nanoparticles. Adv. Drug Deliv. Rev. 2011;63:901–908. DOI: 10.1016/j.addr.2011.05.017
- 179. Nafee N, Husari A, Maurer CK, et al. Antibiotic-free nanotherapeutics: ultra-small, mucus-penetrating solid lipid nanoparticles enhance the pulmonary delivery and anti-virulence efficacy of novel quorum sensing inhibitors. J. Control. Release. 2014;192:131–140. DOI: 10.1016/j.jconrel.2014.06.055
- 180. Almeida AJ, Souto E. Solid lipid nanoparticles as a drug delivery system for peptides and proteins. Adv. Drug Deliv. Rev. 2007;59:478–490. DOI: 10.1016/j.addr.2007.04.007
- 181. Shah RM, Malherbe F, Eldridge D, et al. Physicochemical characterization of solid lipid nanoparticles (SLNs) prepared by a novel microemulsion technique. J. Colloid Interface Sci. 2014;428:286–294. DOI: 10.1016/j.jcis.2014.04.057
- 182. Soares S, Fonte P, Costa A, et al. Effect of freeze-drying, cryoprotectants and storage conditions on the stability of secondary structure of insulin-loaded solid lipid nanoparticles. Int. J. Pharm. 2013;456:370–381. DOI: 10.1016/j.ijpharm.2013.08.076
- 183. Kalhapure RS, Mocktar C, Sikwal DR, et al. Ion pairing with linoleic acid simultaneously enhances encapsulation efficiency and antibacterial activity of vancomycin in solid lipid nanoparticles. Colloids Surf., 2014;117:303–311. DOI: 10.1016/j.colsurfb.2014.02.045
- 184. Hao J, Wang F, Wang X, et al. Development and optimization of baicalin-loaded solid lipid nanoparticles prepared by coacervationmethod using central composite design. Eur. J. Pharm. Sci. 2012;47:497–505. DOI: 10.1016/j.ejps.2012.07.006
- 185. Pardeshi C, Rajput P, Belgamwar V, et al. Solid lipid based nanocarriers: an overview/Nanonosači na bazi čvrstih lipida: pregled. Acta Pharm. 2012; 62(4):433-72. doi: 10.2478/v10007-012-0040-z.
- 186. Muchow M, Maincent P, Müller RH. Lipid nanoparticles with a solid matrix (SLN®, NLC®, LDC®) for oral drug delivery. Drug Dev Ind Pharm. 2008;34(12):1394-405. doi: 10.1080/03639040802130061.
- 187. Muèller RH, Maèder K, Gohla S. Solid lipid nanoparticles (SLN) for controlled drug delivery—a review of the state of the art. Eur J Pharm Biopharm. 2000;50: 161–177. DOI: 10.1016/s0939-6411(00)00087-4
- 188. Borgström B. Importance of phospholipids, pancreatic phospholipase A2, and fatty acid for the digestion of dietary fat. Gastroenterology, 1980;78:954–962.
- 189. Borgström B, Donner J. The polar interactions between pancreatic lipase, colipase and the triglyceride substrate. FEBS Lett. 1977;83:23–26.
- 190. Scow RO, Olivecorona T. Effect of albumin products formed from chylomicron triacylglycerol by lipoprotein lipase in vitro. Biochem Biophys Acta. 1977;487: 472–486. DOI: 10.1016/0005-2760(77)90217-x

- 191. Olbrich C, Müller RH. Enzymatic degradation of SLN effect of surfactant and surfactant mixtures. Int J Pharm. 1999;180:31–39. DOI: 10.1016/s0378-5173(98)00404-9
- 192. Müller RH, Olbrich C. Solid lipid nanoparticles: phagocytic uptake, in vitro cytotoxicity and in vitro biodegradation, 2nd communication. Pharm Ind. 1999;61:564–569.
- 193. Olbrich C, Mehnert W, Müller RH. In vitro degradation properties of solid lipid nanoparticles, in: Proc. 2nd World Meeting APGI/APV, Paris, 1998, pp. 577–578.
- 194. Müller RH, Rühl D, Runge SA. Biodegradation of solid lipid nanoparticles as a function of lipase incubation time. Int J Pharm. 1996;144:115–121.
- 195. Olbrich C, Mehnert W, Müller RH. In vitro degradation properties of solid lipid nanoparticles, in: Proc. 2nd World Meeting APGI/APV, Paris, 1998, pp. 627–628.
- 196. Yang S, Zhu J, Lu Y, et al. Yang, Body distribution of camptothecin solid lipid nanoparticles after oral administration. Pharm Res. 1999;16:751–757. doi.org/10.1023/A:1018888927852
- 197. Penkler L, Müller RH, Runge SA. Pharmaceutical cyclosporin formulation with improved biopharmaceutical properties, improved physical quality and greater stability, and method for producing said formulation, WO 99/56733, 1999.
- 198. Bargoni A, Cavalli R, Caputo O. Solid lipid nanoparticles in lymph and plasma after duodenal administration to rats, Pharm Res. 1998;15:745–75. doi: 10.1023/A:1011975120776 199. Mosmann T. Rapid colorimetric assay of cellular growth and survival: application to proliferation and cytotoxicity assays. J Immunol Meth. 1983;65:55–63. DOI: 10.1016/0022-1759(83)90303-4
- 200. Müller RH, Maaßen S, Weyhers H. Cytotoxicity of magnetite-loaded polylactide, polylactide/glycolide particles and solid lipid nanoparticles. Int J Pharm. 1996;138:85–94. doi.org/10.1016/0378-5173(96)04539-5
- 201. Marcato PD, Duran N. Cytotoxicity and Genotoxicity of Solid Lipid Nanoparticles. In: Duran N, Guterres SS, Alves OL, Eds. Nanotoxicology: Materials, Methodologies, and Assessments. New York, NY: Springer New York 2014; pp. 229-44.
- 202. Almeida H, Amaral MH, Lobao P. Applications of polymeric and lipid nanoparticles in ophthalmic pharmaceutical formulations: Present and future considerations. J Pharm Pharm Sci. 2014;17(3):278-93. DOI:10.18433/J3DP43
- 203. Muller RH, Mader K, Gohla S. Solid lipid nanoparticles (SLN) for controlled drug delivery: a review of the state of the art. Eur J Pharm Biopharm. 2000;50:161-77. DOI: 10.1016/s0939-6411(00)00087-4
- 204. Doktorovova S, Souto EB, Silva AM. Nanotoxicology applied to solid lipid nanoparticles and nanostructured lipid carriers a systematic review of in vitro data. Eur J Pharm Biopharm. 2014;87(1):1-18. doi: 10.1016/j.ejpb.2014.02.005.
- 205. Fortunov S, Peneva P, Andonova V, et al. Toxicity assessment of drug delivery nanocarriers. Science & Technologies 2016; VI(1): 262-6.
- 206. Hillegass J, Shukla A, Lathrop S, et al. Assessing nanotoxicity in cells in vitro. Wiley Interdiscip Rev Nanomed Nanobiotechnol. 2010;2(3):219-231. DOI: 10.1002/wnan.54.
- 207. Maaßen S, Schwarz C, Mehnert W. Comparison of cytotoxicity between polyester nanoparticles and solid lipid nanoparticles. Proc Intern Symp Control Rel Bioact Mater. 1993;20:490–491.
- 208. Müller RH, Maaßen S, Weyhers H, et al. Phagozytic uptake and cytotoxicity of solid lipid nanoparticles (SLN) sterically stabilized with poloxamine 908 and poloxamer 407. J Drug Target. 1996;4(3):161-70. DOI: 10.3109/10611869609015973
- 209. Yu YH, Kim E, Park DE, et al. Cationic solid lipid nanoparticles for co-delivery of paclitaxel and siRNA. Eur. J. Pharm. Biopharm. 2012;80:268–273. DOI:

- 10.1016/j.ejpb.2011.11.002
- 210. Shi S, Han L, Deng L, et al. Dual drugs (microRNA-34a and paclitaxel)-loaded functional solid lipid nanoparticles for synergistic cancer cell suppression. J. Control. Release. 2014;194:228–237. DOI: 10.1016/j.jconrel.2014.09.005
- 211. Jin J, Bae KH, Yang H, et al. In vivo specific delivery of c-Met siRNA to glioblastoma using cationic solid lipid nanoparticles. Bioconjug. Chem. 2011;22:2568–2572. DOI: 10.1021/bc200406n
- 212. Baek JS, Cho CW. Controlled release and reversal of multidrug resistance by coencapsulation of paclitaxel and verapamil in solid lipid nanoparticles. Int. J. Pharm. 2015;478:617–624. DOI: 10.1016/j.ijpharm.2014.12.018
- 213. Kalhapure RS, Mocktar C, Sikwal DR, et al. Ion pairing with linoleic acid simultaneously enhances encapsulation efficiency and antibacterial activity of vancomycin in solid lipid nanoparticles. Colloids Surf. 2014;117:303–311. DOI: 10.1016/j.colsurfb.2014.02.045
- 214. Morishita M, Morishita I, Takayama K, et al. Site-dependent effect of aprotinin, sodium caprate, Na2EDTA and sodium glycocholate on intestinal absorption of insulin. Biol. Pharm. Bull. 1993;16(1):68–72. DOI: 10.1248/bpb.16.68
- 215. Lane ME, O'Driscoll CM, Corrigan OI. Quantitative estimation of the effects of bile salt surfactant systems on insulin stability and permeability in the rat intestine using a mass balance model. J. Pharm. Pharmacol. 2005;57(2):169–175. DOI: 10.1211/0022357055434
- 216. Sharma G, Wilson K, Van Der Walle CF, et al. Microemulsions for oral delivery of insulin: design, development and evaluation in streptozotocin induced diabetic rats. Eur. J. Pharm. Biopharm. 2010;76(2):159–169. DOI: 10.1016/j.ejpb.2010.07.002
- 217. Cilek A, Celebi N, Tirnaksiz F, et al. A lecithin-based microemulsion of rh-insulin with aprotinin for oral administration: investigation of hypoglycemic effects in non-diabetic and STZ-induced diabetic rats. Int. J. Pharm. 2005;298(1):176–185. DOI: 10.1016/j.ijpharm.2005.04.016
- 218. Elsayed A, Remawi MA, Qinna N, et al. Formulation and characterization of an oily-based system for oral delivery of insulin. Eur. J. Pharm. Biopharm. 2009;73(2):269–279. DOI: 10.1016/j.ejpb.2009.06.004
- 219. Toorisaka E, Ono H, Arimori K, et al. Hypoglycemic effect of surfactantcoated insulin solubilized in a novel solid-in-oil-in-water (S/O/W) emulsion. Int. J. Pharm. 2003;252(1–2):271–274. DOI: 10.1016/s0378-5173(02)00674-9
- 220. Watnasirichaikul S, Rades T, Tucker IG, et al. In vitro release and oral bioactivity of insulin in diabetic rats using nanocapsules dispersed in biocompatible microemulsion. J. Pharm. Pharmacol. 2002;54(4):473–480. DOI: 10.1211/0022357021778736
- 221. Constantinides PP, Scalart JP, Lancaster C, et al. Formulation and intestinal-absorption enhancement evaluation of water-in-oil microemulsions incorporating medium-chain glycerides. Pharm. Res. 1994;11(10): 1385–1390. DOI: 10.1023/a:1018927402875
- 222. Ritschel WA. Microemulsions for improved peptide absorption from the gastrointestinal tract. Methods Find. Exp. Clin. Pharmacol. 1991;13(3):205–220.
- 223. Celebi N, Turkyilmaz A, Gonul B, et al. Effects of epidermal growth factor microemulsion formulation on the healing of stress-induced gastric ulcers in rats. J. Control. Release. 2002;83(2):197–210. DOI: 10.1016/s0168-3659(02)00198-0
- 224. Rao SVR, Yajurvedi K, Shao J. Selfnanoemulsifying drug delivery system(SNEDDS) for oral delivery of protein drugs III. In vivo oral absorption study. Int. J. Pharm. 2008;362(1–2):16–19. DOI: 10.1016/j.ijpharm.2008.05.015
- 225. Lyons KC, Charman WN, Miller R, et al. Factors limiting the oral bioavailability of N-acetylglucosaminyl-N-acetylmuramyl dipeptide (GMDP) and enhancement of absorption in

- rats by delivery in a water-in-oil microemulsion. Int. J. Pharm. 2000;199(1):17–28. DOI: 10.1016/s0378-5173(00)00349-5
- 226. Zheng JY, Fulu MY. Decrease of genital organ weights and plasma testosterone levels in rats following oral administration of leuprolide microemulsion. Int. J. Pharm. 2006;307(2):209–215. DOI: 10.1016/j.ijpharm.2005.10.007
- 227. Fagerholm U, Sjostrom B, Sroka-Markovic J, et al. The effect of a drug-delivery system consisting of soybean phosphatidyl choline and medium-chain monoacylglycerol on the intestinal permeability of hexarelin in the rat. The J. Pharm. Pharmacol. 1998;50(5):467–473. DOI: 10.1111/j.2042-7158.1998.tb06187.x
- 228. Aungst BJ, Saitoh H, Burcham DL, et al. Enhancement of the intestinal absorption of peptides and nonpeptides. J. Control. Release. 1996;41(1–2):19–31. https://doi.org/10.1016/0168-3659(96)01353-3
- 229. Hastewell J, Lynch S, Williamson I, et al. Absorption of human calcitonin across the rat colon in vivo. Clin. Sci. 1992;82(5):589–594. DOI: 10.1042/cs0820589
- 230. New R, Littlewood G, Guard P, et al. Intestinal delivery of calcitonin in pig. Int. J. Pharm. 1997;156(1):1–8.
- 231. Fan Y, Li X, Zhou Y, et al. Improved intestinal delivery of salmon calcitonin by water-in-oil microemulsions. Int. J. Pharm. 2011;416(1):323–330. DOI: 10.1016/j.ijpharm.2011.06.029
- 232. Yoshiura H, Tahara Y, Hashida M, et al. Design and in vivo evaluation of solid-in-oil suspension for oral delivery of human growth hormone. Biochem. Eng. J. 2008;41(2):106–110. 10.1016/j.bej.2008.04.001
- 233. FDA. 21 CFR Part 182, Substances Generally Recognized as Safe. 2010.
- 234. U.S. Department of Health and Human Services, Food and Drug Administration. Guidance for industry: non clinical studies for the safety evaluation of pharmaceutical excipients. Office of Training and Communication, Division of Drug Information, HFD-240, Center for Drug Evaluation and Research, Food and Drug Administration, or Office of Communication, Training, and Manufacturers Assistance, HFM-40, Center for Biologics Evaluation and Research, Food and Drug Administration; May 2005. Available from: (http://www.fda.gov/cder/guidance/5544fnl.pdf).
- 235. U.S. Food and Drug Administration. Inactive ingredients search for approved drug products. Division of Labeling and Program Support, Office of Generic Drugs, Center for Drug Evaluation and Research, Food and Drug Administration; 2007. Available from: (http://www.accessdata.fda.gov/scripts/cder/iig/index.cfm) [accessed 29.07.07].
- 236. P. Maincent, "The regulatory environment: the challenges for lipid-based formulations," Bulletin Technique Gattefoss'e, vol. 100, pp. 47–49, 2007.
- 237. Chen M, "Lipid excipients and delivery systems for pharmaceutical development: a regulatory perspective," Advanced Drug Delivery Reviews, vol. 60, no. 6, pp. 768–777, 2008.