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Responsive Nano-structured Cubosomes: Advancements and Therapeutic Applications

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ABSTRACT

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Submitted: June 25, 2024 Revised: May 30, 2025 Accepted: May 30, 2025 ePublished: May 31, 2025 Nanotechnology has revolutionized drug delivery, which offers innovative ways to maximize treatment efficacy while decreasing side effects. The lyotropic liquid crystalline nanoparticles, such as cubosomes and hexosomes, have gained substantial interest because of their distinctive molecular arrangements. Lipophilic, hydrophilic, and amphiphilic drugs can be encapsulated by cubosomes, making them versatile carriers in drug delivery systems. Different types of cubosomes, such as pH-responsive, temperature-responsive, light-responsive, enzyme-responsive, and multi-stimuli-responsive, have been discussed in this review detailing their preparation methods and therapeutic applications. Cubosomes possess high surface area, are biocompatible, and provide enhanced drug protection. However, formulation stability and scalability are the main challenges. This paper highlights the potential of cubosomes for targeted drug delivery, focusing on their ability to optimize bioavailability and controlled drug release.

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Introduction

Nanotechnology has developed drug delivery by providing novel solutions that improve treatment outcomes while reducing adverse effects. Active pharmaceutical ingredients (API) characterized by extremely low solubility present restricted bioavailability upon oral ingestion, diminished diffusion capacity via the outer membrane, the necessity for higher quantities during intravenous administration, and undesired side effects preceding the conventional formulation methods.¹ Integrating nanotechnology in drug delivery mechanisms offers a pathway to overcome these constraints. Lyotropic liquid crystalline nanoparticles (LLCNP) emerge as exemplars of self-assembling nanomaterials, showcasing remarkable potential.^{2–5} Among the array of nanostructured carriers, cubosomes epitomize a burgeoning class of nanosystems engineered to accommodate various active pharmaceutical ingredients, consisting of both hydrophobic and hydrophilic drugs, alongside biotherapeutics such as peptides, proteins, and nucleic acids.^{6–9}

The process of liquid crystalline injectable formulations involves integrating amphiphilic molecules within a solvent, resulting in the formation of LLC phases. The amphiphilic structure of the molecule, additives, and the solution's conditions affect these phases.¹⁰ Friedrich Reinitzer made the first observation of LLCNP in 1999.¹¹ Like liposomes, they have intricate 2D and 3D nonlamellar nanostructures, including inverse hexagonal and cubic mesophases. The distinctions between liposomes and cubosomes are illustrated in Figure **1**.

Cubosomes are square and rounded shapes and possess an internal cubic lattice. They are thermodynamically stable structures characterized by honeycombed SL formulations that create two internal aqueous channels and a substantial interfacial area.^{12,13} Inside the bicontinuous cubic stage framework, there are three stages [Im3m (Schwarz) surface), Pn3m (Diamond surface), Ia3d (gyro surface)] to explain the distinct shapes, all of which additionally display boosted drug transportation in the target site. ^{14–17} According to Ayesha Waheed, the organized mesophase structure and nanoscale size range of liquid crystalline NPs make them useful drug carriers for molecules with different polarity, such as nucleic acids and proteins. A thorough analysis highlights how their 3D structure and tunable coronas support a variety of applications, from theranostics to medication delivery.¹¹

Zhai et al. reflected that liquid crystalline drug delivery systems are promising for the future generation of nanomedicine, with their self-assembling amphiphilic lipids acting as efficient nanocarriers for a variety of medications, peptides, proteins, nucleic acids, and imaging agents.¹⁸

The unique cubic structure not only presents a platform for superior drug encapsulation and protection but also offers tailored solutions for targeted therapeutic interventions.^{19–21} Cubosomes possess high surface area and responsiveness to stimuli. Using this, researchers can design and tailor sophisticated delivery systems capable of precise control over drug release and distribution within the body. ^{22,23} Moreover, the diversity of types of cubosome each with its own set of advantages and challenges, underscores the necessity of a comprehensive exploitation. We intend to inspire further research and innovation in the nanotechnology field, driving advancements in therapeutic interventions, and improving patient care.

pH-Responsive Cubosomes

There are pH differences between normal blood and pathological tissues (e.g., those affected by infection, inflammation, and cancer, which often become more acidic), among specific intracellular compartments such as the cytosol, endosomes, and lysosomes, and along the gastrointestinal tract. These differences are typically targeted by pH-responsive nanosystems. For this scope, "smart" molecules including polymers, lipids, and peptides are used since they are biocompatible and sensitive to specific pH levels because of their functional ionizable groups. ^{24,25}

The pH-responsive mesophase, designed with a lipid-based LLC system composed of mono-linolein and pyridinyl methyl linoleate, transitions its symmetry from a reverse hexagonal phase (H₂) at pH 7.4 to a bi-continuous cubic phase (pn3m) at pH 5.5. This transformation was studied and found to occur due to the protonation of the pyridinyl methyl linoleate's weakly basic head group near its pKa of 5.5. ²⁶ This pH-triggered behavior leverages the acidic conditions in tumor tissues to enhance the release of drugs like Doxorubicin, potentially improving chemotherapy efficacy. By switching its structure under acidic conditions, this mesophase enables targeted drug delivery, improving the efficiency of cancer treatments. ^{27,28}

According to Rajesh et al., pH-responsive cubosomes lessen side effects while allowing a chemotherapeutic chemical to be delivered to tumors selectively. ²⁹ According to Mertins et al., pH-sensitive polymer shells present novel prospects for topical and oral medication delivery that could lead to the development of innovative cancer treatments. Drug compounds that have electrochemical activity might also be advantageous for pH-responsive drug release. ³⁰ At physiological pH, these NPs had a slow-releasing hexagonal structure, whereas, at the acidic pH of the tumor, they displayed a quick-releasing bi-continuous cubic phase. ³¹

According to Manchun et al., pH-sensitive nanosystems have been synthesized to deliver medications to the endosomes or lysosomes within cancer cells, or the mildly acidic extracellular fluids of tumor tissue following endocytosis. After the medication accumulates in tumor tissue via enhanced permeability and retention effect, these systems can release it through specialized mechanisms. Alternatively, they can release the drug within endosomes and lysosomes via pH-controlled hydrolysis after cellular uptake through the endocytic pathway. ³² According to Negrini et al., linoleic acid, a weak acid with a pKa of approximately 5, provides pH responsiveness. At pH 7, it is essentially in the deprotonated charged state, while at pH 2, it is primarily protonated and neutral. This results in changes to the critical packing parameter of the LLC. ³³ In the study by Prajapati et al. pH-responsive cubosomes were synthesized by blending 2-Hydoxyoleic acid with glycerol monooleate at varying mass ratios to examine pH-induced structural transformation for targeted drug delivery to cancer tissues. The research aimed to investigate the composition and pH dependence of drug-loaded NPs, providing insights into their pH-triggered transformation [Figure 2.]. ³⁴

Temperature Responsive Cubosomes

Numerous investigations have corroborated the hypothesis that drug release from thermos-responsive polymers can be triggered by slight temperature variations. However, in recent years, thermos-responsive drug delivery systems have primarily been utilized in the delivery of anti-cancer drugs and imaging agents.³⁵ Poloxamers stand out as widely explores thermo-responsive systems, renowned for their versatility. Thermo-responsive systems offer numerous advantages, such as the formulation of an *in-situ* gelling-controlled release system.³⁶ Mohsen et al. elucidated the significance of a thermos-sensitive system in enhancing the intranasal delivery of Lamotrigine. By integrating optimized cubosomes into a thermosensitive in *situ gel*,the system enhances the physical stability, nasal residence time, and patient compliance. The thermos-sensitive gel undergoes a sol-gel transition in response to temperature changes, facilitating the administration as a liquid that transforms into a gel upon contact with the nasal mucosa. The transition augments drug absorption across the nasal mucosa membrane, thereby improving the efficacy of lamotrigine in epilepsy treatment. The thermosensitive property is attributed to the *in-situ* gel into which the optimized cubosomes are incorporated. The synergistic combination of cubosomes and the thermosensitive gel enhances drug absorption and efficacy by prolonging nasal residence time and facilitating drug release at the target site.³⁷

According to Dabkowska et al., the tiny poly(*N*-isopropylacrylamide) nanogels act as precise thermos-responsive controllers in regulating the hydration of liquid crystalline surface layers.

Their rapid transition from a swollen to a collapsed state, induced by temperature changes, allows controlled release of water from the surface while preserving the integrity of the lipid matrix. This capability enables the secure encapsulation of delicate bioactive molecules within the lipid matrix, presenting a promising avenue for controlled-release applications. ³⁸

Light responsive cubosome

Light-responsive nanocarriers offer a non-invasive, highly adaptable, and precisely controlled method for drug delivery. ³⁹ The advancement of stimuli-responsive materials is crucial, marking the initial utilization of photoswitchable amphiphiles for the creation of light-sensitive cubic LLC dispersions, known as cubosomes. This innovation enables external manipulation of the LLC structure, facilitating the on-demand release of entrapped guest molecules. To produce these cubosomes, azobenzene photo surfactants, which have an azobenzene-alkyl tail and a neutral tetra ethylene glycol head group, are used with monoolein-water systems. ⁴⁰ The lipid/water system can be made photo-responsive by including plasmonic NPs or photochromic compounds. When these components are activated, the lipid bilayer's permeability changes either temporarily or completely, enabling or prohibiting the movement of molecules that are encapsulated [Figure 3.]. By using photothermal and photochromic techniques, this has been accomplished.⁴¹

Chen et al. highlighted that phototherapy, which includes photothermal therapy and photodynamic therapy, has attracted considerable interest among researchers because of its non-invasive nature, precise spatial and temporal selectivity, and low toxicity.⁴² As per Fong et al., plasmonic hydrophobized gold nanorods are integrated into mesophases produced by diverse lipid/water combinations to fabricate light-responsive bulk self-assembly lipid systems. The gold nanorods nestled within the liquid crystalline matrix were stimulated by Near Infrared laser light, inducing localized plasmonic heating of the mesophase. This facilitated the reversible manipulation of nanostructure, contingent upon the concentration of nanorods and the composition and heat capacity of the liquid crystalline matrix. ⁴¹

According to Angelova et al. in host-guest LLC mesophases of lipids intended as molecular switches for the "ondemand" release of chemicals, Small-Angle X-Ray Scattering (SAXS) has shown light-triggered effects. A tiny quantity of a lipid with an azobenzene photoactive unit that was synthesized successfully has been added to host liquid crystalline mesophases made up of oleic acid and monoolein. ⁴³ According to Jia et al., a promising class of photo-switchable molecules that hasn't gotten much attention in the literature is hexaaryl biimidazoles. The one that exhibits remarkable negative photochromism and is sensitive to green light is (2,20-dimethoxy diphenyl imidazole)-1,10-binaphthyl.⁴⁴ The photosensitive cubosomes utilized in the study by Bazylin´ ska et al represent a substantial advancement in targeted therapy for melanoma skin cancer cells. These cubosomes, laden with photosensitizing dyes such as Chlorin e6 or meso- tetraphenylporphine-Mn (III) chloride, exhibited enhanced biocompatibility, heightened therapeutic efficacy with significant cytotoxic effects post-irradiation effective bioimaging capabilities, and potential for future applications in photodynamic therapy and bioimaging of skin malignant melanoma. The innovative approach presents a promising avenue for precise and efficient treatment of melanoma skin cancer, underscoring the potential of cubosomes are versatile drug delivery systems for improved therapeutic outcomes.⁴⁵

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Enzyme-responsive cubosomes

One of the most promising types of smart stimulus-responsive NPs is enzyme-responsive NPs. Enzymes are needed for the body's lipid processing to transform lipids into cellular fuel ⁴¹. The hepatoprotective properties and oral bioavailability of Coenzyme Q10 can be significantly enhanced by utilizing Glycerol Monooleate (GMO) cubosomes stabilized with P407. To overcome the challenges associated with piperine distribution, such as hydrophobicity and 1st pass metabolism, tween 80-modified GMO cubosomes have been developed. *In vivo* studies indicate that these cubosomes markedly amplify cognitive function, suggesting their potential as a noninvasive, brain-targeted delivery system for Alzheimer's disease treatment. Similarly, the incorporation of curcumin into phytantriol cubosomes has demonstrated a substantial increase in oral bioavailability, achieving at least a 14-fold improvement compared to free curcumin. ⁴⁶

According to Tan et al. cubosomes containing the antimicrobial peptide were found to have a notably greater bactericidal impact following enzyme exposure than pure peptide which lost its bactericidal action upon proteolysis⁴⁶. The significance of enzyme-sensitive cubosomes lies in their potential application as drug-delivery systems that can respond to specific enzymatic triggers. In the presence of enzymes present in the target tissue, the cubosomes can be synthesized for releasing payloads in a controlled fashion. This targeted and triggered release mechanism enhances the efficiency and specificity of drug delivery, especially in customized treatments of various diseases, hence offering precise dosage and minimal off-target effects. Fusion of enzyme-sensitive molecules and cubosomes can develop smart nanocarriers that react to specific biological cues such as enzyme activity levels in diseased tissues. ^{47,48}

Based on the study conducted by Li et al, using enzymes as triggers, chemical selectivity and substrate specificity can be achieved. Enzyme-catalyzed reactions can be performed under moderate conditions, low-temperature aqueous environments, and neutral or near-neutral pH levels. Phospholipase exhibited high selectivity hydrolyzing fatty ester bonds at the sn-2 position of glycerophospholipids. Such cubosomes also increased the efficacy of urokinase-type plasminogen activator, a thrombolytic drug. Its encapsulation within the enzyme-responsive cubosomes aid in targeted drug release realized due to specific enzymatic triggers within the thrombus microenvironment. This ensures the thrombolytic agents retain protection while in circulation and are precisely released at the thrombosis site. ⁴⁹

Multi stimuli responsive cubosomes

Dual and multi-responsive, including stimuli-responsive NPs, are the innovative drug delivery strategies developed for combinational chemo-phototherapy. Integrating multiple stimuli - such as pH and redox, pH and temperature, temperature and magnetic field, enzyme activity, and others- resulted in multi-responsive drug delivery systems. For example Co-loading photosensitizer and chemotherapeutic agents onto graphene oxide NPs has shown a marked improvement in cancer treatment efficacy compared to monotherapy.⁵⁰ For targeted photodynamic treatment, a pH-responsive nanophotomedicine (pH-NanoPM) was developed. This nano photomedicine was constructed through the self-assembly of a pH-responsive polymeric photosensitizer (pH-PPS), incorporating approximately 10nm-sized pH-cleavable mPEG (pH-pH-mPEG). When HeLa human cervical cancer cells were exposed to pH-NanoPM, enhanced cellular internalization was observed at the acidic tumor pH compared to the normal pH, leading to a significant increase in cancer cell cytotoxicity. The fusion of metal NPs and stimuli-responsive polymers onto one platform has garnered a lot of attention in recent years. Zhou et al. stated that a drug in combination with a polymer containing selenium may be employed successfully for

multi-stimuli responsive drug release. They created metal-organic frameworks with pH-triggered properties for drug delivery systems and selenium-containing PEG micelles with redox-triggered features. It is observed that the shell can only break down in low pH conditions, the cores collapsed readily in the presence of redox agents. ⁵¹ As reported by Sauraj et al. in their study on pH-sensitive prodrug NPs for targeted chemo-photodynamic therapy, the integrated platform was formed by encapsulating the photosensitizer after connecting the chemotherapeutic agent DOX to the polymer PEG via a pH-sensitive (Schiff base) bond. Under acidic pH conditions, the NPs exhibited pH-responsive release behavior, leading to the simultaneous release of the medication and photosensitizer. An *in vivo* investigation revealed that NPs had higher antitumor efficacy against the cells when compared to free drugs and photosensitizers. ⁵⁰ Details on types of cubosomes and their potential applications are given in Table 1.

Types of responsive cubosomes	Composition	Drug used	Applications	References
pH-responsive	Monoolein Pluronic F127 Ionizable amino lipids	Doxorubicin	Anticancer Therapy	Rajesh et al. ⁵²
Light-responsive cubosomes	Photoswitchable amphiphiles such as azobenzene photo surfactants and monoolein	Nile Red	Controlled drug delivery.	Jones et al. ⁵³
pH-responsive cubosomes	Monoolein and the amino lipids N- (Pyridin-4-ylmethyl) oleamide and N- (2(piperidine- 1yl)ethyl) oleamide	7-ethyl-10-hydroxy camptothecin, which is an active metabolite of the anticancer prodrug irinotecan	Anticancer Therapy	Rajesh et al. ⁵⁴
pH-responsive cubosomes	Monoolein N-arginine-modified chitosan and alginate	Anthelmintic drugs including ivermectin, mebendazole, and praziquantel 1.	oral drug delivery systems	Mathews et al.
Cubosomes in thermos responsive gelling system	Glyceryl monooleate, Pluronic® F127,	Docetaxel	Controlled- release	Rarokar et al. 56
Thermosensitive cubosomes	Poloxamer 407 Glyceryl monooleate	Lamotrigine	intranasal delivery	Mohsen et al.
Thermosensitive cubosomes	poly(N,N-dimethyl acrylamide)-block- poly(N-isopropyl acrylamide) glycerol-monooleate	-	targeted drug delivery	Balestri et al.
pH-responsive cubosomes	Monoolein. Brucea javanica oil	Doxorubicin	combined delivery for cancer treatment.	Li et al. ⁵⁹

Table 1. Types of responsive cubosomes and their potential applications

Top-down and bottom-up approaches for cubosomes preparation

In the bottom-up approach, cubosomes are fashioned through the dispersion of droplets from the inverse micellar phase into an aqueous medium heated to 80°C. subsequently, a gradual cooling process ensues, prompting crystallization and the emergence of cubosomes. The hydrotrope incorporated within the cubosomes formulation assumes a pivotal role in thwarting the development of a bulk cubic gel phase. Its action involves a dissolution of

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the cubic gel, while the subsequent introduction of water, in conjunction with sonication, diminishes the solubility of the liquid crystalline particles, thus fostering the genesis of cubic entities. ⁶⁰

According to Gaballa et al., poloxamer 407 and soulan C24 were used as stabilizers throughout the top-down process of creating GMO cubosomes. ⁶¹ As per Garg et al., the surfactant employed in the production of cubosomes includes poloxamer 407, with the concentration ranging from 0%-20% w/w about the dispersion phase. Typically, a concentration of 2.5%-10% w/w of the total weight of the dispersion is necessary for the monoglyceride/surfactant mixture. In addition to poloxamer, polyvinyl alcohol (PVA) is utilized as a dispersion stabilizer. ⁶² According to Bryant et al. phytantriol solutions in a variety of diluents, such as glycerol, ethanol, honey, lactic acid, and choline chloride-glycerol, were used to create cubosomes. Applying these solutions dropwise to water containing poloxamer 407 stabilizers was done following a well-established cubosome synthesis protocol. ⁶³ According to Gaballa et al. to avoid cubosome dispersion aggregation, F127 or another appropriate stabilizer must be used. The selection of the ideal preparation technique still focuses primarily on stability, biocompatibility, and optimal drug release. ^{64,65}

Challenges and considerations

Addressing the potential challenges inherent in cubosomes production and stability is paramount to fully harnessing their potential in pharmaceutical applications. A significant obstacle is the elevated viscosity of the cubic phase, which complicates large-scale production processes. Moreover, cubosomes tend to exhibit low entrapment efficiency for water-soluble drug molecules due to the significant water content within their structure. This constraint not only has an impact on overall medication loading capacity, but it also impairs the delivery system's effectiveness. Nano-sized nature of cubosomes can undergo particle growth upon prolonged standing and is problematic for parenteral formulations. This causes stability and uniformity issues, demands innovative approaches to mitigate particle proliferation, and assures long-term stability. ^{24,66-68}

There is continuous research on cubosomes in drug delivery aimed at surmounting production limitations and augmenting stability. Researchers hope to unlock the complete potential of cubosomes by tackling difficulties associated with cubosomes such as viscosity maintenance during the synthesis and refinement of drug-loading methods for hydrophilic compounds. The NP engineering and tailored formulations promise to mitigate stability concerns connected to particle growth, thereby enhancing the suitability of cubosomes for various administration routes including parenteral delivery. The challenges can be resolved with interdisciplinary research efforts, shaping the future landscape for customized drug delivery options. ^{23,69–71}

Diverse utilization

Cubosomes have emerged as a promising drug delivery system, presenting advantages over traditional liposomes because of their unique inner cubic structure. This unique configuration provides a significantly larger interfacial surface area, facilitating the encapsulation and protection of higher quantities of hydrophilic and hydrophobic drugs compared to liposomes ^{72–74}. Notably, cubosome preparation primarily employs shear and homogenization techniques, eliminating the need for organic solvents. Furthermore, cubosomes exhibit superior solubilization capacities in contrast to conventional lipid or non-lipid carriers, making them excellent vehicles for protecting delicate drugs, such as peptides and proteins, from enzymatic degradation and *in vivo* degradation.⁷⁵ Nanocarriers, including cubosomes, exhibit minimal toxicity and biocompatible characteristics, making them effective delivery methods for a wide range of substances in various applications. ^{76–79}

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Cubosomes have shown promise as nanocarriers for anticancer medications. The unique structure of cubosomes suggests their potential application in melanoma treatment, with both passive and active targeting strategies demonstrating validity in preclinical and clinical research. In the realm of oral drug delivery, liquid crystalline NP technology emerges as a sophisticated solution, adept at orchestrating precise *in vivo* drug distribution. By strategically releasing particles at distinct absorption sites, such as the upper or lower intestine, it effectively navigates the challenges of regional absorption, a critical consideration for medications characterized by narrow absorption windows. ^{80–84}

In targeted drug delivery, cubosomes have exhibited enhanced permeability and retention when administered to rabbit corneal tissue sections, showcasing their potential for ocular applications. Additionally, cubosomes have been found to increase ocular bioavailability by prolonging the half-life at the corneal surface and exhibiting mucoadhesive properties, enhancing corneal permeability. For topical drug delivery, the bio-adhesive characteristics of cubic phases make them suitable for mucosal depositions and topical drug delivery systems. These systems leverage liquid crystal and liquid crystal NP technology to create bio-adhesive liquid crystalline systems in situ, facilitating precise and efficient drug distribution to mucosal surfaces. In contrast to conventional administration approaches, topical drug delivery systems offer temporary protection to sensitive and irritated skin by creating a thin layer on mucosal surfaces. These systems further fine-tune the nanostructure to attain the desired delivery profiles, representing a sophisticated approach to dermatological care. ^{52,85}

Conclusion

Cubosomes demonstrate strong potential as smart drug delivery systems, owing to their ability to respond to physiological stimuli such as temperature, pH, and enzymatic activity. This review presents that cubosomes support the initial hypothesis of their functional adaptability since they show great potential as carriers for site-specific and controlled drug delivery. Their distinctive structural characteristics and biocompatibility imply useful benefits in improving drug stability and release profiles. By minimizing off-target effects and enhancing delivery precision, cubosomes could contribute meaningfully to the development of safer and more effective therapies. Future studies may centre on optimizing formulation parameters, increasing manufacturing scale, and conducting in vivo studies to support their clinical relevance in particular therapeutic settings.

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Competing Interests

None.

References

1. Yusuf A, Almotairy ARZ, Henidi H, Alshehri OY, Aldughaim MS. Nanoparticles as Drug Delivery Systems: A Review of the Implication of Nanoparticles' Physicochemical Properties on Responses in Biological Systems. *Polymers (Basel)* 2023;15(7):1596. doi: 10.3390/polym15071596

2. Gowda BHJ, Ahmed MG, Alshehri SA, Wahab S, Vora LK, Singh Thakur RR, et al. The cubosomebased nanoplatforms in cancer therapy: Seeking new paradigms for cancer theranostics. *Environ Res* 2023;237. doi: 10.1016/j.envres.2023.116894

3. Sarkar S, Dyett B, Lakic B, Ball AS, Yeo LY, White JF, et al. Cubosome Lipid Nanocarriers As a Drug Delivery Vehicle for Intracellular Mycobacterium tuberculosis Infections. *ACS Appl Mater Interfaces* 2023;15(18):21819-29. doi: 10.1021/acsami.3c00101

4. Mertins O, Mathews PD, Angelova A. Advances in the design of ph-sensitive cubosome liquid crystalline nanocarriers for drug delivery applications. *Nanomaterials* 2020;10(5). doi: 10.3390/nano10050963

5. Bettini R, Catellani PL, Santi P, Massimo G, Peppas NA, Colombo P. *Translocation of Drug Particles in HPMC Matrix Gel Layer: Effect of Drug Solubility and Influence on Release Rate*. Vol 70.; 2001. www.elsevier.com/locate/jconrel

6. Spicer P. Cubosome processing: Industrial nanoparticle technology development. *Chemical Engineering Research and Design* 2005;83(11 A):1283-6. doi: 10.1205/cherd.05087

7. Waheed A, Aqil M. Lyotropic liquid crystalline nanoparticles: Scaffolds for delivery of myriad therapeutics and diagnostics. *J Mol Liq* 2021;338:116919. doi: 10.1016/j.molliq.2021.116919

8. Aleandri S, Bandera D, Mezzenga R, Landau EM. Biotinylated Cubosomes: A Versatile Tool for Active Targeting and Codelivery of Paclitaxel and a Fluorescein-Based Lipid Dye. *Langmuir* 2015;31(46):12770-6. doi: 10.1021/acs.langmuir.5b03469

9. Lancelot A, Sierra T, Serrano JL. Nanostructured liquid-crystalline particles for drug delivery. *Expert Opin Drug Deliv* 2014;11(4):547-64. doi: 10.1517/17425247.2014.884556

10. Kim DH, Jahn A, Cho SJ, Kim JS, Ki MH, Kim DD. Lyotropic liquid crystal systems in drug delivery: a review. *J Pharm Investig* 2015;45(1):1-11. doi: 10.1007/s40005-014-0165-9

11. Waheed A, Aqil M. Lyotropic liquid crystalline nanoparticles: Scaffolds for delivery of myriad therapeutics and diagnostics. *J Mol Liq* 2021;338. doi: 10.1016/j.molliq.2021.116919

12. Sharma P, Dhawan S, Nanda S. Cubosome: A Potential Liquid Crystalline Carrier System. *Curr Pharm Des* 2020;26(27):3300-16. doi: 10.2174/1381612826666200617162424

13. Shetty S, Shetty S. Cubosome-based cosmeceuticals: A breakthrough in skincare. *Drug Discov Today* 2023;28(7). doi: 10.1016/j.drudis.2023.103623

14. Umar H, Wahab HA, Gazzali AM, Tahir H, Ahmad W. Cubosomes: Design, Development, and Tumor-Targeted Drug Delivery Applications. *Polymers (Basel)* 2022;14(15). doi: 10.3390/polym14153118

15. Grandes Reyes CF, Ha S, Kim KT. Synthesis and applications of polymer cubosomes and hexosomes. *Journal of Polymer Science* 2023;61(12):1196-213. doi: 10.1002/pol.20230053

16. Azmi IDM, Moghimi SM, Yaghmur A. Cubosomes and hexosomes as versatile platforms for drug delivery. *Ther Deliv* 2015;6(12):1347-64. doi: 10.4155/tde.15.81

17. Kowark A, Keszei AP, Schneider G, Pilge S, Schneider F, Obert DP, et al. Preoperative Midazolam and Patient-Centered Outcomes of Older Patients: The I-PROMOTE Randomized Clinical Trial. *JAMA Surg* 2024;159(2):129-38. doi: 10.1001/jamasurg.2023.6479

18. Zhai J, Fong C, Tran N, Drummond CJ. Non-Lamellar Lyotropic Liquid Crystalline Lipid Nanoparticles for the Next Generation of Nanomedicine. *ACS Nano* 2019;13(6):6178-206. doi: 10.1021/acsnano.8b07961

19. Faisal MM, Gomaa E, Ibrahim AE, El Deeb S, Al-Harrasi A, Ibrahim TM. Verapamil-Loaded Cubosomes for Enhancing Intranasal Drug Delivery: Development, Characterization, Ex Vivo Permeation, and Brain Biodistribution Studies. *AAPS PharmSciTech* 2024;25(5). doi: 10.1208/s12249-024-02814-w

20. Balakrishnan P, Gopi S. Revolutionizing transdermal drug delivery: unveiling the potential of cubosomes and ethosomes. *J Mater Chem B* 2024;12(18):4335-60. doi: 10.1039/d3tb02927a

21. Khoshdooz S, Khoshdooz P, Bonyad R, Bonyad A, Sheidaei S, Nosrati R. Cubosomes-based hydrogels; A promising advancement for drug delivery. *Int J Pharm* 2025;674. doi: 10.1016/j.ijpharm.2025.125510

22. Mohammad Y, Prentice RN, Boyd BJ, Rizwan SB. Comparison of cubosomes and hexosomes for the delivery of phenytoin to the brain. *J Colloid Interface Sci* 2022;605:146-54. doi: 10.1016/j.jcis.2021.07.070

23. Luo Q, Lin T, Zhang CY, Zhu T, Wang L, Ji Z, et al. A novel glyceryl monoolein-bearing cubosomes for gambogenic acid : Preparation , cytotoxicity and intracellular uptake. *Int J Pharm* 2015;493(1-2):30-9. doi: 10.1016/j.ijpharm.2015.07.036

24. Verkhovskii RA, Ivanov AN, Lengert E V., Tulyakova KA, Shilyagina NY, Ermakov A V. Current Principles, Challenges, and New Metrics in pH-Responsive Drug Delivery Systems for Systemic Cancer Therapy. *Pharmaceutics* 2023;15(5). doi: 10.3390/pharmaceutics15051566

25. Vegad U, Patel M, Khunt D, Zupančič O, Chauhan S, Paudel A. pH stimuli-responsive hydrogels from non-cellulosic biopolymers for drug delivery. *Front Bioeng Biotechnol* 2023;11. doi: 10.3389/fbioe.2023.1270364

26. Chountoulesi M, Pispas S, Tseti IK, Demetzos C. Lyotropic Liquid Crystalline Nanostructures as Drug Delivery Systems and Vaccine Platforms. *Pharmaceuticals* 2022;15(4). doi: 10.3390/ph15040429

27. Alsawaftah NM, Awad NS, Pitt WG, Husseini GA. pH-Responsive Nanocarriers in Cancer Therapy. *Polymers (Basel)* 2022;14(5). doi: 10.3390/polym14050936

28. Mathews PD, Mertins O, Angelov B, Angelova A. Cubosomal lipid nanoassemblies with pH-sensitive shells created by biopolymer complexes: A synchrotron SAXS study. *J Colloid Interface Sci* 2022;607:440-50. doi: 10.1016/j.jcis.2021.08.187

29. Rajesh S, Zhai J, Drummond CJ, Tran N. Novel pH-Responsive Cubosome and Hexosome Lipid Nanocarriers of SN-38 Are Prospective for Cancer Therapy. *Pharmaceutics* 2022;14(10). doi: 10.3390/pharmaceutics14102175

30. Mertins O, Mathews PD, Angelova A. Advances in the design of ph-sensitive cubosome liquid crystalline nanocarriers for drug delivery applications. *Nanomaterials* 2020;10(5). doi: 10.3390/nano10050963

31. Rajesh S, Zhai J, Drummond CJ, Tran N. Novel pH-Responsive Cubosome and Hexosome Lipid Nanocarriers of SN-38 Are Prospective for Cancer Therapy. *Pharmaceutics* 2022;14(10). doi: 10.3390/pharmaceutics14102175

10 | Advanced Pharmaceutical Bulletin 2025

32. Manchun S, Dass CR, Sriamornsak P. Targeted therapy for cancer using pH-responsive nanocarrier systems. *Life Sci* 2012;90(11-12):381-7. doi: 10.1016/j.lfs.2012.01.008

33. Negrini R, Mezzenga R. PH-responsive lyotropic liquid crystals for controlled drug delivery. *Langmuir* 2011;27(9):5296-303. doi: 10.1021/la200591u

34. Prajapati R, Gontsarik M, Yaghmur A, Salentinig S. PH-Responsive Nano-Self-Assemblies of the Anticancer Drug 2-Hydroxyoleic Acid. *Langmuir* 2019;35(24):7954-61. doi: 10.1021/acs.langmuir.9b00838

35. Karimi M, Sahandi Zangabad P, Ghasemi A, Amiri M, Bahrami M, Malekzad H, et al. Temperature-Responsive Smart Nanocarriers for Delivery of Therapeutic Agents: Applications and Recent Advances. *ACS Appl Mater Interfaces* 2016;8(33):21107-33. doi: 10.1021/acsami.6b00371

36. Kojarunchitt T, Hook S, Rizwan S, Rades T, Baldursdottir S. Development and characterisation of modified poloxamer 407 thermoresponsive depot systems containing cubosomes. *Int J Pharm* 2011;408(1-2):20-6. doi: 10.1016/j.ijpharm.2011.01.037

37. Mohsen AM, Salama AAA, Asfour MH. Cubosome-based thermosensitive in situ gelling system for intranasal administration of lamotrigine with enhanced antiepileptic efficacy. *Pharm Dev Technol* 2023;28(6):520-34. doi: 10.1080/10837450.2023.2216755

38. Dabkowska AP, Hirst C, Valldeperas M, Clifton LA, Montis C, Nöjd S, et al. Temperature responsive lipid liquid crystal layers with embedded nanogels. *Chemical Communications* 2017;53(8):1417-20. doi: 10.1039/c6cc09426k

 Jia S, Tan A, Hawley A, Graham B, Boyd BJ. Visible light-triggered cargo release from donor acceptor Stenhouse adduct (DASA)-doped lyotropic liquid crystalline nanoparticles. *J Colloid Interface Sci* 2019;548:151-9. doi: 10.1016/j.jcis.2019.04.032

40. Jones BE, Kelly EA, Cowieson N, Divitini G, Evans RC. Light-Responsive Molecular Release from Cubosomes Using Swell-Squeeze Lattice Control. *J Am Chem Soc* 2022;144(42):19532-41. doi: 10.1021/jacs.2c08583

41. Fong WK, Negrini R, Vallooran JJ, Mezzenga R, Boyd BJ. Responsive self-assembled nanostructured lipid systems for drug delivery and diagnostics. *J Colloid Interface Sci* 2016;484:320-39. doi: 10.1016/j.jcis.2016.08.077

42. Chen Z, Huang Q, Song Y, Feng X, Zeng L, Liu Z, et al. Cubosomes-assisted transdermal delivery of doxorubicin and indocyanine green for chemo-photothermal combination therapy of melanoma. *Biomedicine and Pharmacotherapy* 2023;166. doi: 10.1016/j.biopha.2023.115316

43. Angelova A, Garamus VM, Angelov B, Tian Z, Li Y, Zou A. Advances in structural design of lipidbased nanoparticle carriers for delivery of macromolecular drugs, phytochemicals and anti-tumor agents. *Adv Colloid Interface Sci* 2017;249:331-45. doi: 10.1016/j.cis.2017.04.006

44. Jia S, Graham B, Capuano B, Tan A, Hawley A, Boyd BJ. Hexaarylbiimidazoles(HABI)-functionalized lyotropic liquid crystalline systems as visible light-responsive materials. *J Colloid Interface Sci* 2020;579:379-90. doi: 10.1016/j.jcis.2020.06.006

45. Bazylińska U, Kulbacka J, Schmidt J, Talmon Y, Murgia S. Polymer-free cubosomes for simultaneous bioimaging and photodynamic action of photosensitizers in melanoma skin cancer cells. *J Colloid Interface Sci* 2018;522:163-73. doi: 10.1016/j.jcis.2018.03.063

46. Tan C, Hosseini SF, Jafari SM. Cubosomes and Hexosomes as Novel Nanocarriers for Bioactive Compounds. *J Agric Food Chem* 2022;70(5):1423-37. doi: 10.1021/acs.jafc.1c06747

47. Zhang Y, Wang L, Wang ZD, Zhou Q, Zhou X, Zhou T, et al. Surface-anchored microbial enzymeresponsive solid lipid nanoparticles enabling colonic budesonide release for ulcerative colitis treatment. *J Nanobiotechnology* 2023;21(1). doi: 10.1186/s12951-023-01889-0

48. Umar H, Wahab HA, Gazzali AM, Tahir H, Ahmad W. Cubosomes: Design, Development, and Tumor-Targeted Drug Delivery Applications. *Polymers (Basel)* 2022;14(15). doi: 10.3390/polym14153118

49. Li M, Zhao G, Su WK, Shuai Q. Enzyme-Responsive Nanoparticles for Anti-tumor Drug Delivery. *Front Chem* 2020;8. doi: 10.3389/fchem.2020.00647

50. Fraceto JL, Estefaniaavangelieeramosscamposs G. *Nanotechnology in the Life Sciences Green Nanoparticles Synthesis and Biomedical Applications*. http://www.springer.com/series/15921

51. Alsuraifi A, Curtis A, Lamprou DA, Hoskins C. Stimuli responsive polymeric systems for cancer therapy. *Pharmaceutics* 2018;10(3). doi: 10.3390/pharmaceutics10030136

52. Rajesh S, Zhai J, Drummond CJ, Tran N. Synthetic ionizable aminolipids induce a pH dependent inverse hexagonal to bicontinuous cubic lyotropic liquid crystalline phase transition in monoolein nanoparticles. *J Colloid Interface Sci* 2021;589:85-95. doi: 10.1016/j.jcis.2020.12.060

53. Snell KE, Mevellec JY, Humbert B, Lagugné-Labarthet F, Ishow E. Photochromic organic nanoparticles as innovative platforms for plasmonic nanoassemblies. *ACS Appl Mater Interfaces* 2015;7(3):1932-42. doi: 10.1021/am5076953

54. Rajesh S, Zhai J, Drummond CJ, Tran N. Novel pH-Responsive Cubosome and Hexosome Lipid Nanocarriers of SN-38 Are Prospective for Cancer Therapy. *Pharmaceutics* 2022;14(10). doi: 10.3390/pharmaceutics14102175

55. Mathews PD, Mertins O, Angelov B, Angelova A. Cubosomal lipid nanoassemblies with pH-sensitive shells created by biopolymer complexes: A synchrotron SAXS study. *J Colloid Interface Sci* 2022;607:440-50. doi: 10.1016/j.jcis.2021.08.187

56. Rarokar NR, Saoji SD, Raut NA, Taksande JB, Khedekar PB, Dave VS. Nanostructured Cubosomes in a Thermoresponsive Depot System: An Alternative Approach for the Controlled Delivery of Docetaxel. *AAPS PharmSciTech* 2016;17(2):436-45. doi: 10.1208/s12249-015-0369-y

57. Mohsen AM, Salama AAA, Asfour MH. Cubosome-based thermosensitive in situ gelling system for intranasal administration of lamotrigine with enhanced antiepileptic efficacy. *Pharm Dev Technol* 2023;28(6):520-34. doi: 10.1080/10837450.2023.2216755

58. Balestri A, Lonetti B, Harrisson S, Farias-Mancilla B, Zhang J, Amenitsch H, et al. Thermo-responsive lipophilic NIPAM-based block copolymers as stabilizers for lipid-based cubic nanoparticles. *Colloids Surf B Biointerfaces* 2022;220. doi: 10.1016/j.colsurfb.2022.112884

59. Li Y, Angelova A, Hu F, Garamus VM, Peng C, Li N, et al. PH Responsiveness of Hexosomes and Cubosomes for Combined Delivery of Brucea javanica Oil and Doxorubicin. *Langmuir* 2019;35(45):14532-42. doi: 10.1021/acs.langmuir.9b02257

60. Lakshmi NM, Yalavarthi PR, Vadlamudi HC, Thanniru J, Yaga G. Send Orders for Reprints to Reprints@benthamscience.Net Cubosomes as Targeted Drug Delivery Systems-A Biopharmaceutical Approach. Vol 11.; 2014.

61. Gaballa SA, El Garhy OH, Moharram H, Abdelkader H. Preparation and Evaluation of Cubosomes/Cubosomal Gels for Ocular Delivery of Beclomethasone Dipropionate for Management of Uveitis. *Pharm Res* 2020;37(10). doi: 10.1007/s11095-020-02857-1

62. Garg G, Saraf S, Saraf S. Cubosomes: An Overview Structure of Different Cubosomes. Vol 30.; 2007.

63. Bryant SJ, Bathke EK, Edler KJ. *Bottom-Up Cubosome Synthesis Without Organic Solvents*.

64. Gaballa SA, El Garhy OH, Abdelkader H. *Cubosomes: Composition, Preparation, and Drug Delivery Applications.* http://jabps.journals.ekb.eg

65. Nagao M, Ranneh AH, Iwao Y, Yamamoto K, Ikeda Y. Preparation of Cubosomes with Improved Colloidal and Structural Stability Using a Gemini Surfactant. *Mol Pharm* 2023;20(10):5066-77. doi: 10.1021/acs.molpharmaceut.3c00378

66. N H, C M, T. R M, S S, S N, K. E M, et al. In Vitro Hepatic Models to Assess Herb–Drug Interactions: Approaches and Challenges. *Pharmaceuticals* 2023;16(3):1-12. doi: 10.3390/ph16030409

67. Verlinde CL, Hol WG. Structure-based drug design: progress, results and challenges. *Structure* 1994;2(7):577-87. doi: 10.1016/S0969-2126(00)00060-5

 Kádár R, Spirk S, Nypelö T. Cellulose Nanocrystal Liquid Crystal Phases: Progress and Challenges in Characterization Using Rheology Coupled to Optics, Scattering, and Spectroscopy. ACS Nano 2021;15(5):7931-45. doi: 10.1021/acsnano.0c09829

69. Elnaggar YSR, Etman SM, Abdelmonsif DA, Abdallah OY. Novel piperine-loaded Tween-integrated monoolein cubosomes as brain-targeted oral nanomedicine in Alzheimer's disease: Pharmaceutical, biological, and toxicological studies. *Int J Nanomedicine* 2015;10:5459-73. doi: 10.2147/IJN.S87336

70. Rakotoarisoa M, Angelov B, Garamus VM, Angelova A. Curcumin- and Fish Oil-Loaded Spongosome and Cubosome Nanoparticles with Neuroprotective Potential against H 2 O 2 -Induced Oxidative Stress in Differentiated Human SH-SY5Y Cells. *ACS Omega* 2019;4(2):3061-73. doi: 10.1021/acsomega.8b03101

71. Sivadasan D, Sultan MH, Alqahtani SS, Javed S. Cubosomes in Drug Delivery—A Comprehensive Review on Its Structural Components, Preparation Techniques and Therapeutic Applications. *Biomedicines* 2023;11(4). doi: 10.3390/biomedicines11041114

72. Likhitha U, Nayak UY. Advances in tailored drug delivery systems: Amphiphilic lyotropic mesomorphs for targeted therapeutic intervention. *J Mol Liq* 2024;403. doi: 10.1016/j.molliq.2024.124738

73. Bachhav AA, Pingale PL, Upasani CD, Ahire SA. CUBOSOME: A NOVEL VESICULAR DRUG DELIVERY SYSTEM. *Int J Pharm Sci Res* 2024;15(2):323. doi: 10.13040/IJPSR.0975-8232.15(2).323-39

74. Umar H, Wahab HA, Gazzali AM, Tahir H, Ahmad W. Cubosomes: Design, Development, and Tumor-Targeted Drug Delivery Applications. *Polymers (Basel)* 2022;14(15). doi: 10.3390/polym14153118

75. Zhu C, Duan W, Jing H, Long J, Huang Y, Huang D, et al. Improving the stability and transdermal permeability of phycocyanin loaded cubosomes. *Frontiers in Nanotechnology* 2024;6. doi: 10.3389/fnano.2024.1359219

76. Vitoria Pupo Silvestrini A, Wender Debiasi B, Garcia Praça F, Vitoria Lopes Badra Bentley M. Progress and challenges of lyotropic liquid crystalline nanoparticles for innovative therapies. *Int J Pharm* 2022;628(July):122299. doi: 10.1016/j.ijpharm.2022.122299

77. Chountoulesi M, Pispas S, Tseti IK, Demetzos C. Lyotropic Liquid Crystalline Nanostructures as Drug Delivery Systems and Vaccine Platforms. *Pharmaceuticals* 2022;15(4):1-32. doi: 10.3390/ph15040429

78. Blanco-Fernández G, Blanco-Fernandez B, Fernández-Ferreiro A, Otero-Espinar FJ. Lipidic lyotropic liquid crystals: Insights on biomedical applications. *Adv Colloid Interface Sci* 2023;313(February). doi: 10.1016/j.cis.2023.102867

79. Chavda VP, Dawre S, Pandya A, Vora LK, Modh DH, Shah V, et al. Lyotropic liquid crystals for parenteral drug delivery. *Journal of Controlled Release* 2022;349:533-49. doi: 10.1016/j.jconrel.2022.06.062

80. Psimadas D, Georgoulias P, Valotassiou V, Loudos G. Molecular Nanomedicine Towards Cancer : *J Pharm Sci* 2012;101(7):2271-80. doi: 10.1002/jps

81. Tsakiris N, Papavasileiou M, Bozzato E, Lopes A, Vigneron AM, Préat V. Combinational Drug-Loaded
Lipid Nanocapsules for the Treatment of Cancer. *Int J Pharm* 2019;19(7):1-27. doi: 10.1016/j.ijpharm.2019.118588

82. Gadag S, Narayan R, Nayak Y, Garg S, Nayak UY. Design, development and evaluation of Resveratrol transdermal patches for breast cancer therapy. *Int J Pharm* 2023;632(October 2022):122558. doi: 10.1016/j.ijpharm.2022.122558

83. Gadag S, Narayan R, Sabhahit JN, Hari G, Nayak Y, Pai KSR, et al. Transpapillary iontophoretic delivery of resveratrol loaded transfersomes for localized delivery to breast cancer. *Biomaterials Advances* 2022;140(May):213085. doi: 10.1016/j.bioadv.2022.213085

84. Arti Majumdar, Nidhi Dubey ND. Dermal delivery of docetaxel loaded nano liquid crystals for the treatment of skin cancer. *International Journal of Applied Pharmaceutics* 2019;11(5):8-13.

85. Zhai J, Fong C, Tran N, Drummond CJ. Non-Lamellar Lyotropic Liquid Crystalline Lipid Nanoparticles for the Next Generation of Nanomedicine. *ACS Nano* 2019;13(6):6178-206. doi: 10.1021/acsnano.8b07961

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Figures

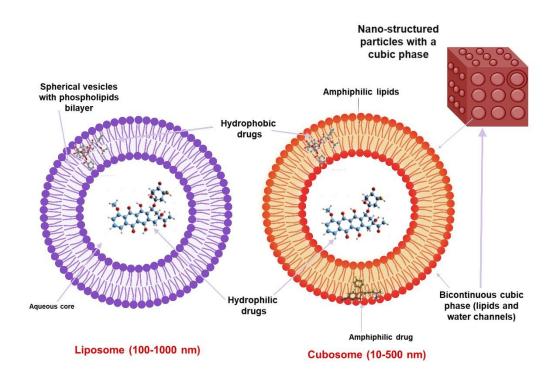


Figure 1.

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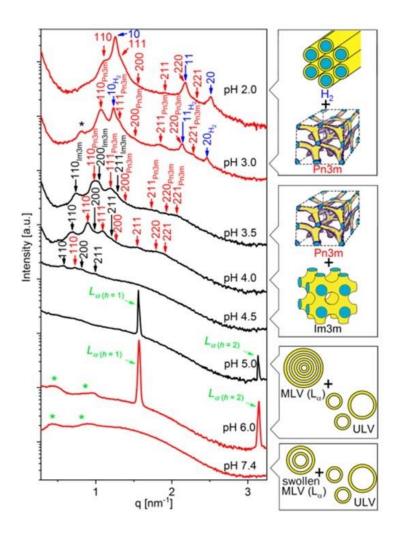
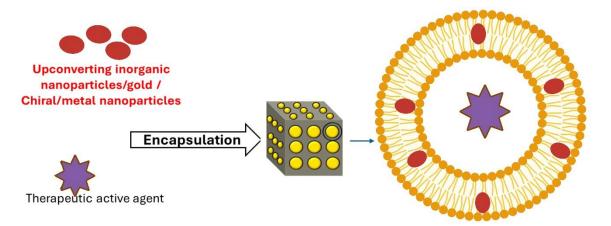


Figure 2.

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Plasmonic Nanoparticles were co-encapsulated with an Active agent within the cubosomes to enhance the therapeutic and diagnostic capabilities of the nanoplatforms

Figure 3.