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Evaluating the Efficacy of a Semi-Solid Formulation with Clove Oil and Curcumin versus Clindamycin in the Treatment of Acne Vulgaris: A Comprehensive Study of Preclinical and Clinical Findings

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#### ARTICLE INFO

#### **ABSTRACT**

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#### **Article History:**

Submitted: January 15, 2025 Revised: August 13, 2025 Accepted: August 15, 2025 ePublished: August 30, 2025 **Purpose:** Acne vulgaris is a chronic inflammatory condition affecting the pilosebaceous units. With the rise of antibiotic resistance and the potential side effects associated with conventional treatments, there is increasing interest in exploring natural alternatives for treating acne.

This study aimed to formulate and clinically evaluate a topical gel containing clove oil and curcumin in patients with mild to moderate acne vulgaris, using topical clindamycin as a standard comparator.

**Methods:** The antibacterial activity of clove oil and curcumin against *Propionibacterium acnes* (*P. acnes*) and *Staphylococcus epidermidis* (*S. epidermidis*) was assessed by determining their minimum inhibitory concentration (MIC) and minimum bactericidal concentration (MBC). Based on these findings, semisolid gel formulations were developed and subjected to in vitro evaluations. Subsequently, a single-blind, randomized, comparative clinical study was conducted in 31 participants diagnosed with mild to moderate acne. Volunteers applied clindamycin gel in the morning and the clove oil–curcumin gel in the evening, or vice versa, over a 4-week period. Clinical outcomes, including acne lesion counts, Acne Severity Index (ASI), and patient satisfaction, were assessed. **Results:** The clove oil-curcumin gel demonstrated comparable efficacy to clindamycin in reducing acne lesions, papules, and ASI. While no significant differences were observed between the two groups in comedone reduction, patient satisfaction increased in clove oil-curcumin gel group.

**Conclusion:** Topical application of a gel containing clove oil and curcumin demonstrated promising results as an effective and well-tolerated alternative or adjunctive therapy for acne vulgaris. These findings support the potential of plant-based formulations in acne management.

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#### **Introduction:**

Acne vulgaris is a prevalent chronic skin condition associated with inflammation of the pilosebaceous units. It typically presents as non-inflammatory and inflammatory lesions, such as comedones, papules, pustules, nodules, and cysts, and may result in permanent scarring. The disease develops through a multifactorial process that includes excessive sebum secretion, abnormal keratinization within hair follicles, bacterial colonization particularly by P. acnes and S. epidermidis and subsequent inflammatory responses.<sup>1,2</sup>

The psychological burden of acne is substantial, as it can lead to emotional distress, reduced self-confidence, social isolation, and in severe cases, anxiety, depression, or suicidal thoughts.3-5 Therefore, effective treatment strategies are not only aimed at lesion reduction but also at improving patients' quality of life.

Currently available treatments comprise topical agents (e.g., benzoyl peroxide, tretinoin, azelaic acid and topical antibiotics e.g., clindamycin) and systemic medications (e.g., oral antibiotics, isotretinoin). Despite their clinical efficacy, these therapies often present drawbacks, including microbial resistance, gastrointestinal side effects, photosensitivity, and potential teratogenicity, especially with long-term use.<sup>7-10</sup> As a result, there is increasing interest in the development of safer, naturally derived alternatives with fewer side effects.

Although various herbal agents have been traditionally used for acne, standardized and officially approved natural products remain scarce in many countries. This prompted research into natural formulations including herbals that may offer therapeutic benefits with enhanced safety profiles.

The current study explores the formulation of two semisolid topical preparations containing natural active pharmaceutical ingredients (APIs) with established antimicrobial and anti-inflammatory properties: clove oil and curcumin.

Clove oil, obtained from Syzygium aromaticum showed broad-spectrum antimicrobial activity, particularly against acne-associated bacteria. It also exhibits anti-inflammatory and antioxidant activities, so its potential of dermatological applications was stablished previously. 11-16

Curcumin, a polyphenol derived from the rhizome of Curcuma longa, exhibits antimicrobial, anti-inflammatory, and antioxidant properties. It inhibits the growth of P. acnes and S. epidermidis, suppresses the production of inflammatory cytokines by P. acnes, and promotes wound healing and scar reduction. 17-28

This study aimed to assess the feasibility of incorporating these two natural agents into topical formulations for the management of mild to moderate acne, potentially contributing to the development of safer and more acceptable treatment options as well as reducing the psychological effects of acne in patients.

#### **Material and Methods**

#### Materials:

Tween 80, Span 80, propylene glycol, methylparaben, propylparaben, and triethanolamine were purchased from Merck (Germany). Microbial culture media were obtained from Merck (Germany) and QUELAM. Curcumin was sourced from Sami Lab (India), and clove oil was provided by Ghatran Shimi Co. (Iran). Standard strains of P. acnes ATCC 6919 and S. epidermidis ATCC 12228 were obtained from the Pasteur Institute of Iran's microbial culture collection. Cultures were maintained on appropriate media under standard conditions.

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Methods:

#### Antimicrobial Susceptibility Testing:

Disk Diffusion Method:

Bacterial suspensions ( $1.5 \times 10^8$  CFU/mL) of *P. acnes* and *S. epidermidis* were prepared and inoculated on blood agar and Mueller-Hinton agar plates, respectively. Sterile filter paper discs were impregnated with various concentrations of clove oil and curcumin solutions for 1 hour and then placed on the inoculated plates. Prediffusion was allowed for 30 minutes at room temperature. Plates were incubated at 37°C for 96 hours for *P. acnes* and 20 hours for *S. epidermidis*. Standard clindamycin discs ( $2 \mu g/disc$ ) served as positive controls. Inhibition zones were measured in millimeters. All experiments were performed in triplicate.<sup>28</sup>

Agar Well Diffusion Method:

Bacterial suspensions ( $1.5 \times 10^8$  CFU/mL equal to 0.5 McFarland) were prepared. Suspensions were swabbed onto the surface of blood agar plates for *P. acnes* and Mueller-Hinton agar plates for *S. epidermidis*. Clove oil solutions in ethanol were prepared at concentrations ranging from 0.0625 to 4% (v/v) for *S. epidermidis* and 0.03125 to 2% (v/v) for *P. acnes*. Curcumin solutions in ethanol were prepared at concentrations ranging from 1000 to 31.25 µg/mL. Wells were created in the agar plates. Solutions were added to the respective wells. Plates were pre-incubated for 30 minutes. Plates were incubated at 37°C for 20 hours for *S. epidermidis* and 96 hours for *P. acnes*. Zones of inhibition were measured. <sup>29</sup>

Broth Micro Dilution Method:

The minimum inhibitory concentrations (MICs) and minimum bactericidal concentrations (MBCs) of clove oil and curcumin were determined using the broth micro dilution method in sterile 96-well microplates.

Experimental Procedure:

Each well was filled with 100  $\mu$ L of Mueller-Hinton broth for *S. epidermidis* and Brain Heart Infusion (BHI) broth for *P. acnes*. Stock solutions of clove oil and curcumin were added to the first wells and subjected to eight-fold serial dilutions across the plate. Bacterial suspensions were adjusted to 0.5 McFarland standard (~1.5  $\times$  10<sup>8</sup> CFU/mL), and 60  $\mu$ L of each suspension was added to the wells, resulting in a final bacterial concentration of approximately 1.5  $\times$  10<sup>8</sup> CFU/mL per well. Positive and negative controls were included in each assay. The negative control consisted of clindamycin and broth media (no bacterial growth), while the positive control consisted of bacterial suspension and media without antimicrobial agents. The plates were incubated at 37°C under anaerobic conditions for 96 hours for P. acnes and under aerobic conditions for 24 hours for S. epidermidis. All tests were performed in triplicate.

MIC Determination:

The MIC was defined as the lowest concentration of the test compound that completely inhibited visible bacterial growth.

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MBC Determination:

Aliquots ( $10 \,\mu\text{L}$ ) from wells at and above the MIC level were sub cultured on appropriate agar plates and incubated under the same conditions described above. The MBC was defined as the lowest concentration of the test agent resulting in no visible bacterial growth on the agar surface. All experiments were conducted in triplicate.

Antibacterial combination study by Checkerboard method:

The checkerboard microdilution method was employed to determine the FIC indices of the active ingredients.

Experimental Procedure:

After determining the individual MICs of each active ingredient, their combined effects were evaluated using the microdilution method. 100  $\mu$ L of selective broth media and 60  $\mu$ L of bacterial inoculum (1.5 × 10<sup>8</sup> CFU/mL) were added to each microtiter plate well. Two-fold serial dilutions of the active ingredients were prepared in combination (1:1 v/v), and 100  $\mu$ L of each combination was added to the wells. Incubation conditions were maintained as previously described for individual MIC determinations.

FIC Index Calculation:

The FIC index was calculated using the following formula:

FIC index = (MIC of EOA in combination / MIC of EOA alone) + (MIC of EOB in combination / MIC of EOB alone)

Where EOA and EOB represent the two different essential oils being tested.

FIC indices were interpreted as follows:

- FICI  $\leq$  0.5: Synergistic interaction
- $0.5 < FICI \le 4$ : Additive interaction
- FICI > 4: Antagonistic interaction

All experiments were performed in triplicate.

#### Formulation of gels:

Tween 80, ethanol, and propylene glycol were used as co-solvents for clove oil, while propylene glycol was selected as the levigating agent for curcumin. The composition of the investigated formulations is summarized in table 1. Carbopol 940 was dispersed in distilled water and stirred at 200–300 rpm for 2 hours. Clove oil and curcumin were levigated with titanium dioxide (used as a masking agent) in propylene glycol. This mixture was subsequently dissolved in ethanol containing 0.25% (w/v) methylparaben and 0.05% (w/v) propylparaben as preservatives. The resulting solution was added to the previously prepared carbomer dispersion and stirred for an additional 10 minutes at 200–300 rpm. The final pH of the formulation was adjusted to 4–6 using triethanolamine.

Table 1. Formulation composition of gels and emulgels

#### Formulation composition of gels

*Formulation	TiO <sub>2</sub>	Carbopol % (w/w)	Ethanol	Propylen Glycol % (w/w)	Tween 80	LLP	Span
A1	0.5	10	5	2	5	-	-
A2	0.6	10	5	2	5	-	-
A3	0.75	10	5	2	5	-	-
A4	1	10	5	2	5	-	-
B1	0.6	10	7.5	2	2.5	-	-
B2	0.6	20	7.5	2	2.5	-	-
В3	0.6	30	7.5	2	2.5	-	-
C1	0.6	30	7.5	2	5	-	-
C2	0.6	30	10	2	5	-	-
		Formul	ation compo	sition of emulgel	s		
D1	5	0.5	7.5	0.84	10	2.16	5
D2	5	0.6	7.5	0.84	10	2.16	5
D3	5	0.75	7.5	0.84	10	2.16	5
D4	5	1	7.5	0.84	10	2.16	5
<b>E</b> 1	7.5	0.6	7.5	0.84	10	2.16	2.5
E2	10	0.6	7.5	0.84	10	2.16	2.5
F1	10	0.6	7.5	0.84	10	2.16	5
F2	10	0.6	10	0.84	10	2.16	5

#### Formulation of Emulgel:

The detailed composition of the investigated emulgel formulations is presented in table 1. The gel base was prepared by dispersing Carbopol 940 in purified water with continuous stirring, followed by neutralization with triethanolamine to adjust the pH to 4–6. The oil-in-water (O/W) emulsion was prepared by separately heating and mixing the oil and aqueous phases. Oil phase was prepared by mixing of light liquid paraffin (LLP) with appropriate amounts of Tween 80 and Span 80. The surfactant ratio was determined based on the required HLB value. For preparing of aqueous phase, methylparaben and propylparaben were dissolved in purified water. Both phases were heated to  $75 \pm 1^{\circ}$ C and  $80 \pm 1^{\circ}$ C, respectively. The aqueous phase was added dropwise to the oil phase under continuous stirring, and the emulsion was allowed to cool gradually to room temperature. Clove oil, curcumin, and titanium dioxide were levigated in propylene glycol and dissolved in ethanol. This solution was then incorporated into the emulsion. Finally, the prepared emulsion was blended with the gel base in a 1:1 ratio under continuous stirring to obtain the final emulgel.

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#### In-vitro evaluation of prepared formulations:

Physical Appearance:

All prepared gel and emulgel formulations were evaluated for color, homogeneity, consistency, grittiness, and phase separation conditions.<sup>30-32</sup>

Spreadability Assessment:

The formulations were manually applied to the back of the hand to assess spread ability. Formulations demonstrating uniform distribution, ease of application, and absence of grittiness were considered acceptable.

Quantitative Measurement:

An accurately weighed amount (0.5 g) of each sample was placed between two glass slides. A 500 g weight was applied for 1 minute to achieve uniform thickness. The diameter of the spread was measured, and spread ability was calculated based on the circular area.<sup>33</sup>

Stress Stability test:

The mechanical stability of the emulgel was evaluated through centrifugation under different rpms and times. After each centrifugation cycle, the formulations were visually inspected for phase separation. Stable emulgels were defined as those exhibiting no signs of creaming, cracking, or phase separation during or after the test.<sup>34</sup>

Determination of Emulgel Type (O/W or W/O):

Water Wash Test: A small amount of emulgel was placed on the hand and washed with water. Ease of removal with water typically indicates an oil-in-water (O/W) emulsion, where water constitutes the continuous external phase.

Methylene Blue Dye Test:

Methylene blue dye is water-soluble and exhibits a blue color in aqueous solutions. For this test, a small amount of methylene blue dye was added to the emulgel. The appearance of a blue color within the emulgel suggests an oil-in-water (O/W) emulsion.

#### pH Determination:

The pH of all formulations was determined using calibrated pH-meter. Each formulation consists of water (propyl paraben 0.02% w/w and methyl paraben 0.18% w/w). All the formulations were neutralized by triethanolamine to pH=4-6.

Viscosity measurement and rheological behavior studies:

The viscosity and rheological behavior of the final formulations were evaluated using a Brookfield viscometer equipped with spindle 52.

0.5 g of each gel sample was placed on the viscometer plate. The cone spindle was carefully lowered onto the sample. Viscosity measurements were conducted at room temperature. The rotational speed (rpm) was varied, and the corresponding viscosity, shear stress, shear rate, and torque values were recorded. A plot of viscosity versus rotational speed (rpm) was generated. The shape of the resulting curve provided insights into the rheological behavior of the gel formulation.<sup>35</sup>

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Heat-cold cycle stability test:

To assess the stability of the formulations under temperature fluctuations, they were subjected to three cycles of temperature, 4°C, ambient temperature and 40°C. After each cycle, preparations were tested for any signs of phase separation, cracking, or other physical instability. Formulations that maintained their uniformity throughout the three cycles were considered stable.<sup>33</sup>

Assay of Active Ingredients:

The amount of both active ingredients in each prepared semi solid preparations, were measured accurately.

Release profile study:

The in vitro drug release profiles of the formulations were evaluated using two complementary methods as following description. Dialysis bag method in which predefined amount of each formulation was enclosed in a synthetic dialysis membrane (molecular weight cut-off: 14,000 Da) and immersed in an appropriate release medium under sink conditions and franz diffusion cell method in which the receptor compartment of the franz diffusion cell was filled with 50 mL of phosphate buffer (pH 6.8) containing 40% ethanol to ensure drug solubility. The formulation was applied to the donor compartment, and the system was maintained at  $37 \pm 0.5$ °C with continuous magnetic stirring at 100 rpm. Aliquots (5 mL) were withdrawn from the receptor compartment at predetermined time intervals over a 24-hour period and replaced with an equal volume of fresh medium to maintain sink conditions. Skin permeability test which can provide more details is recommended for future studies, the presented study was focused on the evaluation of the clinical efficacy of prepared formulations and some in vitro characterizations were carried out before the application of the formulations. Samples collected from both methods were analyzed using UV-Vis spectrophotometry to quantify the amount of drug released. All experiments were performed in triplicate (n = 3), and results are presented as mean  $\pm$  standard deviation

Antimicrobial Activity Assessment:

The antimicrobial activity of the optimized formulation was evaluated using the well diffusion method. Mueller-Hinton agar was used for *S.epidermidis*. Blood agar was used for *P.acnes*. Sterile media were prepared and poured into Petri dishes to solidify.

Suspension of *S. epidermidis* and *P. acnes* was prepared at a concentration of  $1.5 \times 10^8$ CFU/mL. The entire surface of each agar plate was evenly inoculated with the respective bacterial suspension.

Wells were aseptically created in the agar plates. An appropriate volume of the optimized formulation was aseptically dispensed into each well. Plates were incubated at 37°C for 24 hours for *S. epidermidis* and 96 hours for *P. acnes* to allow for optimal microbial growth.

The diameters of the zones of inhibition around each well were measured in millimeters using a ruler. The experiment was performed in triplicate to ensure reproducibility.

Antimicrobial studies revealed that the mean inhibition zone diameters for *S. epidermidis* were 19 mm and 16 mm for the gel (C2) and emulgel (F2), respectively, which were significantly smaller than that of the clindamycin gel control (35 mm; p < 0.0001). No statistically significant difference was observed between the gel (C2) and emulgel (F2) formulations (p = 0.2756). Similarly, for *P. acnes*, the inhibition zones measured 24 mm and 22 mm for the gel (C2) and emulgel (F2), respectively, both of which were significantly smaller than that of the

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clindamycin gel (36 mm; p < 0.001). Again, no significant difference was detected between the gel (C2) and emulgel (F2) (p = 0.6793).

#### Clinical Trial:

A single-blind, comparative clinical study was conducted involving 31 patients diagnosed with mild to moderate acne vulgaris. The study population comprised 13 females and 14 males, with a mean age of 23.96 years (range: 12–37 years).

Inclusion criteria were as follows:

- Age  $\geq$  12 years
- Non-pregnant and non-lactating during the study period
- Provision of written informed consent
- No known hypersensitivity to herbal products, including clove oil and curcumin
- No use of systemic acne treatments within 30 days prior to enrollment
- No use of topical anti-acne therapies within 2 weeks prior to enrollment

Exclusion criterion included withdrawal of consent at any point during the study.

Study Design:

Each participant received two gels including Gel 1 which was Clindamycin gel and Gel 2 that contains combination of clove oil and curcumin.

Treatment Protocol:

To minimize confounding variables, each patient applied both gels to contralateral body sites affected by acne. One gel was applied on one side of his/her body surface with acnes and the other gel was applied in the other side of the body for him/her. So, many variables effects like genetic factors were limited to influence the results of the study.

One group of volunteers were instructed to apply just clindamycin gel (G1) twice a day and Other groups received one time G1 and one time combination gel of clove oil and curcumin (G2) in a day. The use of any other anti-acne medications or products was prohibited during the study period. As our clinical study lacks placebo and the absence of randomization between gel types, randomized controlled trials in future work are recommended.

#### Follow-up:

A one-week follow-up phone call was conducted to assess for any adverse events.

At four weeks, patients were evaluated based on their lesion counts also, patient-reported outcomes assessed through a standardized questionnaire.

Statistical Methods:

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The Kolmogorov-Smirnov test indicated non-normal distribution of all quantitative variables. Therefore, the Wilcoxon signed-rank test was used to compare pre- and post-intervention values within each group, and the Mann-Whitney U test was used to compare changes between groups.

Significant reductions were observed in the sample group compared to baseline for the following: total acne lesion count (p = 0.000), comedone count (p = 0.005), papule count (p = 0.000), pustule count (p = 0.002), and ASI (p = 0.000).

Between-group comparisons revealed significant differences in the reduction of total acne lesions, papules, and ASI (p = 0.000) in favor of the combination therapy group. No significant between-group differences were observed in the reduction of comedones (p = 0.375) or pustules (p = 0.062).

#### Results and Discussion:

#### Antimicrobial evaluation results:

Clove oil and curcumin demonstrated antimicrobial activity against P. acnes and S. epidermidis. The zones of inhibition were recorded in table 2.

Table2. Antibacterial inhibition zones caused by active ingredients

		Concentrations (v/v %)	Inhibition zone (mm) Disc diffusion method	Inhibition zone (mm) Agar diffusion method
		4	12	16
		2	11	13
		1	8	10.5
	S. epidermidis	0.5	7.5	9
		0.25	_	Grown
		0.125		Grown
		0.0625		Grown
~1. II		Control (-)	Grown	Grown
Clove oil		Clindamycin	23	22
-		2	16	22.5
		1	15	22
		0.5	14	21
		0.25	13	19.5
	P. acnes	0.125	Grown	12.5
		0.0625	Grown	Grown
		0.03125	Grown	Grown
		Control (-)	Grown	Grown
		Clindamycin	35	36
		1000	13	17
		500	12	14
		250	Grown	10.5
	S. epidermidis	125	Grown	Grown
	~. opiwei iiiwiii	62.5	Grown	Grown
		31.25	Grown	Grown

Curcumin	Control (-)	Grown	Grown
	Clindamycin	23	22
	2000	12.5	-
	1000	12	16
	500	11	15
P. acnes	250	9.5	12
	125	Grown	Grown
	62.5	_	Grown
	31.25	_	Grown
	Control (-)	Grown	Grown
	Clindamycin	35	36

#### Minimum Inhibitory Concentration (MIC):

MIC of clove oil and curcumin for *P. acnes* was 0.25% (v/v) and  $250 \mu g/mL$  for curcumin, respectively. MIC of clove oil and curcumin for *S. epidermidis* was 0.5% (v/v) and  $500 \mu g/mL$ .

#### Minimum Bactericidal Concentration (MBC):

MBC of clove oil and curcumin for P. acnes was 2.5% (v/v) and 4 mg/mL, respectively and for S. epidermidis was obtained in 4% (v/v) by clove oil and 4 mg/mL by curcumin.

#### Checkerboard Assay:

Against S. epidermidis, the combination of clove oil and curcumin exhibited synergistic activity.

Against *P. acnes*, the combination of clove oil and curcumin demonstrated either additive effects or no significant interaction.

#### **Formulation Development:**

Based on the MBC values determined for both bacteria, gel and emulgel formulations were developed. Subsequent characterization and evaluation of these formulations were conducted.

#### **Visual Inspections:**

All prepared gels and emulgels exhibited a clear, uniform appearance and were free of any discernible odor.

### Evaluation of Extrudability and Spreadability:

The ease of extrusion from the container and the spreadability of the gels and emulgels were evaluated. Formulations containing 0.75% and 1% Carbomer 940 exhibited high consistency, resulting in difficulty in extrusion and poor spreadability on both the skin and glass plates. Formulations containing 0.5% Carbomer 940 demonstrated poor extrudability from the container due to their loose consistency. Formulations containing 0.6% Carbomer 940 demonstrated optimal extrudability and spreadability upon manual application.

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**Formulation Optimization:** 

Formulation optimization was conducted to improve gel stability, appearance, and antimicrobial efficacy.

Increasing propylene glycol concentration to 10% prevented dehydration during storage, while 5% titanium

dioxide effectively masked the yellow color of curcumin. Additionally, incorporating 30% ethanol significantly

enhanced antibacterial activity against Staphylococcus epidermidis, likely due to improved solubility and

synergistic effects with clove oil and curcumin. These modifications were applied to the final gel formulation.

Emulgel Formulation Optimization:

Emulgel formulation was optimized to enhance stability and aesthetic properties. Increasing propylene glycol

concentration to 10% prevented dehydration during storage, while 5% titanium dioxide effectively masked

curcumin's yellow color. Varying light liquid paraffin concentrations (7.5% vs. 10%) showed no notable

difference in appearance. The final emulgel formulations were evaluated for the release profiles of clove oil and

curcumin.

Stability in the Heat-Cold Cycle:

Both the optimized gel and emulgel formulations demonstrated stability after undergoing three cycles of

temperature fluctuations (refrigeration at 4°C, ambient temperature, and incubation at 40°C). No significant

physical changes, such as phase separation or cracking, were observed in formulations.

pH Determination:

The pH of the gel was 6.09, and the pH of the emulgel was 5.86. Both values fall within the acceptable range for

topical skin products (pH 4-6).

Emulgel Type Determination:

Water Wash Test: The emulgel was easily removed by washing with water, indicating an oil-in-water (O/W)

emulsion.

Methylene Blue Dye Test: The addition of methylene blue dye to the emulgel resulted in a green color,

confirming it as an oil-in-water (O/W) emulsion.

Assay of Active Ingredients:

All the results of the assay were in acceptable range of 90-110% of claimed amount.

In vitro Release:

Franz diffusion cells were employed to evaluate the in vitro release profiles of the active ingredients. The results

are presented in Figure 1 as is shown below.

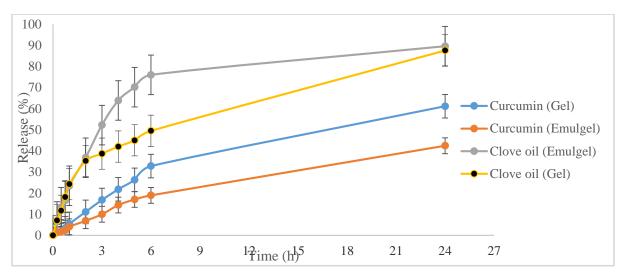


Figure 1. Release study graphs.

#### Viscosity and rheological behavior:

Both gel and emulgel formulations exhibited non-Newtonian, shear-thinning behavior; the gel was thixotropic, while the emulgel showed stable, non-thixotropic flow.

#### Antimicrobial effect of desired formulations and clindamycin gel:

The results of these comparison is presented in table 3.

Table 3. Comparison of antibacterial effects of designed formulations with clindamycin

Type of bacteria	Formulation	Inhibition zone (mm)
S. epidermidis	Gel (C2)	19
5. epiaermais	Emulgel (F2)	16.5
	Clindamycin gel	35
	Gel (C2)	24
P. acnes	Emulgel (F2)	22
	Clindamycin gel	36

Based on the observation of larger zones of inhibition exhibited by the gel formulation against both bacterial strains compared to the emulgel, the gel formulation was selected for subsequent clinical investigation.

#### Clinical studies:

Table 4 presents the demographic characteristics of the study participants. Twenty seven patients completed the study, while 4 were excluded due to factors such as non-adherence to the treatment regimen, missed follow-up appointments, or the concurrent use of other acne medications. The exclusion rate of 12.91% was considered negligible, with no significant impact on the overall study results or the validity of the conclusions.

Table 4. The demographic characteristics of the study participants

Description	Number	Percentage
Age (years)		
≤20	9	33.3
21-30	15	55.6
31-40	3	11.1
Gender		
Male	14	51.9
Female	13	48.1
Drug history for acne		
No history	15	55.6
Topical	3	11.1
Oral	3	11.1
Topical & Oral	6	22.2
Skin diseases history		
No history	11	40.7
Acne	13	48.1
Skin blemish	1	3.7
Skin allergy	1	3.7
Family skin diseases history		
No history	17	63
Acne	5	18.5
Skin blemish	2	7.4
Skin allergy	3	11.1

#### Statistical analysis:

Both treatment groups exhibited good tolerability during the 4-week study period. Minor side effects were observed: burning in four and burning-itching in one patient in the sample group within the first hour of application. In the positive control group, one patient reported dry skin. No other adverse events were noted during the study. The frequency of complications was comparable between groups based on chi-square analysis (p = 0.083). All observed side effects were mild and transient, resolving within two hours. Patient satisfaction with the drug formulation was 100% in both groups. Chi-square analysis revealed a statistically significant difference in patient-reported improvement between groups (p = 0.000), indicating higher satisfaction with the combination therapy (clove oil-curcumin gel and clindamycin gel) compared to clindamycin gel alone. Clinical outcomes demonstrated superior recovery in the combination therapy group. 62/96 patients in the clindamycin gel group and 100% of patients in the combination therapy group showed improvement. Among the 27 patients, 10 (27.04%) experienced excellent recovery, with 9 from the combination therapy group and 1 from the clindamycin gel group.

Tables 5-6 show the treatment groups and the therapeutic response of drug regimens for them. The clinical study of designed formulations was summarized in these tables.

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Table 5. Treatment groups and responses to the therapeutic regimens

	Clindamycin gel treatment regimen (Standard deviation ± mean)		Clindamycin regi	rcumin gel + gel treatment men iation ± mean)
-	Before intervention	After intervention	Before intervention	After intervention
Number of total acne	11.25±11.26	7.07±8.72	15.11±13.93	4.67±6.18
lesions	P-value (comparing with before intervention) =0.000		P-value (comparing with before intervention) =0.000	
Number of	6.15±10.95	4.78±8.29	7.37±13.46	3.15±5.83
Comedones	P-value (comparing with before intervention) =0.011		P-value (comparing with before intervention) =0.005	
Number of acne	4.78±3.18	2.26±2.21	6.93±3.98	1.52±1.42
papules	P-value (comparing with before intervention) =0.000		P-value (comparing with before intervention) =0.000	
Number of pustules	0.33±0.62	0.04±0.19	0.81±1.04	0±0.00
-	P-value (comparing with before intervention) =0.033		P-value (comparing with before intervention) =0.002	
Severity of acne	8.52±6.33	4.72±4.85	12.24±8.07	3.093±3.41
according to ASI index	P-value (comparing with before intervention) =0.000		P-value (comparing with before intervention) =0.000	

Table 6. Statistical analysis of therapeutic responses to the drug regimen

	The amount of changes compared to before the intervention		
_	Clindamycin gel	Clove oil-curcumin gel + Clindamycin gel	
Number of total acne lesions	7.07-11.25=-4.19	4.67-15.11=-10.44	
_	P-value (comparing b	etween two groups) =0.000	
Number of Comedones	-1.37	-4.22	
_	P-value (comparing b	etween two groups) =0.375	
Number of acne papules	-2.52	-5.41	
	P-value (comparing b	etween two groups) =0.000	
Number of pustules	-0.29	-0.81	

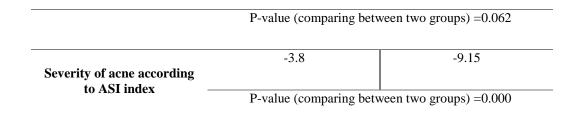


Figure 2 shows the efficacy results of prepared formulation (C2 Gel) on different symptoms.

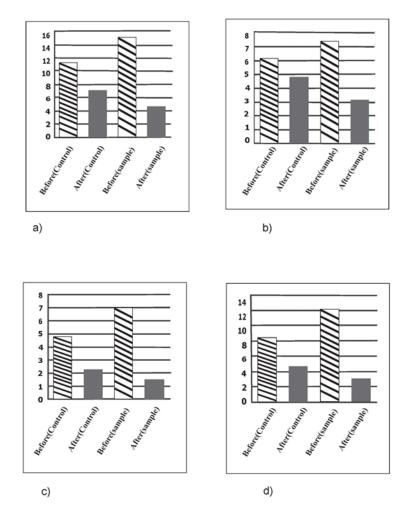


Figure 2. Efficacy results of prepared formulation (C2 Gel) on different symptoms including total acne lesions (a), comedones (b), pustules (c) and papules (d).

The following section discusses the above outcomes in the context of previous researches as well as potential clinical relevance. Acne vulgaris is a chronic inflammatory skin condition affecting the pilosebaceous units. Contributing factors include increased sebum production, follicular hyperkeratosis (blockage of the sebum secretion pathway), *P. acnes* overgrowth, and inflammatory processes. Studies suggest that IL-1 $\beta$ , Th2 cells, and IL-17 play a key role in acne-related inflammation.<sup>24</sup> Furthermore, T lymphocyte accumulation around the follicle is observed during comedone formation, with *P. acnes* playing a primary role in developing inflammatory lesions. Mediators such as prostaglandins, macrophages, cytokines, and interleukins, particularly IL-1 $\alpha$ , IL-1 $\beta$ , and IL-8, are critical contributors to the inflammatory response.<sup>25, 26</sup>

Alternative therapeutic approaches as we did in present study was attractive for researchers due to antibiotic resistance and possible teratogenic and other adverse health effects associated with conventional treatments. Isotretinoin is associated with significant adverse effects, including teratogenicity, with reported risks of 20–35% for major congenital anomalies.<sup>36</sup>

Based on this concept Clove oil and curcumin have garnered increasing interest for their synergistic potential anti-bacterial and anti-inflammatory properties in acne treatment.

Studies have demonstrated the bacteriostatic and bactericidal effects of clove oil against various bacterial strains, including *P. acnes* and *S. epidermidis* (Lamlertthon et al., 2007). Yuji Fu et al. (2009) proposed a unique mechanism of action for clove essential oil, involving damage to the bacterial cell wall and inhibition of protein synthesis (kDa-18 and kDa-33) in *P. acnes*, ultimately leading to bacterial death. Horover in vitro studies suggest that clove extracts or eugenol at non-toxic concentrations can significantly suppress the production of NF-mediated inflammatory factors (TNF-α, IL-1β, IL-6, and IL-8) and matrix metalloproteinase-9 by *P. acnes*-stimulated THP-1 cells, indicating potential for treating acne-induced skin inflammation (Tsai et al., 2017). On the other side Saeed Ta et al. (2020) found that a clove oil emulsion exhibited anti-inflammatory activity in vivo, reducing inflammation and promoting wound healing in carrageenan-induced paw edema and incisional wound models in rats.<sup>27</sup>

Misar et al. (2020) reported that a formulation containing 1% clove oil displayed the strongest antimicrobial activity against acne-associated microorganisms, including *P. acnes, S. epidermidis, Staphylococcus aureus*, and *Candida albicans*. <sup>28</sup>Liu et al. (2013) demonstrated that a combination of 0.43 μg/mL curcumin and 17.2 μg/mL lauric acid could inhibit 50% of P. acnes growth, suggesting its potential application in treating *P. acnes*-related diseases. <sup>16</sup>C. Xu et al. (2019) revealed that *P. acnes* induces the secretion of inflammatory cytokines (TNF-α, IL-8, and IL-6) by monocytes and keratinocytes through the Toll-like receptor (TLR) pathway. They further demonstrated the anti-inflammatory effects of curcumin by downregulating TLR2 expression, inhibiting IL-6 production, and blocking TLR2-NF-κB signaling. A study by Mr. Zaman et al. (2013) evaluated the antioxidant activity of a 5% turmeric rhizome methanolic extract cream formulation using the DPPH method. The study also investigated the effect of the cream on sebum secretion in 13 human volunteers using a sebometer. The results indicated that the turmeric extract could potentially reduce sebum secretion in individuals with acne. <sup>19</sup>

Sumit et al. (2018) investigated liposomal gel formulations containing curcumin, lauric acid, azithromycin, and a combination of curcumin and lauric acid. The liposomal gel formulation with a 1:1 ratio of curcumin and lauric acid demonstrated the strongest antibacterial activity against *P. acnes* based on the agar diffusion method. In vivo

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studies using a mouse ear model revealed that the curcumin-lauric acid combination gel significantly reduced comedones and cytokine levels (TNF- $\alpha$  and IL-1 $\beta$ ) compared to the placebo group.

Alihosseini et al. (2016) highlighted the potential role of turmeric and its active ingredient, curcumin, as a natural antioxidant in wound healing due to its ability to inhibit bacteria growth as well as healing agent.<sup>37</sup> Previous studies indicates that herbal treatments can serve as effective alternatives or adjuncts to traditional therapies for both mild and moderate acne vulgaris.<sup>38-40</sup> This study investigated the antimicrobial activity of clove oil and curcumin against *P. acnes* and *S. epidermidis*. Semi solid preparations were prepared and applied on the skin of volunteers suffering from acne, the synergic effect of the formulations with clindamycin gel was demonstrated.

#### Conclusion

Acne vulgaris is a chronic inflammatory condition involving multiple pathogenic factors including P. acnes, immune responses, and excessive sebum production. Concerns over antibiotic resistance and the side effects of conventional treatments like isotretinoin have led to growing interest in herbal alternatives.<sup>37-44</sup> Clove oil and curcumin have demonstrated strong antibacterial and anti-inflammatory effects against *P. acnes* and *S. epidermidis*, through mechanisms such as disruption of bacterial membranes and suppression of pro-inflammatory cytokines. This study explored their combined efficacy in topical formulations, highlighting their potential as complementary therapies in acne management. The results indicate that the combination of clindamycin gel with a clove oil-curcumin formulation significantly reduced acne lesions, papule count, and ASI scores compared to clindamycin alone. These findings support the potential of this combination therapy as a more effective approach for acne management. Further large-scale, controlled clinical studies are needed to validate these outcomes.

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#### **Ethical Approval**

The study was approved by the Ethic Committee of Islamic Azad University. Code: IR.IAU.PS.REC.1400.212.

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# The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final form.

#### References:

- 1. Tahir CM. Pathogenesis of acne vulgaris: simplified. JPAD. 2010; 20:93-7.
- 2. Habif TP. Clinical dermatology. 6th ed. Elsevier Health Sciences; 2015. p. 20–35.
- 3. Eyüboglu M, Kalay I, Eyüboglu D. Evaluation of adolescents diagnosed with acne vulgaris for quality of life and psychosocial challenges. Indian J Dermatol. 2018; 63(2):131-5. doi:10.4103/ijd.IJD\_671\_16
- 4. Lukaviciute L, Navickas P, Navickas A, Grigaitiene J, Ganceviciene R, Zouboulis CC. Quality of life, anxiety prevalence, depression symptomatology and suicidal ideation among acne patients in Lithuania. J Eur Acad Dermatol Venereol. 2017; 31(11):1900-6. doi:10.1111/jdv.14477
- 5. Vasam M, Korutla S, Bohara RA. Acne vulgaris: A review of the pathophysiology, treatment, and recent nanotechnology based advances. Biochem Biophys Rep. 2023; 36:101578. doi:10.1016/j.bbrep.2023.101578
- 6. Clark C. Acne-general practice management. The Practitioner. 1993; 237(1523):160-4.
- 7. Leyden JJ, McGinley KJ, Cavalieri S, Webster GF, Mills OH, Kligman AM. Propionibacterium acnes resistance to antibiotics in acne patients. J Am Acad Dermatol. 1983;8(1):41-5. doi:10.1016/S0190-9622(83)70005-8
- 8. Coskey R. Side effects of antibiotic therapy in acne vulgaris. Cutis. 1975; 15(5):710–2.
- 9. Milstone EB, McDonald AJ, Scholhamer CF. Pseudomembranous colitis after topical application of clindamycin. Arch Dermatol. 1981; 117(3):154-5. doi:10.1001/archderm.1981.01650030032015
- 10. Lammer EJ, Chen DT, Hoar RM, Agnish ND, Benke PJ, Braun JT, et al. Retinoic acid embryopathy. N Engl J Med. 1985; 313(14):837-41. doi:10.1056/NEJM198510033131401
- 11. Lamlertthon S, Luangnarumitchai S, Tiyaboonchai W. Antimicrobial activity of essentials oils against five strains of Propionibacterium acnes. Mahidol Univ J Pharm Sci. 2007; 34(1-4):60-4.
- 12. Fu Y, Chen L, Zu Y, Liu Z, Liu X, Liu Y, et al. The antibacterial activity of clove essential oil against Propionibacterium acnes and its mechanism of action. Arch Dermatol. 2009; 145(1):86-8. doi:10.1001/archdermatol.2008.549
- 13. Singh V, Pahuja C, Ali M, Sultana S. Analysis and bioactivities of essential oil of the flower buds of Syzygium aromaticum. J Med Plants Stud. 2018; 6(6):79–83.
- 14. Tsai T-H, Huang W-C, Lien T-J, Huang Y-H, Chang H, Yu C-H, et al. Clove extract and eugenol suppress inflammatory responses elicited by Propionibacterium acnes in vitro and in vivo. Food Agric Immunol. 2017; 28(5):916-31. doi:10.1080/09540105.2017.1320357
- 15. Sugihartini N, Lestari G, Yuliani S. Anti-inflammatory Capacity of Clove Bud (Syzygium aromaticum) Essential Oil in O/W and W/O Creams. Pharmaciana. 2019; 9(1):109-18. doi:10.12928/pharmaciana.v9i1.10023
- 16. Liu C-H, Huang H-Y. In vitro anti-propionibacterium activity by curcumin containing vesicle system. Chem Pharm Bull. 2013; 61(4):419-25. doi:10.1248/cpb.c12-01043

# The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final form.

- 17. Liu C-H, Huang H-Y. Antimicrobial activity of curcumin-loaded myristic acid microemulsions against Staphylococcus epidermidis. Chem Pharm Bull. 2012; 60(9):1118–24. doi:10.1248/cpb.c12-00220
- 18. Xu C, Zeng R, Chen Q, Du L, Tong J, He Y, et al. Curcumin suppresses interleukin-6 production in THP-1 monocytes induced by Propionibacterium acnes extracts via downregulation of Toll-like receptor 2 expression and the nuclear factor kappa B pathway. Br J Dermatol. 2019; 181(6):1320-2. doi:10.1111/bjd.18228
- 19. Zaman S, Akhtar N. Effect of turmeric (Curcuma longa Zingiberaceae) extract cream on human skin sebum secretion. Trop J Pharm Res. 2013; 12(5):665-9. doi:10.4314/tjpr.v12i5.1
- 20. Madan S, Nehate C, Barman TK, Rathore AS, Koul V. Design, preparation, and evaluation of liposomal gel formulations for treatment of acne: in vitro and in vivo studies. Drug Dev Ind Pharm. 2019; 45(3):395-404. doi:10.1080/03639045.2018.1546310
- 21. Lawhavinit O-a, Kongkathip N, Kongkathip B. Antimicrobial activity of curcuminoids from Curcuma longa L. on pathogenic bacteria of shrimp and chicken. Agriculture and Natural Resources. 2010; 44(3):364-71.
- 22. Niamsa N, Sittiwet C. Antimicrobial activity of Curcuma longa aqueous extract. J Pharmacol and Toxicol. 2009; 4(4): 173-177. doi:10.3923/jpt.2009.173.177
- 23. Jagetia GC, Rajanikant GK. Curcumin treatment enhances the repair and regeneration of wounds in mice exposed to hemibody γ-irradiation. *Plast Reconstr Surg.* 2005; 115(2):515–28. doi:10.1097/01.PRS.0000148372.75342.D9
- 24. Suh D, Kwon H. What's new in the physiopathology of acne? Br J Dermatol. 2015; 172:13–9. doi:10.1111/bjd.13634
- 25. Farrar MD, Ingham E. Acne: inflammation. Dermatol Clin. 2004; 22(5):380-4. doi:10.1016/j.clindermatol.2004.03.006
- 26. Barak IV, Halperin T, Birkenfeld S, Ginsburg I. PADMA-28, a Tibetan herbal preparation is an inhibitor of inflammatory cytokine production. Eur Cytokine New. 2004; 15(3):203-9.
- 27. Saeed TA, Osman OA, Amin AE, El Badwi SM. Safety assessment and potential anti-inflammatory effect of ethanolic extract of Syzygium aromaticum in albino rats. Adv Biosci Biotechnol. 2017; 8(11):411. doi:10.4236/abb.2017.811030
- 28. Misar K, Kulkarni S, Gurnule W. Formulation and evaluation of antiacne cream by using Clove oil. Mater Today Proc. 2020; 29:1251-8. doi:10.1016/j.matpr.2020.06.106
- 29. Balouiri M, Sadiki M, Ibnsouda SK. Methods for in vitro evaluating antimicrobial activity: A review. J Pharm Biomed Anal. 2016; 6(2):71–9. doi:10.1016/j.jpha.2015.11.005
- 30. Donskey CJ. Antibiotic regimens and intestinal colonization with antibiotic-resistant gram-negative bacilli. Clin Infect Dis. 2006; 43:62-9.
- 31. United States Pharmacopeial Convention, editor. The United States Pharmacopeia. 2000.
- 32. Sreevidya V. An overview on emulgel. Int J Pharm Phytopharm Res. 2019;9(1):92-7.

#### **Accepted Manuscript (unedited)** The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final form.

- 33. Lasic DD. Liposomes: from physics to applications. Elsevier. 1993.
- 34. Wilkinson J, Moore R. Harry's Cosmetology. 7th ed. George-Godwin Publ.; 1982.
- 35. Mortazavi M, GKM, Mahboubi A, Mortazavi A. Review on Pharmaceutics. Etminan; 1396 (Persian calendar).
- 36. Zhao R, Guo H, Yan T, Li J, Xu W, Deng Y, et al. Fabrication of multifunctional materials based on chitosan/gelatin incorporating curcumin-clove oil emulsion for meat freshness monitoring and shelf-life extension. Int J Biol Macromol. 2022; 223:837–50. doi:10.1016/j.ijbiomac.2022.10.271
- 37. Alihosseini F, Azarmi S, Ghaffari S, Haghighat S, Sorkhabadi SMR. Synergic antibacterial effect of curcumin with ampicillin; free drug solutions in comparison with SLN dispersions. Adv Pharm Bull. 2016; 6(3):461. doi:10.15171/apb.2016.060
- 38. Proença AC, Luis A, Duarte AP. The role of herbal medicine in the treatment of acne vulgaris: a systematic review of clinical trials. Evid Based Complement Alternat Med. 2022; 2022:2011945. doi:10.1155/2022/2011945
- 39. Nojek P, Pawlik W, Zimonczyk M, Zawół M, Błaszczyński G, Nowotarska A. Effectiveness of different acne treatment methods: a comparison of pharmacological, procedural and natural therapies. J Educ Health Sport. 2024; 71:55977. doi:10.12775/JEHS.2024.71.55977
- 40. Sinha P, Srivastava S, Mishra N, Yadav NP. New perspectives on antiacne plant drugs: contribution to modern therapeutics. Biomed Res Int. 2014; 2014:301304. doi:10.1155/2014/301304.
- 41. Draghici CC, Miulescu RG, Petca RC, Petca A, Dumitrascu MC, Sandru F. Teratogenic effect of isotretinoin in both fertile females and males. Exp Ther Med. 2021; 21(5):534. doi:10.3892/etm.2021.9966
- 42. Nawaz A, Farid A, Safdar M, Latif MS, Ghazanfar S, Akhtar N, et al. Formulation development and ex-vivo permeability of curcumin hydrogels under the influence of natural chemical enhancers. Gels. 2022;8(6):384. doi:10.3390/gels8060384
- 43. Bendary MM, Elmanakhly AR, Mosallam FM, Alblwi NAN, Mosbah RA, Alshareef WA, et al. Boosting the anti-Helicobacter efficacy of azithromycin through natural compounds: Insights from in vitro, in vivo, histopathological, and molecular docking investigations. Helicobacter. 2024; 29(4):e13110. doi:10.1111/hel.13110
- 44. Jourghanian P, Ghaffari S, Ardjmand M, Haghighat S, Mohammadnejad M. Sustained release curcumin loaded solid lipid nanoparticles. Adv Pharm Bull. 2016; 6(1):17. doi:10.15171/apb.2016.04