

A Molecular Study on Drug Delivery System Based on Carbon Nanotube Compared to Silicon Carbide Nanotube for Encapsulation of Platinum-Based Anticancer Drug

Zahra Khatti¹, Seyed Majid Hashemianzadeh², Seyed Ali Shafiei^{3*}

¹ Department of Chemistry, Iran University of Science and Technology, Tehran, Iran.

² Molecular Simulation Research Laboratory, Department of Chemistry, Iran University of Science and Technology, Tehran, Iran.

³ Neurology and Neuroscience Research Center, Qom University of Medical Sciences, Qom, Iran.

Article info

Article History:

Received: 13 November 2017

Revised: 25 February 2018

Accepted: 26 February 2018

ePublished: 18 March 2018

Keywords:

- Molecular Dynamic Simulation
- Carbon nanotube
- Silicon Carbide nanotube
- Platinum-based anticancer drug

Abstract

Purpose: Drug delivery has a critical role in the treatment of cancer, in particular, carbon nanotubes for their potential use in various biomedical devices and therapies. From many other materials which could be more biocompatible and biodegradable and which could form single-walled nanotubes, silicon carbide was selected.

Methods: To compare two drug delivery systems based on single-walled nanotubes, molecular dynamic simulations were applied and encapsulation behavior of the drug carboplatin was investigated inside the silicon carbide nanotube and the carbon nanotube.

Results: Localization of the carboplatin inside the nanotubes indicated that the carboplatin moves throughout the tubes and possesses a greater probability of finding the drug molecule along the nanotubes in the first quarter of the tubes. The energy analysis exhibited the lowest free energy of binding belongs to the encapsulation of the drug carboplatin in the silicon carbide nanotube, about -145 Kcal/mol.

Conclusion: The results confirmed that the silicon carbide nanotube is a more suitable model than the carbon nanotube for drug delivery system based on nanotubes as a carrier of platinum-based anticancer drugs.

Introduction

Carbon nanotubes (CNTs), due to their unique atomic configuration, mechanical, optical and electronic properties, have been envisioned in designing biomedical devices and therapies on novel delivery platforms.¹⁻⁵ The physicochemical versatility of carbon nanotubes is related to their high surface-area-to-volume ratios and facile functionalization along the nanotube axis and a great inner content that can be filled with the desired drug molecules.⁶⁻⁸ Additionally, the ability of single-walled carbon nanotubes (SWCNT) to incorporate inside cells has been proven, independent of cell type and functional groups linked to the nanotubes.^{9,10} In vivo assays have demonstrated that SWCNTs as carriers have no obvious toxicity,¹¹ and in animal models have demonstrated the efficacy of drug-loaded CNTs through targeting tumors.^{6,12} Whereas water is the main component in biological systems and CNTs are hydrophobic, heterogeneous CNTs could be considered including silicon carbide nanotube (SiCNT) in aqueous media.¹³ In previous studies, several advantages have been shown for SiCNTs compared to CNTs. First, there is the relative stability increase from the CNTs to SiCNTs when the ratio of Si over C is 50:50, due to alternative sp² and sp³ hybridization bond structures that

are more stable than a smooth-walled tube.¹⁴ Additionally, the external surface of SiCNTs has higher reactivity than that of CNTs to facilitate aimed side wall functionalization.^{15,16} Furthermore, the experimental results prove the biocompatibility of silicon nanotubes and hence an alternative option for applications in nanomedicine.^{17,18} Platinum-based anticancer drugs are used to treat many types of solid tumors via binding to DNA and inducing cellular apoptosis, despite their adverse side effects,^{19,20} so that many of these side effects for healthy cells can be greatly reduced by nanoscale drug delivery. A fast and reliable tool to evaluate theoretically such systems is molecular modeling, which could interpret the details of the interaction between the drug and both the DNA and the nanostructures.²¹ On the other hand, our previous study allowed the assessment of a drug delivery system caused by another nanotube apart from CNTs, as carrier on theoretical level for the first time.²² Therefore, in the present study, molecular dynamics (MD) simulation was employed to investigate another so-named nanotube SiCNT as delivery system compared to CNT. Carboplatin (diammineplatinum(II) cyclobutane-1,1-dicarboxylate) was selected as a drug model because it

*Corresponding author: Seyed Ali Shafiei, Tel: +98 2533209121, Fax: +98 2533209127, Email: sashafiei@muq.ac.ir

©2018 The Authors. This is an Open Access article distributed under the terms of the Creative Commons Attribution (CC BY), which permits unrestricted use, distribution, and reproduction in any medium, as long as the original authors and source are cited. No permission is required from the authors or the publishers.

encompasses fewer adverse effects and has greater water solubility than other platinum agents.²³⁻²⁵ Our simulations have been performed to assess the encapsulation behavior and localization of the anticancer drug carboplatin inside pristine CNT and SiCNT. For this purpose, the placement of the drug inside nanotubes and the free energy calculations were implemented to determine the preference between the two predicted drug delivery systems.

Materials and Methods

Systems preparation

In the present work, the zigzag open-ended single-walled nanotubes with 14 Å diameter and 40 Å length were considered, so (18, 0) carbon nanotube and (14, 0) silicon carbide nanotube were applied. Molecular dynamic simulations were applied for two systems; both of them containing a carboplatin molecule at a position along the CNT and SiCNT z-axis, i.e., drug_CNT and drug_SiCNT. Two systems were solvated in an aqueous solution. The parameters of CNT were modeled using the AMBER99SB force field.^{26,27} All the Lennard-Jones parameters and partial charge values for the carbon and silicon atoms for SiCNT were obtained from density functional theory calculations, summarized in Table 1.^{14,28} Other parameters were taken from the DREIDING force field and used in MD simulations. The structure of carboplatin was obtained from our previous works,^{22,29} and the parameters were identified from the literature and the GAFF,^{30,31} additionally, the carboplatins' partial charges using the RESP module of AMBER 12 were calculated. The desired structures were immersed in a periodic water box of octagonal shape with a minimal distance of 12 Å from the system surface. Two systems were solvated with a TIP3P solvation model.³²

Table 1. Lennard-Jones parameters and partial charges for SiCNT atoms

Atom	ϵ (kcal/mol)	σ (Å)	Atomic Charge (e)
C	0.086	3.4	0.45
Si	0.469	3.7364	-0.45

Molecular dynamics simulations

Molecular dynamics simulations were performed by employing the AMBER 12 simulation package.³³ All simulations were conducted under the isothermal-isobaric ensemble at 1 atm and 300 K using the SANDER module in the AMBER 12 program. Constant temperature and pressure were maintained using Langevin dynamics.³⁴ Bond lengths containing hydrogen atoms were constrained using the SHAKE algorithm.³⁵ Periodic boundary conditions were applied and the long-range electrostatic interactions were handled with the particle mesh Ewald method; the nonbonded interactions were treated with cutoff at 12Å.³⁶ Each simulation in energy minimization stage included 5000 steps for solvent and 5000 steps for solute relaxation. All systems

were then heated from 0K to 300K for 120 ps and equilibrated at 300K for 200 ps. The production stages were run for 10 ns with a time step of 2 fs, and the trajectories for structural coordinates were saved every 1 ps for data analysis. Configuration analysis of the trajectories was performed with the ptraj module included in AmberTools14. Additionally, binding free energies were evaluated using the MM/PBSA (molecular mechanics/Poisson-Boltzmann surface area) method by previous experiment.^{29,37} The analysis of the interaction energies between the nanotubes and the drug was accomplished with AmberTools14 and implementation of MMPBSA.py script.

Results

The results from MD simulations of the two systems revealed that throughout the simulation time, both encapsulated carboplatin molecules resided inside the CNT and SiCNT cavity. Time-averaged radial distribution functions (RDF) were calculated to assess the localization of carboplatin inside the nanotubes (Figure 1), also RMSD plots were illustrated the drug movement inside the carbon and silicon carbide nanotubes (Figure 2). Moreover, MM/PBSA analysis of the trajectories was performed to estimate the binding free energy of the drug_CNT and the drug_SiCNT. Binding and absolute free energies of molecules are evaluated using this attractive method:

$$\Delta G_{binding} = [G_{complex}] - [G_{drug}] - [G_{CNT}]$$

Additionally, separation of the total free energy of binding into the van der Waals, electrostatic, and solute-solvent interactions, and discussion of each of the terms can be allowed and is calculated from:

$$\Delta G_{tot} = \Delta E_{MM} + \Delta G_{solv} - T\Delta S$$

$$\Delta E_{MM} = E_{int} + E_{vdW} + E_{ele}$$

$$\Delta G_{solv}^{PBSA} = \Delta G_{solv}^{nonpolar} + \Delta G_{solv}^{electrostatic}$$

where the molecular mechanics energy of the molecule (ΔE_{MM}) includes the van der Waals (E_{vdW}), internal energy (bonds, angles and dihedrals) (E_{int}) and the electrostatic energy (E_{ele}) terms. The solvation energy (G_{solv}) containing the polar and nonpolar parts is obtained from an implicit solvent description. Table 2 summarizes all energy terms for the two simulated systems in order to evaluate binding free energy using the MM/PBSA method.

Discussion

Localization of carboplatin inside the CNT and SiCNT

To assess the localization of carboplatin inside the nanotubes, time-averaged radial distribution functions (RDF) were calculated. Figure 1a illustrates the RDF plot between the carboplatin center of mass and the carbon atoms of the CNT. In addition, Figure 1b illustrates the RDF plot between the carboplatin center of mass and the

carbon and silicon atoms of the SiCNT. Both plots indicate that the drug moves throughout the tubes and there is a greater probability of finding the drug along the CNT and SiCNT in the first quarter of the tubes. Since drug exposure at the end of the nanotube is an important

option for the drug molecules to release,³⁸ the encapsulation and the preferential position of the carboplatin inside the tubes have a significant contribution in drug delivery.

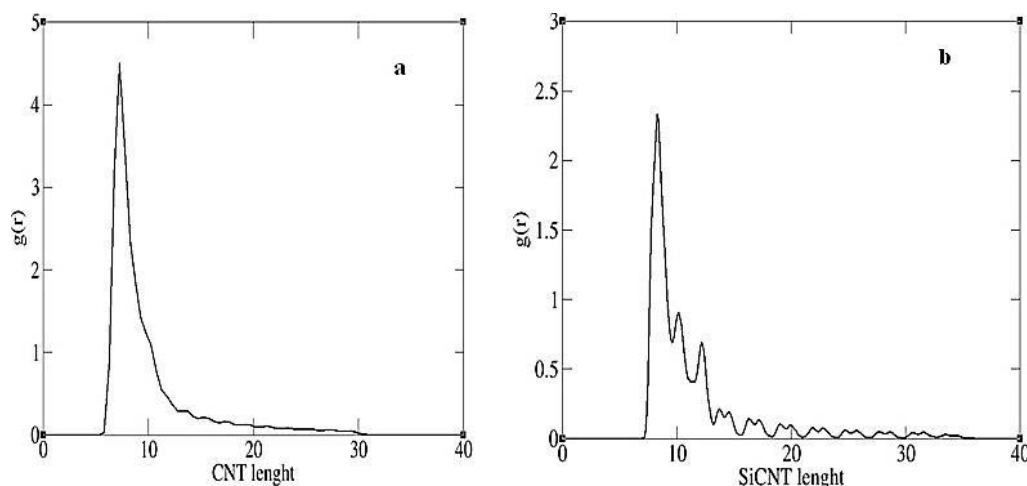


Figure 1. The RDF plot between the carboplatin center of mass and (a) the carbon atoms of the CNT, (b) the carbon and silicon atoms of the SiCNT

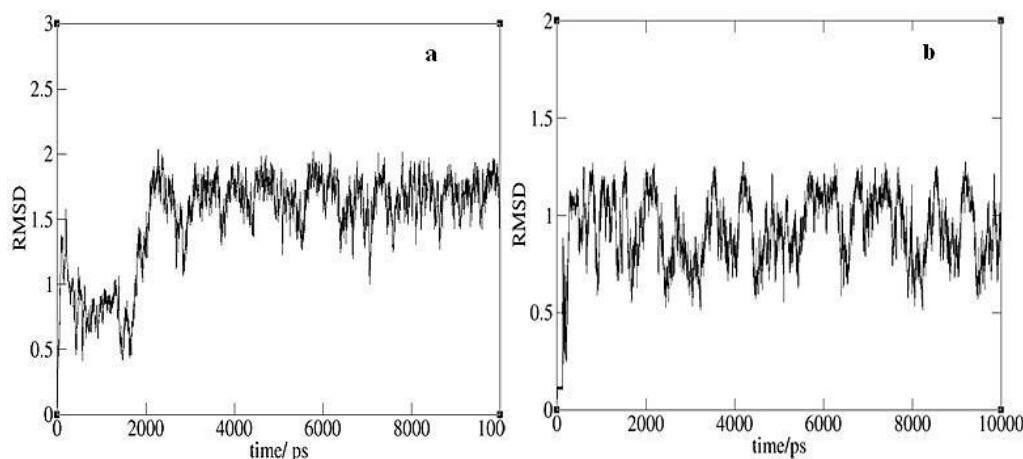


Figure 2. RMSD plots of encapsulation of the carboplatin inside the (a) CNT, (b) SiCNT.

Energy analysis

With regard to RMSD plots of drug movement inside the carbon and silicon carbide nanotubes (Figure 2), it is obvious that the two systems have stable equilibration, so it is possible to assess the thermodynamic properties of systems by sampling in simulation time and achieving the localization and the encapsulation behavior of the carboplatin inside the CNT and SiCNT. From the results of energy terms in Table 2, the total binding free energy in the drug_CNT system with an average of about $-46 \text{ kcal mol}^{-1}$ and the drug_SiCNT with $-145 \text{ kcal mol}^{-1}$ remains constant. It is derived that the dominant contribution of interactions is related to nonbonded van der Waals energy; evidently indicating that encapsulation of carboplatin inside the silicon carbide nanotube has stronger vdW interactions. The solvation free energies ($G_{\text{solv}}^{\text{PBSA}}$) have a positive and small contribution of the

binding free energy for both systems (Table 2), due to the hydrophobic structures of nanotubes; however, this is slightly reduced in the drug_SiCNT system. Whereas the partial charge on carbon atoms of CNT is zero, this reduction of solvation free energies can be related to partial charges on carbon and silicon atoms of the silicon carbide nanotube's structure. Regarding these results, the larger van der Waals value belongs to the drug_SiCNT, so the carboplatin drug has stronger interaction inside the SiCNT than it does inside the carbon nanotube. Comparison of these energy profiles reveals that the carboplatin molecule prefers to spend more time inside the SiCNT until it reaches the target cell. Consequently, using SiCNT nanotube as a platinum drug carrier can be the more suitable option for drug delivery with this type of nanostructure.

Table 2. Energy values for encapsulation of drug inside the nanotubes

Complex	drug_CNT	drug_SiCNT
E_{vdW}	-50.7566	-147.1443
E_{ele}	-0.9297	-2.9051
G_{solv}^{ele}	7.0830	6.0232
$G_{solv}^{nonpolar}$	-1.7366	-1.0369
E_{MM}	-51.6863	-150.0495
G_{solv}^{PBSA}	5.3464	4.9863
$\Delta G_{binding}$	-46.3399	-145.0631

All values in this table are in kcal/mol.

Conclusion

Molecular dynamic simulations to investigate the encapsulation behavior of the drug carboplatin inside the silicon carbide nanotube were applied as a comparison with the carbon nanotube as an anticancer drug delivery system based on single-walled nanotubes. The RDF plots show the localization of carboplatin inside the nanotubes, indicating that the drug moves throughout the tubes and has a greater probability of finding the carboplatin along the CNT and SiCNT in the first quarter of the tubes. Additionally, the binding free energy profiles in the encapsulation of drug inside both systems were investigated. The results confirmed that the appropriate drug delivery system for platinum drug is the use of SiCNTs to CNTs. Since the free energy of binding in the silicon carbide nanotube is about three times that of the carbon nanotube, the length of time remaining for the drug in this system will be greater, and the probability of releasing the drug will be less than with carbon nanotube, before reaching the target cells.

Acknowledgments

The authors give special thanks to S. Skies for editing this manuscript.

Ethical Issues

Not applicable.

Conflict of Interest

The authors declare no conflict of interests.

References

- Baughman RH, Zakhidov AA, de Heer WA. Carbon nanotubes--the route toward applications. *Science* 2002;297(5582):787-92. doi: 10.1126/science.1060928
- Sharma A, Jain N, Sareen R. Nanocarriers for diagnosis and targeting of breast cancer. *BioMed Res Int* 2013;2013:960821. doi: 10.1155/2013/960821
- Wong CH, Vijayaraghavan V. Compressive characteristics of single walled carbon nanotube with water interactions investigated by using molecular

dynamics simulation. *Phys Lett A* 2014;378(5-6):570-6. doi: 10.1016/j.physleta.2013.12.026

- Adeli M, Hakimpoor F, Ashiri M, Kabiri R, Bavadi M. Anticancer drug delivery systems based on noncovalent interactions between carbon nanotubes and linear-dendritic copolymers. *Soft Matter* 2011;7(8):4062-70. doi: 10.1039/c0sm01550d
- Madani SY, Mandel A, Seifalian AM. A concise review of carbon nanotube's toxicology. *Nano Reviews* 2013;4. doi: 10.3402/nano.v4i0.21521
- McDevitt MR, Chattopadhyay D, Kappel BJ, Jaggi JS, Schiffman SR, Antczak C, et al. Tumor targeting with antibody-functionalized, radiolabeled carbon nanotubes. *J Nucl Med* 2007;48(7):1180-9. doi: 10.2967/jnumed.106.039131
- Liu Z, Chen K, Davis C, Sherlock S, Cao Q, Chen X, et al. Drug delivery with carbon nanotubes for in vivo cancer treatment. *Cancer Res* 2008;68(16):6652-60. doi: 10.1158/0008-5472.CAN-08-1468
- Castle AB, Gracia-Espino E, Nieto-Delgado C, Terrones H, Terrones M, Hussain S. Hydroxyl-functionalized and N-doped multiwalled carbon nanotubes decorated with silver nanoparticles preserve cellular function. *ACS Nano* 2011;5(4):2458-66. doi: 10.1021/nn200178c
- Kang B, Chang S, Dai Y, Yu D, Chen D. Cell response to carbon nanotubes: Size-dependent intracellular uptake mechanism and subcellular fate. *Small* 2010;6(21):2362-6. doi: 10.1002/sml.201001260
- Kostarelos K, Lacerda L, Pastorin G, Wu W, Wieckowski S, Luangsivilay J, et al. Cellular uptake of functionalized carbon nanotubes is independent of functional group and cell type. *Nat Nanotechnol* 2007;2(2):108-13. doi: 10.1038/nnano.2006.209
- Schipper ML, Nakayama-Ratchford N, Davis CR, Kam NW, Chu P, Liu Z, et al. A pilot toxicology study of single-walled carbon nanotubes in a small sample of mice. *Nat Nanotechnol* 2008;3(4):216-21. doi: 10.1038/nnano.2008.68
- Liu Z, Cai W, He L, Nakayama N, Chen K, Sun X, et al. In vivo biodistribution and highly efficient tumour targeting of carbon nanotubes in mice. *Nat Nanotechnol* 2007;2(1):47-52. doi: 10.1038/nnano.2006.170
- Lee C, Drelich J, Yap Y. Superhydrophobicity of boron nitride nanotubes grown on silicon substrates. *Langmuir* 2009;25(9):4853-60. doi: 10.1021/la900511z
- Mavrandonakis A, Froudakis GE, Schnell M, Mühlhäuser M. From pure carbon to silicon-carbon nanotubes: An Ab-initio study. *Nano Lett* 2003;3(11):1481-4. doi: 10.1021/nl0343250
- Wu JJ, Guo GY. Optical properties of SiC nanotubes: An ab initio study. *Phys Rev B* 2007;76(3):035343.
- Miyamoto Y, Yu BD. Computational designing of graphitic silicon carbide and its tubular forms. *Appl Phys Lett* 2002;80(4):586-8. doi: 10.1063/1.1445474

17. Mu C, Zhao Q, Xu D, Zhuang Q, Shao Y. Silicon nanotube array/gold electrode for direct electrochemistry of cytochrome c. *J Phys Chem B* 2007;111(6):1491-5. doi: 10.1021/jp0657944
18. Sahu T, Ghosh B, Pradhan SK, Ganguly T. Diverse role of silicon carbide in the domain of nanomaterials. *Int J Electrochem* 2012;2012:271285. doi: 10.1155/2012/271285
19. Wang D, Lippard SJ. Cellular processing of platinum anticancer drugs. *Nat Rev Drug Discov* 2005;4(4):307-20. doi: 10.1038/nrd1691
20. McWhinney SR, Goldberg RM, McLeod HL. Platinum neurotoxicity pharmacogenetics. *Mol Cancer Ther* 2009;8(1):10-6. doi: 10.1158/1535-7163.MCT-08-0840
21. Sargolzaei M, Nikoofard H, Afshar M. DNA binding mode and affinity of antitumor drugs of 2-aryolbenzofuran-3-ols: Molecular dynamics simulation study. *Pharm Chem J* 2016;50(3):137-42. doi: 10.1007/s11094-016-1411-4
22. Khatti Z, Hashemianzadeh SM. Boron nitride nanotube as a delivery system for platinum drugs: Drug encapsulation and diffusion coefficient prediction. *Eur J Pharm Sci* 2016;88:291-7. doi: 10.1016/j.ejps.2016.04.011
23. Kelland L. The resurgence of platinum-based cancer chemotherapy. *Nat Rev Cancer* 2007;7(8):573-84. doi: 10.1038/nrc2167
24. Arlt M, Haase D, Hampel S, Oswald S, Bachmatiuk A, Klingeler R, et al. Delivery of carboplatin by carbon-based nanocontainers mediates increased cancer cell death. *Nanotechnology* 2010;21(33):335101. doi: 10.1088/0957-4484/21/33/335101
25. Negureanu L, Salisbury FR Jr. Non-specificity and synergy at the binding site of the carboplatin-induced DNA adduct via molecular dynamics simulations of the muts α -DNA recognition complex. *J Biomol Struct Dyn* 2014;32(6):969-92. doi: 10.1080/07391102.2013.799437
26. Cornell WD, Cieplak P, Bayly CI, Gould IR, Merz KM, Ferguson DM, et al. A second generation force field for the simulation of proteins, nucleic acids, and organic molecules. *J Am Chem Soc* 1995;117(19):5179-97. doi: 10.1021/ja00124a002
27. Hummer G, Rasaiah JC, Noworyta JP. Water conduction through the hydrophobic channel of a carbon nanotube. *Nature* 2001;414(6860):188-90. doi: 10.1038/35102535
28. Taghavi F, Javadian S, Hashemianzadeh SM. Molecular dynamics simulation of single-walled silicon carbide nanotubes immersed in water. *J Mol Graph Model* 2013;44:33-43. doi: 10.1016/j.jmkgm.2013.04.012
29. Khatti Z, Hashemianzadeh SM. Investigation of thermodynamic and structural properties of drug delivery system based on carbon nanotubes as a carboplatin drug carrier by molecular dynamics simulations. *J Incl Phenom Macrocycl Chem* 2015;83(1-2):131-40. doi: 10.1007/s10847-015-0549-0
30. Yao S, Plasteras JP, Marzilli LG. A molecular mechanics amber-type force field for modeling platinum complexes of guanine derivatives. *Inorg Chem* 1994;33(26):6061-77. doi: 10.1021/ic00104a015
31. Cundari TR, Fu W, Moody EW, Slavin LL, Snyder LA, Sommerer SO, et al. Molecular mechanics force field for platinum coordination complexes. *J Phys Chem* 1996;100(46):18057-64. doi: 10.1021/jp961240x
32. Jorgensen WL, Chandrasekhar J, Madura JD, Impey RW, Klein ML. Comparison of simple potential functions for simulating liquid water. *J Chem Phys* 1983;79(2):926-35. doi: 10.1063/1.445869
33. Case D, Darden T, Cheatham III T, Simmerling C, Wang J, Duke R, et al. Amber 12. San Francisco: University of California; 2012.
34. Cerutti DS, Duke R, Freddolino PL, Fan H, Lybrand TP. Vulnerability in popular molecular dynamics packages concerning langevin and andersen dynamics. *J Chem Theory Comput* 2008;4(10):1669-80. doi: 10.1021/ct8002173
35. Miyamoto S, Kollman PA. Settle: An analytical version of the SHAKE and RATTLE algorithm for rigid water models. *J Comput Chem* 1992;13(8):952-62. doi: 10.1002/jcc.540130805
36. Essmann U, Perera L, Berkowitz ML, Darden T, Lee H, Pedersen LG. A smooth particle mesh ewald method. *J Chem Phys* 1995;103(19):8577-93. doi: 10.1063/1.470117
37. Kollman PA, Massova I, Reyes C, Kuhn B, Huo S, Chong L, et al. Calculating structures and free energies of complex molecules: Combining molecular mechanics and continuum models. *Acc Chem Res* 2000;33(12):889-97.
38. Hampel S, Kunze D, Haase D, Kramer K, Rauschenbach M, Ritschel M, et al. Carbon nanotubes filled with a chemotherapeutic agent: A nanocarrier mediates inhibition of tumor cell growth. *Nanomedicine (Lond)* 2008;3(2):175-82. doi: 10.2217/17435889.3.2.175