Advanced Pharmaceutical Bulletin

Review Article

Adv Pharm Bull, 2019, 9(3), 335-347 doi: 10.15171/apb.2019.041 https://apb.tbzmed.ac.ir



Drug Resistance and the Prevention Strategies in Food Borne Bacteria: An Update Review

Fataneh Hashempour-Baltork¹⁰, Hedayat Hosseini^{2,3*}, Saeedeh Shojaee-Aliabadi²⁰, Mohammadali Torbati⁴⁰, Adel Mirza Alizadeh¹⁰, Matin Alizadeh⁵⁰

¹Student Research Committee, Department of Food Science and Technology, National Nutrition and Food Technology Research Institute, Faculty of Nutrition Science and Food Technology, Shahid Beheshti University of Medical Sciences, Tehran, Iran. ²Department of Food Science and Technology, National Nutrition and Food Technology Research Institute, Faculty of Nutrition

Science and Food Science and Jechnology, National Nutrition and Food Jechnology Research Institute, Faculty of Nutrition Science and Food Technology, Shahid Beheshti University of Medical Sciences, Tehran, Iran.

³Food Safety Research Center, Shahid Beheshti University of Medical Sciences, Tehran, Iran.

Abstract

bacteria.

⁴Department of Food Science and Technology, Faculty of Nutrition, Tabriz University of Medical Sciences, Tabriz, Iran. ⁵Department of Clinical Sciences (Surgery), Faculty of Specialized Veterinary Sciences, Science and Research Branch, Islamic

Antibiotic therapy is among the most important treatments against infectious diseases and

has tremendously improved effects on public health. Nowadays, development in using this treatment has led us to the emergence and enhancement of drug-resistant pathogens which can

result in some problems including treatment failure, increased mortality as well as treatment

costs, reduced infection control efficiency, and spread of resistant pathogens from hospital to community. Therefore, many researches have tried to find new alternative approaches to

control and prevent this problem. This study, has been revealed some possible and effective

approaches such as using farming practice, natural antibiotics, nano-antibiotics, lactic acid

bacteria, bacteriocin, cyclopeptid, bacteriophage, synthetic biology and predatory bacteria

as alternatives for traditional antibiotics to prevent or reduce the emergence of drug resistant

Azad University, Tehran, Iran.

Article info

Article History: Received: 3 Nov. 2018 Revised: 13 May 2019 Accepted: 13 May 2019 epublished: 1 Aug. 2019

Keywords:

- Drug resistance
- Food borne pathogens
- Food safety
- Health
- Prevention strategies

Introduction

Antimicrobials have a lot of usage in medicine for human, plant and veterinary for a few decades. These substances can be used on agriculture and veterinary for different means of feed efficiency, growth improve and almost simultaneously for control, prevent and treat infections.^{1,2} Antibiotics are vital medical materials which can be natural, synthetic or semi-synthetic and can kill or interfere the growth of bacteria, and are used both in animals and humans for control or treat infections.^{1,2} In meatproducing animals, the most common problems due to antibiotics use are including bovine pneumonia, shipping fever and diarrhea,³ respiratory diseases, liver abscesses and improvement in growth.⁴ On the other hand, studies reported some safety adverse reactions for antibiotics including anaphylaxis, cardiotoxicity, nephrotoxicity, neurotoxicity and hepatotoxicity, and also documented number of hematological and gastrointestinal problems.5 Therefore, in relation to antimicrobial consumption in animal or human, some important items should be considered, including drug characterization (toxicity,

pharmacodynamics, pharmacokinetics, cost and tissue distribution), the age and immune system of animal or human and appropriate drug dose which can be achieved by antimicrobial susceptibility test.⁶ According to studies, antimicrobials can be used as growth enhancer in low subtherapuetic doses, but these doses cannot destroy the bacteria and allow them to achieve more resistance to the drug.⁷ Concerns about misuse or overuse of antimicrobials as nontherapeutic and appearance of drug resistance have arisen when antimicrobial dose increased to 100 percent in aquaculture in the 1994–2004.^{8,9} Antimicrobial resistance gointed to the situation that a microorganism shows resistance to a drug that was effective for its killing or destroying previously.¹⁰ Todays, this issue have significant effect on mortality and morbidity of humans each year

and has reported that antibiotic-resistant bacteria caused death of 700,000 people globally and has predicted that this rate tend to enhance approximately to 10 million by 2050.¹¹ Adaptation of bacteria to various environmental stresses such as antibiotics, approve that they are quite adaptive organisms.¹⁰

*Corresponding Author: Hedayat Hosseini, Tel: +98 21 22376426; Fax: +98 21 2236 0660, Email: hedayat@sbmu.ac.ir

^{© 2019} The Author (s). This is an Open Access article distributed under the terms of the Creative Commons Attribution (CC BY), which permits unrestricted use, distribution, and reproduction in any medium, as long as the original authors and source are cited. No permission is required from the authors or the publishers.

There are two types of mechanisms for creation and spreading the resistant bacteria population: vertical gene transfer and horizontal gene transfer. The former, which is also called *intrinsic* resistance, occurs in evolutionary phase and genetic errors accumulate in the plasmid or chromosome of bacterial cells. However in the horizontal gene transfer or *acquired* resistance, the exchanges are within and between bacterial species in which the organisms gain new genes on their mobile genetic elements including plasmids, insertion sequences, phage-related elements and integrons, transposons.¹²

Antibacterial resistance can be spread by food chain through direct or indirect exposure. Direct exposure occurs, following the contact of human with animal or its blood, saliva, milk, semen, feces and urine which is very simple and rapid way for spreading resistant bacteria. The indirect contact occurs, following by consumption of contaminated food products such as egg, meat and dairy products which is more complex and far-reaching pathway.¹³⁻¹⁵ The other particular transport routs are related to environment which can be the source of antibiotic-resistance genes.^{16,17} As a result, the bacteria as a reservoir of resistance genes in addition of their pathogenicity, can be a hidden hazard for public health. The appearance of antimicrobial resistance by the food chain is a cross-sectorial problem; the first, antibiotics are extensively used in veterinary, aquaculture and agriculture, the second, antibacterial-resistant bacteria and genes can simply spread at each step of the food chain, and the last can be related to infectious diseases in humans.¹⁸ On the other hand, antibacterial-resistance can have globally dissemination by food chain due to extension of population, international travels and trade in food products.¹² In preparing food animals, vegetables and fishes, in different ecosystems with numerous bacteria, large types of antibiotics are used which can cause to appearance resistant bacteria.19

Todays, antibiotic-resistance, especially that which is transferred from food chain to human is a global concern, and a lot of researches have been conducted to find approaches for solving this critical problem. In the present study, we tried to express some approaches for preventing the appearance of drug-resistant bacteria.

Drug resistant bacteria & food safety

Food safety is a scientific course which has been focused on prevention and controlling the food borne diseases in all processes of the food production process including transport, storage, handling, preparation and in ensuring health and safety of foods for human consumption.²⁰

Resistant food-borne diseases are one of the most important public health problems associated with the risk of emergence of antibacterial resistance in the food production chain. Literatures have indicated that increase in antibiotic resistance bacteria has been caused to an augment in food borne diseases.²¹ Besides, it should be noted that two-third of severity illnesses were related to Gram negative resistant bacteria which the importance and treatment ways of different resistant bacteria are dissimilar.²² Todays, different types of resistant bacteria have been identified in food products and humans, however, some basic and simple food safety measures such as appropriate hand-washing, convenient vegetable-washing, effective cooking temperatures and food storage situation can efficiently reduce and control the spread of antibacterial resistance foodborne pathogens.²⁰ According to Mensah et al,²³ antibiotic residues in food products can have different adverse effects for public health including allergic reactions, hepatotoxicity, mutagenicity, carcinogenicity, toxic effects, nephropathy and antibacterial resistance.

In overall, the results of spreading antibiotic-resistant bacteria and infectious diseases could be summarized as: (1) delay or unsuccessfulness in treatment, (2) limiting in selection of antimicrobials, (3) surviving of resistant strains in treatment of other bacterial illnesses, (4) coexistence and increased pathogenicity of resistance genes in result of selection.²⁴⁻²⁶ According to literatures, there are various food borne pathogens which indicates resistance to different drugs and antibiotics. Some important resistant food borne pathogens including:

Thermotolerant *Campylobacter*: its related disease has short duration with low mortality rate and public health problem. Studies found that some *Campylobacter* spp. were resistant to macrolides, quinolones, chloramphenicol, ampicillin, tetracycline, lincosamides, aminoglycosides and other tylosin, β -lactams and cotrimoxazole.^{27,28}

Salmonella: one of the food borne pathogens which is very high risk factor for human health with remarkable worldwide distribution. *Salmonella* spp. have indicated multidrug resistance toward tetracyclines, kanamycin, sulfonamides, chloramphenicol, streptomycin, cephalosporins and penicillins.²⁹

Staphylococcus aureus spp. are common pathogens for animals and human which reported as resistant-pathogens to penicillins as early as 1948.³⁰ These resistant pathogens are important in dairy product.

Enterococci spp. are common bacteria in the intestinal tract of birds and mammals and known as indicators for enteric contamination of foods. These pathogens can endure unfavorable conditions such as low or high pH, temperature and saline waters^{31,32} which reveals that resistant-*Enterococci* can be an important factor of community-acquired infections.³³ Furthermore, resistant *Enterococci* can transfer resistance gene to human-adapted strains and have adverse effect, indirectly.³²

Yersinia: This genus is composed of various species, including *Y. pestis*, *Y. enterocolitica* and *Y. pseudotuberculosis*, which are pathogenic strains.³⁴ *Y. enterocolitica* may lead to septicemia, septic arthritis, pneumonia, cellulitis, meningitis, empyema, osteomyelitis and panophthalmitis. According to literatures, strains

of biovar 1B (serovar O:8) indicated resistance to carbenicillin, cephalothin and ticarcillin,³⁵ whereas, biovar 3 (0:1,2,3 and O:5,27) strains were resistant to cefoxitin and amoxicillin/clavulanic acid.³⁶

Recent publications where the food borne disease, resistant pathogen and their resistance mechanisms have been reported are shown in Table 1.

Preventive approaches

Farming practices

Biosecurity measures in agriculture and food products have significant role in reduction of antimicrobial resistant bacteria and its transmission from farm to fork. According to the FAO and the WHO, 2011, biosecurity is defined as the measures for reduce or eliminate the threat of the emergence or spreading of diseases at region or country-levels.^{56,57} These measures which called as expressions such as hazard analysis and critical control points, good agricultural practices, good veterinary practices, good hygiene practices focus on health and management/assessment of microbiological risk.^{56,57} Therefore, biosecurity can play economical role in public health control plans, especially in agricultural production, because good and correct practice, wellcontrolled farming and using low chemical materials , leads to healthy animals and reduction in requirement of antibiotic treatments. On the other hand in the situation of appearance of resistant bacteria, effective practices can prevent its dissemination and ensure the food safety and food health.⁵⁸⁻⁶⁰ According to Österberg et al,⁶¹ organic plants indicated a significantly lower amount of resistant bacteria of *E. coli* rather than conventional ones. Figure 1 indicates the ways of appearance of drug- resistant bacteria in society.

Beside, rapid diagnostics of resistant bacteria and their genes will aid to agricultural practitioners to early detect and separate the infectious plants and animals and prevent of its quick spread.¹ Almost, few of rapid tools such as PCR and microarray can identify the pattern of resistance and aid to define appropriate treatment methods.⁶² In applying these diagnostic tools, it should be considered that these methods must be rapid, fundamental, repeatable and easy

Table 1. A few food borne pathogens, related disease, antimicrobials and resistance mechanism

| Pathogen | Related disease symptoms | Antimicrobial group | Resistance mechanism | |
|---|--|--|---|----|
| Mycobacterium tuberculosis | Tuberculosis | Fluoroquinolones | Modifying enzymes, target mimicry | 37 |
| Streptococcus pneumonia | Pneumococcal meningitis | Penicillin | Genetic alteration of penicillin-binding protein | |
| Vibrio cholerae | Severe watery diarrhea | Sulfonamides Chromosomal alterations in encoding dihydropter synthase | | 39 |
| | | Tetracycline | Preventing binding of the antibiotic | 40 |
| Shigella dysenteriae | Severe diarrhea | Chloramphenicol, tetracycline | Decreased permeability, efflux | |
| Salmonella thyphi | Typhoid | Chloramphenicol | Alteration in target site, production of chloramphenicol acetyltransferase, active efflux | |
| Campylobacter jejuni | Gastrointestinal illnesses | Tetracycline | Target protection, Change in ribosomal conformation and preventing binding of the antibiotic | |
| Candida krusei, C. albicans, C. glabrata | Yeast infections, oral Thrush | Azoles | Alteration in ergosterol sites, incorporation of different sterols in plasma membrane, reduction in membrane permeability, effect on efflux pumps | |
| Streptococcus spp | Sore throat, scarlet fever | Tetracycline | Target protection, Change in ribosomal conformation and preventing binding of the antibiotic | |
| Enterobacter, Serratia, Pseudomonas, Citrobacter | - | B-lactam antibiotics | Decrease permeability cell membrane | 45 |
| | | Quinolone | Alteration in DNA gyrase, antibiotic efflux systems | 46 |
| | | Aminoglycoside | Cell-wall impermeability, enzymatic modification. | 47 |
| P. aeruginosa | Blood infection, necrotizing enterocolitis | Carbapenem | Mutational loss of porin channel, acquired zinc b-lactamase | |
| Enterococci | Soft tissue infections | Vancomycin | Bypass of antibiotic target | |
| Staphylococcus aureus | Vomiting, diarrhea, dehydration | Methicillin, | Related to mecA gene, decrease affinity of all b-lactams | 50 |
| | | Streptogramin antibiotics, macrolide, lincosamide | Inactivating enzymes, modification of target sites, active efflux | 51 |
| | | Quinolone | Active efflux, changing in DNA topoisomerases | 52 |
| Neisseria Meningitidis | Pyogenic, meningitis and septicemia | Chloramphenicol | Production of chloramphenicol acetyltransferase | 53 |
| | | Rifampin | Alteration in RNA polymerase, membrane permeability | 54 |
| | | Sulfonamides | Chromosomal alterations in encoding dihydropteroate synthase | 55 |
| | | Penicillin | Alterations in penicillin-binding proteins | 53 |



Figure 1. The cycle of emergence and remaining of resistant bacteria (adopted by https://apua.org/science-of-resistance).

to use.12

Todays, a lot of innovative and modern methods have been suggested for rapid and reliable detection of resistance bacteria in food process chain including, Fourier Transform Infrared Spectroscopy, ATR-FTIR,⁶³ Nano scale materials (such as gold nano-wire), magnetic nanoparticles which are based on bacterial metabolic activity and also antibiotic susceptibility in blood or milk.⁶⁴

Alternatives for antibiotics for food animals *Natural antibiotics*

As previous expressed, todays injudicious use of drugs and especially antibiotics is the most important reason in the appearance of drug resistance thus a lot of studies have investigated using the natural and effective antimicrobial agents as alternative and complementary therapy method.⁶⁵ Essential oils are biological and active substances which are produced by plants and can have antibacterial, antifungal, sedative, antioxidant, digestive, anticancer, anti-inflammatory and antiviral activities.⁶⁶⁻⁶⁸ The efficiency of these substances depends on their genotypes, chemical composition, agronomic and environmental conditions.⁶⁹

Recent studies have revealed that essential oils can be used as preservative in foods and even can have prevention role in developing multidrug-resistant bacteria.65,66,70 According to literatures, some essential oils can have synergistic inhibitory effect in combination with conventional drugs which can lead to reducing effective dose of drugs and therefore, lessen their adverse effect.⁷¹ For example, study of Fadli et al,⁷² indicated that combination of conventional antibiotic and essential oil of Moroccan endemic thyme could have synergistic effect in antimicrobial activity and resulting in reduction of required effective dose, toxic side effects and also cost concerning of drug resistant bacteria. On the other hand, combination of essential oils with standard antibiotics can lead to different inhibition mechanisms in resistant bacteria and this can be a new choice to suppress the microbial resistance.73 Duarte et al,71 has studied on antimicrobial effect of classical antibiotics and coriander essential oil against A. baumannii and observed that the essential oil can improve antimicrobial effect of ciprofloxacin, gentamicin and tetracycline.

Essential oils are composed of many different terpenic compounds which lead to have various health effects. These biological substances have been used in different dosage forms including capsules, creams, aerosols, ointments, syrups, sprays and suppositories.⁶⁹

However, essential oils have more advantages as antibiotic alternative, but their utility is limited by very low solubility in water and their sensitivity to oxygen, heat and moisture. Although many researches try to solve these problems. For example, nano-sized formulation or nano-encapsulation of these substances can be viable solutions against these problems.^{74,75}

Table 2 reports recent publications where the effect of using classical antibiotics with essential oils are shown.

Nano-antibiotics

Classical antibiotic therapy lead to antibiotic resistance and this can prevent by using nano-antibiotics.⁸⁵ "Nanoantibiotics" defined as all nano sized materials which have additional antimicrobial activity and can augment the overall efficacy and safety in consumption.⁶⁴ Recent researches have been observed that using nano encapsulation system can improve efficacy of antibiotics.^{86,87}

On the other hand, nano-carriers indicated functional role which can improve drug absorption by enhancing solubility, preventing from drug degradation, controlling intracellular penetration and etc.⁸⁸ Today, biological nanocarrier systems such as liposomes and chitosan based capsules have been used as drug carrier because of their advantages (including biodegradability, economically, biocompatibility, having no or minimal side effects).⁸⁹ The other advantage of nano-antibiotics is their functional properties by several approaches such as (*i*) producing different antibiotics by the same nanoparticles, (*ii*) using different mechanisms to prevent bacteria growth, (*iii*)

Probiotics and their effects on drug resistance

Table 2. Fractional inhibitory concentration (FIC) indices of classical and natural antimicrobial pairs against antibiotic resistant bacteria

| Combination | FIC | FICI | Target bacteria | Effect | Ref |
|---|-------|-------------|------------------------------------|------------------------------------|-----|
| Meropenem–peppermint - 0.26 E. coli J53 pMG321 | | | | | |
| Meropenem (µg/mL) | 0.13 | | | Synergistic | 76 |
| Peppermint (%, v/v) | 0.13 | | | | |
| Cefoperazone-coriander - 0.750 Acinetobacter baumannii 1025 | | | | | |
| Cefoperazone (µg/mL) | 0.500 | | | No interaction | 71 |
| Coriander oil(%, v/v) | 0.250 | | | | |
| Erythromycin-eugenol | - | < 0.50 | Proteus vulgaris NCIM 2813 | Synergistic | 77 |
| Ampicillin-eugenol | - | 1 | Staphylococcus aureus blaZ | No interaction | 78 |
| Gentamicin-tea tree | - | 0.5 | Acinetobacter baumannii ATCC 19606 | Synergistic | 79 |
| Meropenem-cinnamon | - | 1.5 | Acinetobacter baumannii | No interaction | 80 |
| Chloramphenicol-coriander | - | 0.312 | Acinetobacter baumannii 1025 | | |
| Chloramphenicol (μg/mL) | 0.062 | | | Synergistic | 71 |
| Coriander(%, v/v) | 0.250 | | | | |
| Tetracycline- Lemon thyme | - | 0.95-1.08 | Pseudomonas aeruginosa ATCC 27853 | No interaction | 81 |
| Meropenem–tea tree | - | 1 | E. coli J53 pMG321 | | |
| Meropenem (μg/mL) | 0.50 | | · | No interaction | 76 |
| Tea tree (%, v/v) | 0.50 | | | | |
| Tetracycline-cinnamaldehyde | - | 0.37 | E. coli N00666 | Synergistic | 78 |
| Chloramphenicol-coriander | - | 0.047 | Acinetobacter baumannii 1041 | | |
| loramphenicol (μg/ml) 0.016 | | | Synergistic | 71 | |
| Coriander(%, v/v) | 0.031 | | | _ , 0 | |
| Gentamicin-rosewood | - | 0.11 | Acinetobacter baumannii ATCC 19606 | Synergistic | 79 |
| Piperacillin- Coriander | - | 0.625 | Acinetobacter baumannii 1041 | , 0 | |
| Piperacillin (µg/mL) | 0.125 | | | No interaction | 71 |
| Coriander(%, v/v) | 0.500 | | | _ | |
| Penicillin-carvacrol | _ | 0.32 | Salmonella Typhimurium SGI 1 | Synergistic | 78 |
| Ampicillin-eugenol | _ | < 0.50 | Enterobacter aerogenes NCIM 5139 | Synergistic | 77 |
| Penicillin-Allyl isothiocyanate | - | 0.66 | E. coli SGI1 | No interaction | 78 |
| Meropenem-lemon | _ | 2 | Acinetobacter baumannii | No interaction | 80 |
| Tetracycline-coriander | | 0.185 | Acinetobacter baumannii 1041 | | |
| Tetracycline (µg/mL) | 0.125 | _ | | Synergistic | 71 |
| Coriander oil (%, v/v) | 0.062 | _ | | , | |
| Cefixime-thyme | - | 1.25 | Escherichia coli ATCC 25922 | No interaction | 82 |
| Cefixime-thyme | - | 1 | Klebsiella pneumoniae | No interaction | 72 |
| Ampicillin-thymol | - | 0.12 | Escherichia coli N00 666 | Synergistic | 78 |
| Amoxicillin-sandarac | | 1 | Escherichia coli ATCC 10536 | No interaction | 83 |
| Chloramphenicol-geraniol | | 0.32–0.87 | Klebsiella pneumonia ATCC 700603 | Synergistic | 84 |
| Ceftazidime–cinnamon bark - 2 <i>E. coli</i> J53 pMG309. | | Syncigistic | 76 | | |
| eftazidime–cinnamon bark - 2 E. Coll 55 pMG309. eftazidime (µg/mL) 1 - | | | | No interaction | |
| Cenazianne (µg/me) | 1 | - | | - no meraction | |

FIC of oil = MIC of oil in mixing by antibiotic/MIC of oil alone.

FIC of antibiotic = MIC of antibiotic in mixing by oil/MIC of antibiotic alone.

FIC index = FIC of oil + FIC of antibiotic.

improving in drug efflux, (iv) releasing high amount of antibiotics at the infection site.^{64,90} Therefore, manufacture and development of nano-antibiotics, in addition of prevention of emerging resistant bacteria and reduction the side effects of drugs, can facilitate their storage for long time which can be more economic, although, this science needs investment for developing and becoming commercial technology.

Synthetic biology

Synthetic biology is defined as new patch in using of

genome synthesis technologies to generate novel living systems with functional application.⁹¹ This new approach is offered for the improvement and exploitation of natural products to suppress the emergence of antibiotic resistant pathogens. Novel products are small molecules with genetical codes that have chemical structures which can be used as new antimicrobials.⁹¹ This state-of-art technology can design more cost-effective antibiotics with novel and specific activity, which connect biological systems to engineering processes. With emergence of resistance mechanisms, the antibiotics which are created based on random mutations, facing to obsolesce and till now only a few drugs are new and producing by pharmaceutical sector. Synthetic biology is based on building and integrating gene molecules for reaching to desired results. These biological and functional units can be used for biomedical applications.^{92,93}

Biological systems can generate new drugs relied on the intelligent approaches, such systems are composed by products of biosynthetic genes for new production pathways. This technology can define biological machineries for different bacteria to predict their ability in production of antibiotics and designing synthetic bacteria and also new antibiotics with beneficial functionality by engineered control systems.⁹⁴

Furthermore, this engineering framework, has potential uses in countering bacterial infections, biofuel production, synthesis of natural products and industrial chemicals substitutes.92,95,96 According to reports of genome sequencing of various fungi and bacteria, different species of microorganisms can be used for production of antibiotic.⁹¹ Although, as any developing technology, synthetic biology has some problems which leads to limit its application in industry. Fortunately, in antibiotics discovery, these limitations are far less than other fields.⁹¹ In the application of this technology, it should be considered to microbe safety in production of new antibiotics, security of drug discovery in fermentation tanks and their isolation and to economic concerns of synthetic biology in biosynthetic production of antibiotics.

In biosynthetic gene clusters some antimicrobial compounds including nonribosomal peptides, alkaloids, polyketide antibiotics, bacteriocins, saccharides and terpenoids⁹⁷ which are used for encoding, can control the pathogenic microorganisms.⁹⁸ Todays, by advancing in synthetic biology, DNA sequencing and synthesis, it is possible to access more diversity of antimicrobials and study on these organisms without laboratory culture.⁹⁹

Chu et al., by using nonribosomal peptide synthetases and advanced synthetic biology, could produce new antimicrobials against methicillin-resistant *S. aureus*, which called humimycins, and enhanced the effect of β -lactam antibiotics.¹⁰⁰ In overall, synthetic biology have an important role in steps of design, production and modification of natural products.¹⁰¹ Improvement in gene expression and developing strong regulators and promoters can be useful to produce larger and advanced compounds in future.

Lactic acid bacteria

Lactic acid bacteria can be the other alternative approach for traditional antibiotics. According to literatures the foods containing probiotics, prebiotics, synbiotics have significant role in human and animal health.⁶² Probiotics or in particular lactic acid bacteria, are a group of Grampositive, non-spore forming and acid tolerant organisms which are used as reformer of texture and taste in fermented

foods.^{102,103} These bacteria are characterized by reduction in redox potential and pH, producing lactic acids, diacyls, bacteriocins, hydrogen peroxide, etc., which can degrade mycotoxins, prevent the growth and activity of pathogenic and food spoilage bacteria.^{104,105} Furthermore, according to recent studies, kimchi (traditional fermented food) caused by Lactobacillus plantarum LBP-K10, indicated high antifungal activity and also antiviral activity.^{106,107} The other solutions for preventing or treatment of diseases is association with gut bacterial microecosystem which can improve by prebiotics, probiotics, and synbiotics foods.^{108,109} Probiotics can protect the gut microbial flora, improve immune system and prevent colonization of pathogens.¹¹⁰ Therefore, finding new technologies for identifying and characterizing new strains of these bacteria with more antimicrobial activities can have effective role in protection of public health.

Bacteriocin

As previous, LAB and their bacteriocins have indicated antimicrobial activities against various pathogens and can be used as effective replacement for antibiotics and other chemicals in the food technology. Bacteriocins are a group of potent antimicrobial peptides which compete with related and mostly Gram-positive organisms to gain more nutrients.¹¹¹ These primary metabolites are small and cationic molecules with 30–60 amino acids that are heat stable at 100°C for 10 minutes and are different in mode of action, genetic origin, molecular weight (MW) and biochemical properties.¹¹²

In concept of anti-bacterial resistant bacteria, some novel properties of LAB and bacteriocins including sitespecific delivery of drugs and strategies of anti-quorum sensing can have fascinating roles which lead to increase their application in future. Nisin and pediocin, two bacteriocins isolated from fermented foods, have been approved as natural preservatives by the FDA.¹¹² Some of these antibacterial substances can be effective on both Gram-negative and Gram-positive food borne bacterial pathogens which can be used as biopreservatives and be important in preying human pathogens. Action site of bacteriocins is the cytoplasmic membrane of bacteria.¹⁰⁴ Their drawback is associated to the inhibition effect of probiotics against mainly more related organisms and even desirable bacteria as starter cultures. Furthermore, they often have no activity on gram-negative food spoilers and pathogens but according to Kwak et al,106,113 using chelating agents can increase susceptibility of gramnegative bacteria to bacteriocins. According to recent study, heterofermentative Lactobacillus spp. can reduce the harmful microorganisms in Dutch-type cheese production, control human pathogens and therefore can affect on safety, quality and shelf life of food products.¹¹⁴

In spite of the excellent properties of these metabolites, it has been shown that resistant gene could have been horizontal transferred to benefit bacteria (that produce bacteriocin) in uncontrolled in vitro and in vivo analyses.¹¹² Therefore, more researches need to ensure the efficacy and safety of these bacteria and their bacteriocins for health claims and clinical application.

Cyclopeptide

Antimicrobial peptides are a group of metabolites of Lactobacillus spp. which are considered as an approach for novel pharmaceutical applications. These bioactive agents are some small substances including cyclic dipeptides and 2,5-diketopiperazines.¹¹⁵ Borthwick suggested that their inhibitory effect can be related to the presence of double bonds in amino acid residues at the α , β -positions of cyclic dipeptides, and also the NH proton in pyrroline and diketopiperazine ring.115 Dipeptidyl cyclic rings have been introduced as signal molecules which could reduce virulence-factor and prevent microbial growth for few decades 116,117 These peptides such as some diketopiperazines (2,3-, 2,5-, and 2,6-diketopiperazines and their derivatives) indicate inhibitory effect against fungi, Gram- negative and - positive bacteria.¹¹⁸ According to Kwak et al,107 cis-cyclo (L-Phe-L-Pro) and cis-cyclo (L-Leu-L-Pro) have inhibitory effect on influenza A virus and also cis-cyclo (L-Phe-L-Pro) and cis-cyclo (L-Val-L-Pro) are active against growth of Ganoderma boninense and Candida albicans in plant and human, respectively.¹⁰⁶

Liu et al¹¹⁹ reported some of these peptides were isolated from kimchi (a Chinese fermented food) and had antifungal and antibacterial effects. Studies showed that antioxidant peptides have some functional roles and suggested a novel microbial diketopiperazine from the cyclo (His-Leu) which produced by Bacillus subtilis.^{120,121} Researchers reported that cyclo (His-Pro) plus high doses of zinc have anti-hyperglycemic effect and stimulating consumption of muscle glucose¹²² and reduce obese diabetic (ob/ob) which can have significant positive effect on human health.¹²³ Furthermore, Cyclo (Phe-Phe) can inhibit the development of dementia and Alzheimer's disease by preventing acetylcholinesterase and serotonin transporter.¹²⁴ According to Lee *et al.*,¹²⁵ Cyclo (Trp-Trp) from some strains of Streptomyces indicated significant inhibitory effect on multi drug resistance of Acinetobacter baumannii and the other fungal and bacterial strains. Lind et al,126 observed antifungal activity against Rhodotorula mucilaginosa and Aspergillus fumigatus by cyclo (Ile-Pro) and Cyclo (Phe-Pro).

Therefore, antimicrobial peptides can be an alternative for traditional antibiotics, also these substances indicate bioactive activities, such as antiviral, antifungal, antitumor, antiprion and glycosidase inhibitor activities.¹²⁷ However, some of these components have shown toxic effects on mammalian cells.¹²⁸

Immunostimulant

Vaccination is an infection preventive strategy which is based on immunological memory and body immune

response to the foreign agent.¹²⁹ Vaccines are inactivated or attenuated pathogens which can have protection role in again exposure to the same pathogen in future.^{130,131} In modern medicine, antibiotics and vaccines are used as the two greatest measures for prevention and treatment. The vaccines can suppress the antibacterial resistant bacteria by reducing the pathogen population which cause to reduce the level of antibacterial use and appearance of antibacterial resistant bacteria.¹³² Vaccines have prevalence role on resistant bacteria by direct and indirect mechanisms. They reduce the use of antibiotics in individuals for the same pathogen and also help to prevent transmission of disease to others. Furthermore, vaccines can affect on non-bacterial pathogens which can associate to superinfects and need to medicine treatments.133 Unfortunately, there are no certain treatment for many infectious diseases for example malaria, HIV, tuberculosis and salmonella which can lead to global problem.¹³⁴ Vaccination treatment of these diseases may need more cost but can be an economical solution in long term and can inhibit the drug resistance which in turn protect millions of lives. The use of vaccination as an immunization program is a preventing method in animals and can significantly increase the productivity. According to studies, using lower antibiotics with appropriated novel vaccines program will be able to lessen the worldwide spread of infective diseases.130,133

Bacteriophage

Bacteriophages are specifically viruses which act against bacteria as an alternative for antibiotics with breaking cell wall for solving appearance of resistance in bacteria.¹³⁵ Bacteriophages with DNA or RNA genomes can encode endolysin enzymes that lysis cell wall by cleavage peptidoglycan. Besides, Bacteriophages genome encode proteins that are called amurins which inhibit cell wall synthesis resulting in breakage cell wall.¹³⁶

Efforts for using bacteriophage for treatment is related to before discovering discovery of antibiotics.137 Bacteriophages have two lysogenic and lytic life cycles which have potential role in treatment options.135 A characteristic of bacteriophages is their specificity that can only act against targeted bacteria without any adverse effect on normal flora which is very important for improvement of health. On the other hand, this specificity lead to some problems in immunity issue of phage therapy and besides need to high specific procedures.¹³⁸ Furthermore, this specificity causes problem for infections which are generally colonized by different strains of bacteria.¹³⁹ Although a few studies confirmed the safety of oral administration of phage,^{140,141} the important issue is correct phage translocation across the intestinal epithelium.142 Of course, studies indicated this translocation can be useful for the body by regulation of immune system to indigenous microbe antigens through the prevention of tumor necrosis factor, interleukin-2 and interferon gamma production.¹⁴² However, some other studies did not observe significant increase in levels of cytokine through phage treatment.¹⁴³ Despite of limited data about phage therapy, literatures have reported that phage therapy in addition to reduction of the gut pathogenic flora, causes very lower perturbation in comparison to traditional antibiotics.^{144,145}

Regional specificity lead to finding phages with the highest infectivity against the target pathogen. This can be more helpful when looking for phages for antibiotic resistant bacteria especially in hospital.¹⁴⁶ Moreover, phages have some enzymes such as extracellular polymeric substances depolymerase which can degrade bacterial biofilms and extracellular polymeric substances but antibiotic cannot treat the infection of biofilm-based bacteria.¹⁴⁷ Gabisoniya et al¹⁴⁸ observed that phages inhibited additional biofilm formation and also destroyed existing biofilm of the *Pseudomonas aeruginosa*.

Predatory bacteria

The last alternative solution against resistant bacteria infections is the use of some bacteria known as "predatory bacteria". Micavibrio aeruginosavorus, Bdellovibrio bacteriovorus and associated organisms are gram-negative bacteria and belong to two subgroup of proteobacteria.^{149,150} These bacteria due to their proteases and DNases, demonstrated potential in predation of various pathogenic bacteria and are not specific against gram-negative bacteria.^{115,134,135} The interesting factor of these predatory bacteria is that they can prey effectively even bacterial biofilms and multidrug-resistant pathogens including E. coli, P. aeruginosa, A. baumannii, Pseudomonas putida and Klebsiella pneumonia.110,151,152 Predatory bacteria could serve both as probiotic and antibiotic organisms.¹⁰ Studies reported that these bacteria are effective in treatment of ocular diseases caused by Shigella flexneri and Moraxella bovis in rabbits and cows, respectively.¹⁵³ With the development of antibacterial resistant bacteria and inadequately treatment by conventional antibiotics, predatory bacteria as live antibacterials can have effective role in human health and treatment of diseases.^{153,154}

In spite of useful properties of these bacteria, they have some limitations in their application. Predatory bacteria can have negative effect on the natural microbiota of the body.¹⁵⁵ On the other hand, they may have incomplete predation on bacteria and remain a few number of bacteria.¹⁵¹ Furthermore, existence of Gram-positive bacteria can reduce their predation efficacy.¹⁵¹

Conclusion

Drug resistant bacteria are a global concern phenomenon which can have adverse effects on public health and economy. The assurance of food safety from farm to fork is significantly affected by inappropriate use of drugs and especially antibiotics as treatment agents and growth promoters in food animals. Judicious use of antibiotics has been led to the emergence of anti-bacterial resistant pathogens which can contaminate the food products, reach human body and cause different problems to health. For solving this issue, many researchers have investigated on alternatives for antibiotics in farm practice and also human treatment. Simple biosecurity measures in agriculture and food production process are really beneficial practices which have significant role in food safety and transmission of drug resistant bacteria to humans. Furthermore, combination therapy, combining essential oils and traditional antibiotics, as a new method in pharmacology can be an effective and economic approach against antibiotic resistance. However, these methods need more consideration, encouragement and investment. The discovery and development of novel drugs with new technologies as alternatives to classical antibiotics is a vital issue in global health.

Ethical Issues

Not applicable.

Conflict of Interest

Authors declare no conflict of interest in this study.

Acknowledgments

This study is related to the project NO. 1397/71592 stimulated to Student Research Committee, Shahid Beheshti University of Medical Sciences, Tehran, Iran. We also appreciate the "Student Research Committee" and "Research & Technology Chancellor" in Shahid Beheshti University of Medical Sciences for their financial support of this study.

References

- 1. O'Neill J. *Tackling a global health crisis: initial steps*. London: Review on Antimicrobial Resistance; 2015.
- 2. World Health Organization (WHO). *World health statistics* 2015. Geneva: WHO; 2015.
- McEwen SA, Fedorka-Cray PJ. Antimicrobial use and resistance in animals. *Clin Infect Dis* 2002;34 Suppl 3:S93-s106. doi: 10.1086/340246
- USDA. Part III: Health management and biosecurity in us feedlots, 1999. Available from: https://www.aphis.usda. gov/animal_health/nahms/feedlot/downloads/feedlot99/ Feedlot99_dr_PartIII.pdf.
- Granowitz EV, Brown RB. Antibiotic adverse reactions and drug interactions. *Crit Care Clin* 2008;24(2):421-42, xi. doi: 10.1016/j.ccc.2007.12.011
- Watts JL, Sweeney MT, Lubbers BV. Antimicrobial susceptibility testing of bacteria of veterinary origin. *Microbiol Spectr* 2018;6(2). doi: 10.1128/microbiolspec. ARBA-0001-2017
- 7. van den Bogaard AE, Stobberingh EE. Epidemiology of resistance to antibiotics. Links between animals and humans. *Int J Antimicrob Agents* 2000;14(4):327-35.
- Cabello FC. Heavy use of prophylactic antibiotics in aquaculture: a growing problem for human and animal health and for the environment. *Environ Microbiol* 2006;8(7):1137-44. doi: 10.1111/j.1462-2920.2006.01054.x
- 9. Heuer OE, Kruse H, Grave K, Collignon P, Karunasagar I, Angulo FJ. Human health consequences of use of

antimicrobial agents in aquaculture. *Clin Infect Dis* 2009;49(8):1248-53. doi: 10.1086/605667

- Economou V, Gousia P. Agriculture and food animals as a source of antimicrobial-resistant bacteria. *Infect Drug Resist* 2015;8:49-61. doi: 10.2147/idr.s55778
- McCullough AR, Parekh S, Rathbone J, Del Mar CB, Hoffmann TC. A systematic review of the public's knowledge and beliefs about antibiotic resistance. J Antimicrob Chemother 2016;71(1):27-33. doi: 10.1093/jac/ dkv310
- Holmes AH, Moore LS, Sundsfjord A, Steinbakk M, Regmi S, Karkey A, et al. Understanding the mechanisms and drivers of antimicrobial resistance. *Lancet* 2016;387(10014):176-87. doi: 10.1016/s0140-6736(15)00473-0
- Chang Q, Wang W, Regev-Yochay G, Lipsitch M, Hanage WP. Antibiotics in agriculture and the risk to human health: how worried should we be? *Evol Appl* 2015;8(3):240-7. doi: 10.1111/eva.12185
- Coetzee J, Corcoran C, Prentice E, Moodley M, Mendelson M, Poirel L, et al. Emergence of plasmid-mediated colistin resistance (MCR-1) among *Escherichia coli* isolated from South African patients. *S Afr Med J* 2016;106(5):35-6. doi: 10.7196/SAMJ.2016.v106i5.10710
- Liu YY, Wang Y, Walsh TR, Yi LX, Zhang R, Spencer J, et al. Emergence of plasmid-mediated colistin resistance mechanism MCR-1 in animals and human beings in China: a microbiological and molecular biological study. *Lancet Infect Dis* 2016;16(2):161-8. doi: 10.1016/s1473-3099(15)00424-7
- D'Costa VM, McGrann KM, Hughes DW, Wright GD. Sampling the antibiotic resistome. *Science* 2006;311(5759):374-7. doi: 10.1126/science.1120800
- Riesenfeld CS, Schloss PD, Handelsman J. Metagenomics: genomic analysis of microbial communities. *Annu Rev Genet* 2004;38:525-52. doi: 10.1146/annurev. genet.38.072902.091216
- da Costa PM, Loureiro L, Matos AJ. Transfer of multidrugresistant bacteria between intermingled ecological niches: the interface between humans, animals and the environment. *Int J Environ Res Public Health* 2013;10(1):278-94. doi: 10.3390/ijerph10010278
- 19. Acar JF, Moulin G. Antimicrobial resistance at farm level. *Rev Sci Tech* 2006;25(2):775-92.
- 20. Founou LL, Founou RC, Essack SY. Antibiotic resistance in the food chain: a developing country-perspective. *Front Microbiol* 2016;7:1881. doi: 10.3389/fmicb.2016.01881
- 21. Verraes C, Van Boxstael S, Van Meervenne E, Van Coillie E, Butaye P, Catry B, et al. Antimicrobial resistance in the food chain: a review. *Int J Environ Res Public Health* 2013;10(7):2643-69. doi: 10.3390/ijerph10072643
- 22. Hamers FF. European Centre for Disease Prevention and Control issues guidance for the introduction of human papillomavirus (HPV) vaccines in European Union countries. *Euro Surveill* 2008;13(4):1854-61. doi: 10.2807/ ese.13.04.08022-en
- Mensah SE, Koudande OD, Sanders P, Laurentie M, Mensah GA, Abiola FA. Antimicrobial residues in foods of animal origin in Africa: public health risks. *Rev Sci Tech* 2014;33(3):987-96, 75-86.
- 24. Fluit AC. Towards more virulent and antibiotic-resistant Salmonella? *FEMS Immunol Med Microbiol* 2005;43(1):1-11. doi: 10.1016/j.femsim.2004.10.007

- 25. Gooderham WJ, Hancock RE. Regulation of virulence and antibiotic resistance by two-component regulatory systems in *Pseudomonas aeruginosa*. *FEMS Microbiol Rev* 2009;33(2):279-94. doi: 10.1111/j.1574-6976.2008.00135.x
- 26. Guerra B, Junker E, Miko A, Helmuth R, Mendoza MC. Characterization and localization of drug resistance determinants in multidrug-resistant, integroncarrying Salmonella enterica serotype Typhimurium strains. Microb Drug Resist 2004;10(2):83-91. doi: 10.1089/1076629041310136
- Alfredson DA, Korolik V. Antibiotic resistance and resistance mechanisms in *Campylobacter jejuni* and *Campylobacter coli*. FEMS Microbiol Lett 2007;277(2):123-32. doi: 10.1111/j.1574-6968.2007.00935.x
- Koluman A, Dikici A. Antimicrobial resistance of emerging foodborne pathogens: status quo and global trends. *Crit Rev Microbiol* 2013;39(1):57-69. doi: 10.3109/1040841x.2012.691458
- 29. Olsen SJ, Ying M, Davis MF, Deasy M, Holland B, Iampietro L, et al. Multidrug-resistant Salmonella Typhimurium infection from milk contaminated after pasteurization. *Emerg Infect Dis* 2004;10(5):932-5. doi: 10.3201/eid1005.030484
- 30. Huttner A, Harbarth S, Carlet J, Cosgrove S, Goossens H, Holmes A, et al. Antimicrobial resistance: a global view from the 2013 World Healthcare-Associated Infections Forum. *Antimicrob Resist Infect Control* 2013;2:31. doi: 10.1186/2047-2994-2-31
- Hummel A, Holzapfel WH, Franz CM. Characterisation and transfer of antibiotic resistance genes from enterococci isolated from food. *Syst Appl Microbiol* 2007;30(1):1-7. doi: 10.1016/j.syapm.2006.02.004
- Werner G, Coque TM, Franz CM, Grohmann E, Hegstad K, Jensen L, et al. Antibiotic resistant enterococci-tales of a drug resistance gene trafficker. *Int J Med Microbiol* 2013;303(6-7):360-79. doi: 10.1016/j.ijmm.2013.03.001
- Staley C, Dunny GM, Sadowsky MJ. Environmental and animal-associated enterococci. Adv Appl Microbiol 2014;87:147-86. doi: 10.1016/b978-0-12-800261-2.00004-9
- Carniel E. Plasmids and pathogenicity islands of yersinia. In: Hacker J, Kaper JB, eds. *Pathogenicity islands and the evolution of pathogenic microbes*. Berlin, Heidelberg: Springer; 2002. p. 89-108. doi: 10.1007/978-3-662-09217-0_6
- Preston MA, Brown S, Borczyk AA, Riley G, Krishnan C. Antimicrobial susceptibility of pathogenic *Yersinia* enterocolitica isolated in Canada from 1972 to 1990. Antimicrob Agents Chemother 1994;38(9):2121-4. doi: 10.1128/aac.38.9.2121
- Pham JN, Bell SM, Lanzarone JY. Biotype and antibiotic sensitivity of 100 clinical isolates of *Yersinia enterocolitica*. *J Antimicrob Chemother* 1991;28(1):13-8. doi: 10.1093/ jac/28.1.13
- Fong DH, Berghuis AM. Substrate promiscuity of an aminoglycoside antibiotic resistance enzyme via target mimicry. *EMBO J* 2002;21(10):2323-31. doi: 10.1093/ emboj/21.10.2323
- Philippe J, Gallet B, Morlot C, Denapaite D, Hakenbeck R, Chen Y, et al. Mechanism of beta-lactam action in *Streptococcus pneumoniae*: the piperacillin paradox. *Antimicrob Agents Chemother* 2015;59(1):609-21. doi: 10.1128/aac.04283-14

- Kitaoka M, Miyata ST, Unterweger D, Pukatzki S. Antibiotic resistance mechanisms of Vibrio cholerae. J Med Microbiol 2011;60(Pt 4):397-407. doi: 10.1099/jmm.0.023051-0
- 40. Kar SK, Pal BB, Khuntia HK, Achary KG, Khuntia CP. Emergence and spread of tetracycline resistant Vibrio cholerae O1 El Tor variant during 2010 cholera epidemic in the tribal areas of Odisha, India. *Int J Infect Dis* 2015;33:45-9. doi: 10.1016/j.ijid.2014.12.025
- Puzari M, Sharma M, Chetia P. Emergence of antibiotic resistant Shigella species: A matter of concern. *J Infect Public Health* 2018;11(4):451-4. doi: 10.1016/j.jiph.2017.09.025
- 42. Ugboko H, De N. Mechanisms of Antibiotic resistance in Salmonella typhi. *Int J Curr Microbiol App Sci* 2014;3(12):461-76.
- Donhofer A, Franckenberg S, Wickles S, Berninghausen O, Beckmann R, Wilson DN. Structural basis for TetMmediated tetracycline resistance. *Proc Natl Acad Sci U S A* 2012;109(42):16900-5. doi: 10.1073/pnas.1208037109
- 44. Whaley SG, Berkow EL, Rybak JM, Nishimoto AT, Barker KS, Rogers PD. Azole antifungal resistance in Candida albicans and emerging non-albicans *Candida* species. *Front Microbiol* 2016;7:2173. doi: 10.3389/fmicb.2016.02173
- Ruppe E, Woerther PL, Barbier F. Mechanisms of antimicrobial resistance in Gram-negative bacilli. *Ann Intensive Care* 2015;5(1):61. doi: 10.1186/s13613-015-0061-0
- Hooper DC, Jacoby GA. Mechanisms of drug resistance: quinolone resistance. *Ann N Y Acad Sci* 2015;1354:12-31. doi: 10.1111/nyas.12830
- 47. Gad GF, Mohamed HA, Ashour HM. Aminoglycoside resistance rates, phenotypes, and mechanisms of Gramnegative bacteria from infected patients in upper Egypt. *PLoS One* 2011;6(2):e17224. doi: 10.1371/journal. pone.0017224
- Fuste E, Lopez-Jimenez L, Segura C, Gainza E, Vinuesa T, Vinas M. Carbapenem-resistance mechanisms of multidrugresistant *Pseudomonas aeruginosa. J Med Microbiol* 2013;62(Pt 9):1317-25. doi: 10.1099/jmm.0.058354-0
- Miller WR, Munita JM, Arias CA. Mechanisms of antibiotic resistance in enterococci. *Expert Rev Anti Infect Ther* 2014;12(10):1221-36. doi: 10.1586/14787210.2014.956092
- Peacock SJ, Paterson GK. Mechanisms of methicillin resistance in *Staphylococcus aureus*. *Annu Rev Biochem* 2015;84:577-601. doi: 10.1146/annurevbiochem-060614-034516
- 51. Otsuka T, Zaraket H, Takano T, Saito K, Dohmae S, Higuchi W, et al. Macrolide-lincosamide-streptogramin B resistance phenotypes and genotypes among Staphylococcus aureus clinical isolates in Japan. *Clin Microbiol Infect* 2007;13(3):325-7. doi: 10.1111/j.1469-0691.2006.01632.x
- Redgrave LS, Sutton SB, Webber MA, Piddock LJ. Fluoroquinolone resistance: mechanisms, impact on bacteria, and role in evolutionary success. *Trends Microbiol* 2014;22(8):438-45. doi: 10.1016/j.tim.2014.04.007
- Hedberg ST, Fredlund H, Nicolas P, Caugant DA, Olcen P, Unemo M. Antibiotic susceptibility and characteristics of *Neisseria meningitidis* isolates from the African meningitis belt, 2000 to 2006: phenotypic and genotypic perspectives. *Antimicrob Agents Chemother* 2009;53(4):1561-6. doi: 10.1128/aac.00994-08
- 54. Nolte O, Muller M, Reitz S, Ledig S, Ehrhard I, Sonntag HG. Description of new mutations in the rpoB gene in

rifampicin-resistant *Neisseria meningitidis* selected in vitro in a stepwise manner. *J Med Microbiol* 2003;52(Pt 12):1077-81. doi: 10.1099/jmm.0.05371-0

- 55. Fermer C, Kristiansen BE, Skold O, Swedberg G. Sulfonamide resistance in *Neisseria meningitidis* as defined by site-directed mutagenesis could have its origin in other species. *J Bacteriol* 1995;177(16):4669-75. doi: 10.1128/ jb.177.16.4669-4675.1995
- 56. FAO. Guidelines for risk analysis of food borne antimicrobial resistance (cac/gl77-2011). Food and Agriculture Organization of the United Nations (FAO) and World Health Organization (WHO); 2011.
- 57. FAO. Biosecurity in food and agriculture. Rome: Food and Agriculture Organization of the United Nations. 2003.
- Nahar A, Siddiquee M, Nahar S, Anwar KS, Islam S. Multidrug resistant-proteus mirabilis isolated from chicken droppings in commercial poultry farms: Bio-security concern and emerging public health threat in Bangladesh. *J Biosafety Health Educ* 2014;2(2):1-5. doi:10.4172/2332-0893.1000120
- 59. Postma M, Backhans A, Collineau L, Loesken S, Sjolund M, Belloc C, et al. Evaluation of the relationship between the biosecurity status, production parameters, herd characteristics and antimicrobial usage in farrow-to-finish pig production in four EU countries. *Porcine Health Manag* 2016;2:9. doi: 10.1186/s40813-016-0028-z
- 60. Sivapuram PVRK, Sano D, Srivastava N. *Food Safety in the Asia-Pacific Region: Current status, policy perspectives and a way forward.* In: Sustainable Consumption and Production in the Asia-Pacific Region: Effective Responses in a Resource Constrained World, Institute for Global Environmental Strategies, White Paper III. Hayama, Japan: Institute for Global Environmental Strategies; 2010. p. 215-38.
- 61. Österberg J, Wingstrand A, Nygaard Jensen A, Kerouanton A, Cibin V, Barco L, et al. Antibiotic resistance in Escherichia coli from pigs in organic and conventional farming in four European countries. *PLoS One* 2016;11(6):e0157049. doi: 10.1371/journal.pone.0157049
- 62. Woolhouse M, Ward M, van Bunnik B, Farrar J. Antimicrobial resistance in humans, livestock and the wider environment. *Philos Trans R Soc Lond B Biol Sci* 2015;370(1670):20140083. doi: 10.1098/rstb.2014.0083
- 63. Lechowicz L, Urbaniak M, Adamus-Bialek W, Kaca W. The use of infrared spectroscopy and artificial neural networks for detection of uropathogenic *Escherichia coli* strains' susceptibility to cephalothin. *Acta Biochim Pol* 2013;60(4):713-8.
- Huh AJ, Kwon YJ. "Nanoantibiotics": a new paradigm for treating infectious diseases using nanomaterials in the antibiotics resistant era. *J Control Release* 2011;156(2):128-45. doi: 10.1016/j.jconrel.2011.07.002
- Calo JR, Crandall PG, O'Bryan CA, Ricke SC. Essential oils as antimicrobials in food systems–A review. *Food Control* 2015;54:111-9. doi: 10.1016/j.foodcont.2014.12.040
- 66. Aumeeruddy-Elalfi Z, Gurib-Fakim A, Mahomoodally F. Antimicrobial, antibiotic potentiating activity and phytochemical profile of essential oils from exotic and endemic medicinal plants of Mauritius. *Ind Crops Prod* 2015;71:197-204. doi: 10.1016/j.indcrop.2015.03.058
- 67. Sharifi-Rad J, Hoseini-Alfatemi SM, Sharifi-Rad M, Setzer WN. Chemical Composition, Antifungal and Antibacterial

Activities of Essential Oil from Lallemantia Royleana (Benth. in Wall.) Benth. *J Food Saf* 2015;35(1):19-25. doi: 10.1111/jfs.12139

- Pilevar Z, Hosseini H. Chemical composition, antimicrobial and antioxidant activity of *Echinophora platyloba* DC. J *Pharm Nutr Sci* 2013;3(4):270-83.
- 69. Mohamed AA, Ali SI, El-Baz FK, Hegazy AK, Kord MA. Chemical composition of essential oil and in vitro antioxidant and antimicrobial activities of crude extracts of *Commiphora myrrha* resin. *Ind Crops Prod* 2014;57:10-6. doi: 10.1016/j.indcrop.2014.03.017
- 70. Pilevar Z, Hosseini H, Hajimehdipoor H, Shahraz F, Alizadeh L, Khaneghah AM, et al. The anti-Staphylococcus aureus effect of combined *Echinophora platyloba* essential oil and liquid smoke in beef. *Food Technol Biotechnol* 2017;55(1):117-24. doi: 10.17113/ftb.55.01.17.4633
- Duarte A, Ferreira S, Silva F, Domingues FC. Synergistic activity of coriander oil and conventional antibiotics against *Acinetobacter baumannii. Phytomedicine* 2012;19(3-4):236-8. doi: 10.1016/j.phymed.2011.11.010
- 72. Fadli M, Saad A, Sayadi S, Chevalier J, Mezrioui NE, Pages JM, et al. Antibacterial activity of *Thymus maroccanus* and *Thymus broussonetii* essential oils against nosocomial infection bacteria and their synergistic potential with antibiotics. *Phytomedicine* 2012;19(5):464-71. doi: 10.1016/j.phymed.2011.12.003
- 73. Yap PS, Yiap BC, Ping HC, Lim SH. Essential oils, a new horizon in combating bacterial antibiotic resistance. *Open Microbiol J* 2014;8:6-14. doi: 10.2174/1874285801408010006
- 74. Jamil B, Abbasi R, Abbasi S, Imran M, Khan SU, Ihsan A, et al. Encapsulation of cardamom essential oil in chitosan nano-composites: in-vitro efficacy on antibiotic-resistant bacterial pathogens and cytotoxicity studies s. *Front Microbiol* 2016;7:1580. doi: 10.3389/fmicb.2016.01580
- 75. El Asbahani, Miladi K, Ait Addi EH, Bitar A, Casabianca H, El Mousadik A, et al. Antimicrobial activity of nano-encapsulated essential oils: Comparison to non-encapsulated essential oils. *J Colloid Sci Biotechnol* 2015;4(1):39-48. doi: 10.1166/jcsb.2015.1118
- 76. Yap PS, Lim SH, Hu CP, Yiap BC. Combination of essential oils and antibiotics reduce antibiotic resistance in plasmidconferred multidrug resistant bacteria. *Phytomedicine* 2013;20(8-9):710-3. doi: 10.1016/j.phymed.2013.02.013
- Hemaiswarya S, Doble M. Synergistic interaction of eugenol with antibiotics against Gram negative bacteria. *Phytomedicine* 2009;16(11):997-1005. doi: 10.1016/j. phymed.2009.04.006
- Palaniappan K, Holley RA. Use of natural antimicrobials to increase antibiotic susceptibility of drug resistant bacteria. *Int J Food Microbiol* 2010;140(2-3):164-8. doi: 10.1016/j. ijfoodmicro.2010.04.001
- Rosato A, Piarulli M, Corbo F, Muraglia M, Carone A, Vitali ME, et al. In vitro synergistic antibacterial action of certain combinations of gentamicin and essential oils. *Curr Med Chem* 2010;17(28):3289-95.
- Guerra FQ, Mendes JM, Sousa JP, Morais-Braga MF, Santos BH, Melo Coutinho HD, et al. Increasing antibiotic activity against a multidrug-resistant *Acinetobacter* spp by essential oils of *Citrus limon* and *Cinnamomum zeylanicum*. *Nat Prod Res* 2012;26(23):2235-8. doi: 10.1080/14786419.2011.647019
- 81. Miladinovic DL, Ilic BS, Miladinovic LC, Kocic BD, Ciric

VM, Stankov-Jovanovic VP, et al. Antibacterial activity of *Thymus pulegioides* essential oil and its synergistic potential with antibiotics: a chemometric approach. In: Govil JN, Bhattacharya S, eds. *Essential Oils III and Phytopharmacology*. Houston: Studium Press LLC; 2013:101-36.

- Kasrati A, Alaoui Jamali C, Fadli M, Bekkouche K, Hassani L, Wohlmuth H, et al. Antioxidative activity and synergistic effect of *Thymus saturejoides* Coss. essential oils with cefixime against selected food-borne bacteria. *Ind Crops Prod* 2014;61:338-44. doi: 10.1016/j.indcrop.2014.07.024
- 83. Djouahri A, Saka B, Boudarene L, Benseradj F, Aberrane S, Aitmoussa S, et al. In vitro synergistic/antagonistic antibacterial and anti-inflammatory effect of various extracts/essential oil from cones of *Tetraclinis articulata* (Vahl) Masters with antibiotic and anti-inflammatory agents. *Ind Crops Prod* 2014;56:60-6. doi: 10.1016/j. indcrop.2014.02.035
- van Vuuren SF, Suliman S, Viljoen AM. The antimicrobial activity of four commercial essential oils in combination with conventional antimicrobials. *Lett Appl Microbiol* 2009;48(4):440-6. doi: 10.1111/j.1472-765X.2008.02548.x
- 85. Jun SH, Cha SH, Kim JH, Yoon M, Cho S, Park Y. Silver nanoparticles synthesized using *Caesalpinia sappan* extract as potential novel nanoantibiotics against methicillinresistant *Staphylococcus aureus*. *J Nanosci Nanotechnol* 2015;15(8):5543-52.
- Hajipour MJ, Fromm KM, Ashkarran AA, Jimenez de Aberasturi D, de Larramendi IR, Rojo T, et al. Antibacterial properties of nanoparticles. *Trends Biotechnol* 2012;30(10):499-511. doi: 10.1016/j.tibtech.2012.06.004
- Jamil B, Syed MA. Nano-antimicrobials: A Viable Approach to Tackle Multidrug-Resistant Pathogens. In: Rai M., dos Santos CA, eds. *Nanotechnology applied to pharmaceutical technology*. Cham: Springer; 2017:31-54. doi: 10.1007/978-3-319-70299-5_2
- Thorley AJ, Ruenraroengsak P, Potter TE, Tetley TD. Critical determinants of uptake and translocation of nanoparticles by the human pulmonary alveolar epithelium. *ACS Nano* 2014;8(11):11778-89. doi: 10.1021/nn505399e
- Kumari A, Yadav SK, Yadav SC. Biodegradable polymeric nanoparticles based drug delivery systems. *Colloids Surf B Biointerfaces* 2010;75(1):1-18. doi: 10.1016/j. colsurfb.2009.09.001
- Pelgrift RY, Friedman AJ. Nanotechnology as a therapeutic tool to combat microbial resistance. Adv Drug Deliv Rev 2013;65(13-14):1803-15. doi: 10.1016/j.addr.2013.07.011
- Thaker MN, Wright GD. Opportunities for synthetic biology in antibiotics: expanding glycopeptide chemical diversity. ACS Synth Biol 2015;4(3):195-206. doi: 10.1021/ sb300092n
- 92. Schukur L, Geering B, Charpin-El Hamri G, Fussenegger M. Implantable synthetic cytokine converter cells with AND-gate logic treat experimental psoriasis. *Sci Transl Med* 2015;7(318):318ra201. doi: 10.1126/scitranslmed.aac4964
- 93. Waters CM, Bassler BL. Quorum sensing: cellto-cell communication in bacteria. Annu Rev Cell Dev Biol 2005;21:319-46. doi: 10.1146/annurev. cellbio.21.012704.131001
- 94. Krishnamurthy M, Moore RT, Rajamani S, Panchal RG. Bacterial genome engineering and synthetic biology: combating pathogens. *BMC Microbiol* 2016;16(1):258. doi:

10.1186/s12866-016-0876-3

- 95. Keasling JD. Synthetic biology and the development of tools for metabolic engineering. *Metab Eng* 2012;14(3):189-95. doi: 10.1016/j.ymben.2012.01.004
- Ruder WC, Lu T, Collins JJ. Synthetic biology moving into the clinic. *Science* 2011;333(6047):1248-52. doi: 10.1126/ science.1206843
- Cotter PD, Ross RP, Hill C. Bacteriocins a viable alternative to antibiotics? *Nat Rev Microbiol* 2013;11(2):95-105. doi: 10.1038/nrmicro2937
- Medema MH, Kottmann R, Yilmaz P, Cummings M, Biggins JB, Blin K, et al. Minimum Information about a biosynthetic gene cluster. *Nat Chem Biol* 2015;11(9):625-31. doi: 10.1038/nchembio.1890
- Culligan EP, Sleator RD. Antibiotics v2.0: computational and synthetic biology approaches to combat antibiotic resistance. *Future Microbiol* 2017;12:267-9. doi: 10.2217/ fmb-2017-0005
- 100. Chu J, Vila-Farres X, Inoyama D, Ternei M, Cohen LJ, Gordon EA, et al. Discovery of MRSA active antibiotics using primary sequence from the human microbiome. *Nat Chem Biol* 2016;12(12):1004-6. doi: 10.1038/nchembio.2207
- 101. Smanski MJ, Zhou H, Claesen J, Shen B, Fischbach MA, Voigt CA. Synthetic biology to access and expand nature's chemical diversity. *Nat Rev Microbiol* 2016;14(3):135-49. doi: 10.1038/nrmicro.2015.24
- 102. Kaban G, Kaya M. Identification of lactic acid bacteria and gram-positive catalase-positive cocci isolated from naturally fermented sausage (sucuk). *J Food Sci* 2008;73(8):M385-8. doi: 10.1111/j.1750-3841.2008.00906.x
- 103. Hati S, Mandal S, Prajapati J. Novel starters for value added fermented dairy products. *Curr Res Nutr Food Sci* 2013;1(1):83-91, doi: 10.12944/CRNFSJ.1.1.09
- 104. Parada JL, Caron CR, Medeiros ABP, Soccol CR. Bacteriocins from lactic acid bacteria: purification, properties and use as biopreservatives. *Braz Arch Biol Technol* 2007;50(3):512-42. doi: 10.1590/S1516-89132007000300018
- 105. De Vuyst L, Leroy F. Bacteriocins from lactic acid bacteria: production, purification, and food applications. *J Mol Microbiol Biotechnol* 2007;13(4):194-9. doi: 10.1159/000104752
- 106. Kwak MK, Liu R, Kwon JO, Kim MK, Kim AH, Kang SO. Cyclic dipeptides from lactic acid bacteria inhibit proliferation of the influenza A virus. J Microbiol 2013;51(6):836-43. doi: 10.1007/s12275-013-3521-y
- 107. Kwak MK, Liu R, Kim MK, Moon D, Kim AH, Song SH, et al. Cyclic dipeptides from lactic acid bacteria inhibit the proliferation of pathogenic fungi. *J Microbiol* 2014;52(1):64-70. doi: 10.1007/s12275-014-3520-7
- 108. Gaggia F, Mattarelli P, Biavati B. Probiotics and prebiotics in animal feeding for safe food production. *Int J Food Microbiol* 2010;141 Suppl 1:S15-28. doi: 10.1016/j. ijfoodmicro.2010.02.031
- 109. Callaway TR, Edrington TS, Anderson RC, Harvey RB, Genovese KJ, Kennedy CN, et al. Probiotics, prebiotics and competitive exclusion for prophylaxis against bacterial disease. *Anim Health Res Rev* 2008;9(2):217-25. doi: 10.1017/s1466252308001540
- 110. Allen HK, Trachsel J, Looft T, Casey TA. Finding alternatives to antibiotics. Ann N Y Acad Sci 2014;1323:91-100. doi: 10.1111/nyas.12468
- 111. Zacharof MP, Lovitt RW. Bacteriocins produced by lactic

acid bacteria a review article. *APCBEE Procedia* 2012;2:50-6. doi: 10.1016/j.apcbee.2012.06.010

- 112. Mokoena MP. Lactic acid bacteria and their bacteriocins: Classification, biosynthesis and applications against uropathogens: A mini-review. *Molecules* 2017;22(8). doi: 10.3390/molecules22081255
- 113. Pilevar Z, Hosseini H. Effects of starter cultures on the properties of meat products: A review. Annu Res Rev Biol 2017;17(6):1-17. doi: 10.9734/ARRB/2017/36330
- 114. Djadouni F, Kihal M. Antimicrobial activity of lactic acid bacteriaandthespectrumoftheirbiopeptidesagainstspoiling germs in foods. *Braz Arch Biol Technol* 2012;55(3):435-44. doi: 10.1590/S1516-89132012000300015
- 115. Borthwick AD. 2,5-Diketopiperazines: synthesis, reactions, medicinal chemistry, and bioactive natural products. *Chem Rev* 2012;112(7):3641-716. doi: 10.1021/cr200398y
- 116. Campbell J, Lin Q, Geske GD, Blackwell HE. New and unexpected insights into the modulation of LuxR-type quorum sensing by cyclic dipeptides. ACS Chem Biol 2009;4(12):1051-9. doi: 10.1021/cb900165y
- 117. Sauguet L, Moutiez M, Li Y, Belin P, Seguin J, Le Du MH, et al. Cyclodipeptide synthases, a family of class-I aminoacyltRNA synthetase-like enzymes involved in non-ribosomal peptide synthesis. *Nucleic Acids Res* 2011;39(10):4475-89. doi: 10.1093/nar/gkr027
- 118. Borthwick AD, Da Costa NC. 2,5-diketopiperazines in food and beverages: Taste and bioactivity. *Crit Rev Food Sci Nutr* 2017;57(4):718-42. doi: 10.1080/10408398.2014.911142
- 119. Liu R, Kim AH, Kwak MK, Kang SO. Proline-based cyclic dipeptides from Korean fermented vegetable kimchi and from *Leuconostoc mesenteroides* LBP-K06 have activities against multidrug-resistant bacteria. *Front Microbiol* 2017;8:761. doi: 10.3389/fmicb.2017.00761
- 120. Elkahoui S, Abdel rahim H, Tabbene O, Shaaban M, Limam F, Laatsch H. Cyclo-(His,Leu): a new microbial diketopiperazine from a terrestrial *Bacillus subtilis* strain B38. *Nat Prod Res* 2013;27(2):108-16. doi: 10.1080/14786419.2012.660635
- 121. Minelli A, Bellezza I, Grottelli S, Galli F. Focus on cyclo(His-Pro): history and perspectives as antioxidant peptide. *Amino Acids* 2008;35(2):283-9. doi: 10.1007/s00726-007-0629-6
- 122. Uyemura K, Dhanani S, Yamaguchi DT, Song MK. Metabolism and toxicity of high doses of cyclo (his-pro) plus zinc in healthy human subjects. *J Drug Metab Toxicol* 2010;1:105. doi: 10.4172/2157-7609.1000105
- 123. Jung EY, Hong YH, Park C, Suh HJ. Effects of Cyclo-His-Pro-enriched yeast hydrolysate on blood glucose levels and lipid metabolism in obese diabetic ob/ob mice. *Nutr Res Pract* 2016;10(2):154-60. doi: 10.4162/nrp.2016.10.2.154
- 124. Tsuruoka N, Beppu Y, Koda H, Doe N, Watanabe H, Abe K. A DKP cyclo(L-Phe-L-Phe) found in chicken essence is a dual inhibitor of the serotonin transporter and acetylcholinesterase. *PLoS One* 2012;7(11):e50824. doi: 10.1371/journal.pone.0050824
- 125. Lee KH, Kim KW, Rhee KH. Identification of *Streptomyces* sp. KH29, which produces an antibiotic substance processing an inhibitory activity against multidrug-resistant *Acinetobacter baumannii*. *J Microbiol Biotechnol* 2010;20(12):1672-6.
- 126. Lind H, Sjogren J, Gohil S, Kenne L, Schnurer J, Broberg A. Antifungal compounds from cultures of dairy

propionibacteria type strains. *FEMS Microbiol Lett* 2007;271(2):310-5. doi: 10.1111/j.1574-6968.2007.00730.x

- 127. Wang X, Li Y, Zhang X, Lai D, Zhou L. Structural diversity and biological activities of the cyclodipeptides from fungi. *Molecules* 2017;22(12). doi: 10.3390/molecules22122026
- 128. Papagianni M, Anastasiadou S. Pediocins: The bacteriocins of Pediococci. Sources, production, properties and applications. *Microb Cell Fact* 2009;8:3. doi: 10.1186/1475-2859-8-3
- 129. Sanghi DK, Tiwle R. A detail comprehensive review on vaccines. *Int J Res Dev Pharm Life Sci* 2014;3(2):887-95.
- 130. Atkins KE, Flasche S. Vaccination to reduce antimicrobial resistance. *Lancet Glob Health* 2018;6(3):e252. doi: 10.1016/ s2214-109x(18)30043-3
- 131. Ginsburg AS, Klugman KP. Vaccination to reduce antimicrobial resistance. *Lancet Glob Health* 2017;5(12):e1176-e7. doi: 10.1016/s2214-109x(17)30364-9
- 132. Wilder-Smith A, Longini I, Zuber PL, Barnighausen T, Edmunds WJ, Dean N, et al. The public health value of vaccines beyond efficacy: methods, measures and outcomes. *BMC Med* 2017;15(1):138. doi: 10.1186/s12916-017-0911-8
- 133. Lipsitch M, Siber GR. How can vaccines contribute to solving the antimicrobial resistance problem? *MBio* 2016;7(3):e00428-16. doi: 10.1128/mBio.00428-16
- 134. Efficacy and safety of RTS,S/AS01 malaria vaccine with or without a booster dose in infants and children in Africa: final results of a phase 3, individually randomised, controlled trial. *Lancet* 2015;386(9988):31-45. doi: 10.1016/ s0140-6736(15)60721-8
- 135. Bragg R, van der Westhuizen W, Lee JY, Coetsee E, Boucher C. Bacteriophages as potential treatment option for antibiotic resistant bacteria. *Adv Exp Med Biol* 2014;807:97-110. doi: 10.1007/978-81-322-1777-0_7
- 136. Woznica WM, Bigos J, Lobocka MB. [Lysis of bacterial cells in the process of bacteriophage release--canonical and newly discovered mechanisms]. *Postepy Hig Med Dosw* (*Online*) 2015;69:114-26.
- 137. Chanishvili N. Phage therapy--history from Twort and d'Herelle through Soviet experience to current approaches. *Adv Virus Res* 2012;83:3-40. doi: 10.1016/b978-0-12-394438-2.00001-3
- 138. Lin DM, Koskella B, Lin HC. Phage therapy: An alternative to antibiotics in the age of multi-drug resistance. World J Gastrointest Pharmacol Ther 2017;8(3):162-73. doi: 10.4292/wjgpt.v8.i3.162
- 139. Servick K. Beleaguered phage therapy trial presses on. *Science* 2016;352(6293):1506. doi: 10.1126/science.352.6293.1506
- 140. Bruttin A, Brussow H. Human volunteers receiving Escherichia coli phage T4 orally: a safety test of phage therapy. *Antimicrob Agents Chemother* 2005;49(7):2874-8. doi: 10.1128/aac.49.7.2874-2878.2005
- 141. McCallin S, Alam Sarker S, Barretto C, Sultana S, Berger B, Huq S, et al. Safety analysis of a Russian phage cocktail: from metagenomic analysis to oral application in healthy human subjects. *Virology* 2013;443(2):187-96. doi: 10.1016/j. virol.2013.05.022
- 142. Gorski A, Wazna E, Dabrowska BW, Dabrowska K, Switala-Jelen K, Miedzybrodzki R. Bacteriophage translocation. *FEMS Immunol Med Microbiol* 2006;46(3):313-9. doi:

10.1111/j.1574-695X.2006.00044.x

- 143. Hong Y, Thimmapuram J, Zhang J, Collings CK, Bhide K, Schmidt K, et al. The impact of orally administered phages on host immune response and surrounding microbial communities. *Bacteriophage* 2016;6(3):e1211066. doi: 10.1080/21597081.2016.1211066
- 144. Mai V, Ukhanova M, Reinhard MK, Li M, Sulakvelidze A. Bacteriophage administration significantly reduces Shigella colonization and shedding by Shigella-challenged mice without deleterious side effects and distortions in the gut microbiota. *Bacteriophage* 2015;5(4):e1088124. doi: 10.1080/21597081.2015.1088124
- 145. Galtier M, De Sordi L, Maura D, Arachchi H, Volant S, Dillies MA, et al. Bacteriophages to reduce gut carriage of antibiotic resistant uropathogens with low impact on microbiota composition. *Environ Microbiol* 2016;18(7):2237-45. doi: 10.1111/1462-2920.13284
- 146. Latz S, Wahida A, Arif A, Hafner H, Hoss M, Ritter K, et al. Preliminary survey of local bacteriophages with lytic activity against multi-drug resistant bacteria. J Basic Microbiol 2016;56(10):1117-23. doi: 10.1002/jobm.201600108
- 147. Abedon ST. Ecology of anti-biofilm agents I: antibiotics versus bacteriophages. *Pharmaceuticals (Basel)* 2015;8(3):525-58. doi: 10.3390/ph8030525
- 148. Gabisoniya TG, Loladze MZ, Nadiradze MM, Chakhunashvili NK, Alibegashvili MG, Tamarashvili NG, et al. [Effects of bacteriophages on biofilm formation by strains of *Pseudomonas aeruginosa*]. *Prikl Biokhim Mikrobiol* 2016;52(3):312-7.
- 149. Davidov Y, Huchon D, Koval SF, Jurkevitch E. A new alpha-proteobacterial clade of Bdellovibrio-like predators: implications for the mitochondrial endosymbiotic theory. *Environ Microbiol* 2006;8(12):2179-88. doi: 10.1111/j.1462-2920.2006.01101.x
- 150. Davidov Y, Jurkevitch E. Diversity and evolution of Bdellovibrio-and-like organisms (BALOs), reclassification of Bacteriovorax starrii as Peredibacter starrii gen. nov., comb. nov., and description of the Bacteriovorax-Peredibacter clade as Bacteriovoracaceae fam. nov. Int J Syst Evol Microbiol 2004;54(Pt 5):1439-52. doi: 10.1099/ ijs.0.02978-0
- 151. Kadouri DE, To K, Shanks RM, Doi Y. Predatory bacteria: a potential ally against multidrug-resistant Gram-negative pathogens. *PLoS One* 2013;8(5):e63397. doi: 10.1371/ journal.pone.0063397
- 152. Lambert C, Sockett RE. Nucleases in *Bdellovibrio* bacteriovorus contribute towards efficient self-biofilm formation and eradication of preformed prey biofilms. *FEMS Microbiol Lett* 2013;340(2):109-16. doi: 10.1111/1574-6968.12075
- 153. Dwidar M, Monnappa AK, Mitchell RJ. The dual probiotic and antibiotic nature of *Bdellovibrio bacteriovorus*. *BMB Rep* 2012;45(2):71-8. doi: 10.5483/BMBRep.2012.45.2.71
- 154. Sockett RE, Lambert C. Bdellovibrio as therapeutic agents: a predatory renaissance? *Nat Rev Microbiol* 2004;2(8):669-75. doi: 10.1038/nrmicro959
- 155. Schoeffield AJ, Williams HN, Turng B, Fackler WA Jr. A comparison of the survival of intraperiplasmic and attack phase Bdellovibrios with reduced oxygen. *Microb Ecol* 1996;32(1):35-46.