Adv Pharm Bull, 2020, 10(3), 350-358 doi: 10.34172/apb.2020.043 https://apb.tbzmed.ac.ir



CrossMark

Microbial Alpha-Amylase Production: Progress, Challenges and Perspectives

Babak Elyasi Far^{1,2}, Yassin Ahmadi³, Ahmad Yari Khosroushahi^{1,4}, Azita Dilmaghani^{1,2*}

¹Drug Applied Research Center, Tabriz University of Medical Sciences, Tabriz, Iran.

²Department of Pharmaceutical Biotechnology, Faculty of Pharmacy, Tabriz University of Medical Sciences, Tabriz, Iran.

³Student Research Committee, Tabriz University of Medical Sciences, Tabriz, Iran.

⁴Department of Medical Nanotechnology, Faculty of Advanced Medical Science, Tabriz University of Medical Sciences, Tabriz, Iran.

Article info

Article History: Received: 29 Apr. 2019 Revised: 23 Oct. 2019 Accepted: 9 Nov. 2019 epublished: 11 May 2020

Keywords:

- Microbial
- Alpha-amylase
- RMS
- Stability
- Optimization

Introduction

Compared to the chemical methods that need harsh conditions such as high pressure and temperature, using of microorganism is considered in many purposes including heavy metal absorption,¹ gene engeering,² digestion,³ production of novel anti-microbs⁴ and particullary for producing of industrial enzymes.^{5,6} Demand on the high-quality productions leads to development of novel methods to improve industrial products such as protease and amylase that are frequently applied in industry and medical science.

There have been identified three types of amylase including α -amylase, β -amylase, and γ -amylase. Alphaamylase is an industrial enzyme (EC 3.2.1.1.), which cleaves internal alpha 1-4 glycosidic bands of starch and other polysaccharides to produce several products such as glucose and maltose.^{7,8} It belongs to the family of GH13 (most of them), GH57, GH119, and GH126⁹ and is one of the most widely used commercial enzymes.¹⁰

Most of alpha-amylases are secreted extracellularly, however some intracellular alpha-amylase have been reported. Regarding the outstanding application of α -amylase, there is an urgent need to develop the costeffective techniques to produce stable and efficient alpha-

Abstract

Alpha-amylase reputes for starch modification by breaking of 1-4 glycosidic bands and is widely applied in different industrial sectors. Microorganisms express unique alpha-amylases with thermostable and halotolerant characteristics dependent on the microorganism's intrinsic features. Likewise, genetic engineering methods are applied to produce enzymes with higher stability in contrast to wild types. As there are widespread application of α -amylase in industry, optimization methods like RSM are used to improve the production of the enzyme ex vivo. This study aimed to review the latest researches on the production improvement and stability of α -amylase.

amylase.¹¹ Here we aim to present different methods of the amylase production and how it is possible to improve the efficiency of alpha-amylase.

Alpha-amylase production in microbial source

A wide range of organisms, including microorganisms such as aquatic bacteria,¹² fungi, actinomycetes, plants, and animals, can produce alpha-amylase.¹³⁻¹⁵

Regarding the high rate of proliferation and growth, microorganisms are the primary source of alphaamylase producing a high volume of the enzyme. Also, the genetically manipulated microorganisms are forced to produce alpha-amylase with novel characteristics like thermos-stability.^{14,16} Additionally, the microorganisms produce large quantity of enzyme, which can be simply optimized by various methods such as response surface methodology.¹⁷ The most widely used microorganisms for the production of alpha-amylase include bacteria, actinomycetes, and fungi.¹⁵

Several bacteria have been shown are capable of producing a tremendous amount of alpha-amylase for industrial applications, these bacteria include *Bacillus amyloliquefaciens*, *Bacillus licheniformis*, and *Bacillus stearothermophilus*. Some bacteria can produce

^{*}Corresponding Author: Azita Dilmaghani, Email: dilmaghani.a@tbzmed.ac.ir

^{© 2020} The Author (s). This is an Open Access article distributed under the terms of the Creative Commons Attribution (CC BY), which permits unrestricted use, distribution, and reproduction in any medium, as long as the original authors and source are cited. No permission is required from the authors or the publishers.

alpha-amylase in harsh conditions; for instance, some thermophilic bacteria produce alpha-amylase at high temperatures. Most of the starch processing steps, including saccharification, gelatinization, and liquefaction, need high temperature, so the thermostable alpha-amylase is useful to progress the possessing steps in such harsh conditions.¹⁸ The most common sources of thermostable α -amylase are Geobacillus bacterium isolated from Manikaran hot springs. The thermophilic alpha-amylases (BLA) have been shown to have more structural flexibility than mesophilic alpha-amylases (BAA). Although the optimum temperature for this enzyme is 80°C, there is an essential need for amylases resistant to other harsh conditions, especially in the industrial process.¹⁹⁻²¹ The halophilic α -amylase tolerates saline and high-temperature conditions.¹⁴ Additionally, this enzyme is resistant to organic solvents and keep its activity at low-water conditions. One of the halophilic bacterial sources of alpha-amylase is Nesterenkonia sp. strain F, which produces enzymes with catalytic activity even in the organic solvents such as chloroform, benzene, cyclohexane, and toluene.22 Due to acidic residues on the surface of halophilic alpha-amylase, this enzyme is stable at low water conditions.²³⁻²⁵

Other types of bacteria can remain alive in low temperatures, such as marine bacteria, and produce coldactive enzymes. Cold-active alpha-amylase is widely used in industry for saving energy²⁶; for example, in detergent contents, it is not needed to heat water for washing and also cold water help to protect clothes' color. In the backing process, cold-stable amylase efficiently reduces the time of dough fermentation and improves bread quality. *Pseudoalteromonas* sp. M175 has been isolated from Antarctic sea ice, which is a common source of cold-stable alpha-amylase.²⁷ The microorganisms produce cold-active amylases that have a flexible polypeptide chain to make an easier accommodation of substrates at the low-temperature condition. Also, the enzymes contain lipid composition to maintain greater membrane fluidity.²⁸

Also, some types of actinomycetes such as *Nocardiopsis aegyptia* can produce cold-adapted enzymes.¹⁵ Actinomycetes, such as *Streptomyces fragilis* DA7-7 produce thermostable alpha-amylase.²⁹

Other microbial sources of alpha-amylase for commercial purposes include *Aspergillus niger*, *Aspergillus awamori*, and *Aspergillus oryzae*.¹⁴ Due to the extracellular secretion of alpha-amylase that is easily isolatable from the microbial culture medium, fungi can be a good source for α -amylase production in the industry.³⁰ Also, other fungi species have been reported to possess some unique features making them suitable for industrial goals; for example, an alpha-amylase produced by *Aspergillus flavus* NSH9 is thermally stable at 50°C.³¹ Table 1 shows different microbial sources for alpha-amylase production.

Application of alpha-amylase

Alpha-amylase is currently used in a broad array of industrial applications such as the production of ethanol and high fructose corn syrup, food, textile, paper, and detergent industries.³²⁻³⁵ Table 2 shows the most current applications of α -amylase in different industries.

Alpha-amylase production *Starch*

Starch is known as a carbon source and the main substrate of alpha-amylase, which is comprised of two parts, amylose (25–30%) and amylopectin (70–75%).^{36,37} Amylose contains glucose monomers that are linked to each other via α (1-4) glycosidic bands and its molecular weight spans 1 × 10⁵ – 1×10⁶ Da. The other polymer is amylopectin, which is polymerized by α (1-4) glycosidic bands and is branched by α (1-6) glycosidic bands with the

molecular weight about 1×107 - 1×109 Da.38

According to the digestibility features, starch is divided into three main groups, including rapidly digestible starch (RDS), slowly digestible starch, and resistant starch (RS).³⁹ The most current RDS are used in the foods and gelatinized waxy. RDS in the digestive system is rapidly degraded (20 minutes) into glucose units and so elevates the blood glucose rapidly.¹⁰

RS is very resistant against digestive mechanisms because of their low glycemic index and these starches are used mainly by bacteria in the colon to generate shortchain fatty acids, which is essential for human health. RS consists of five different types, including encapsulated starch, resistant granules, retrograded amylose, chemically

Table 1. Microbial sources for alpha amylase production

Source	Microbial type	Feature of alpha-amylase
Bacillus stearothermophilus	Bacteria	Thermophile alpha-amylase
Geobacillus bacterium	Bacteria	Thermophile alpha-amylase
Nesterenkonia sp. strain F	Bacteria	Halophilic enzyme
Bacterium Pseudoalteromonas sp. M175	Bacteria	Cold-active alpha-amylase
Nocardiopsis aegyptia	Actinomycetes	Cold-active alpha-amylase
Streptomyces fragilis	Actinomycetes	Cold-active alpha-amylase
Aspergillus niger	Fungi	Commercial production
Aspergillus awamori	Fungi	Commercial production
Aspergillus oryzae	Fungi	Commercial production
Aspergillus flavus	Fungi	Thermophile alpha-amylase

Table 2. Industrial applications of α -amylase

Industry	Application	Further explanation	Microorganism
Detergent (laundry and dish wash)	Starch stain removal	Detergents containing chemicals don't break down easily in waste-water and cause pollution and eutrophication in the rivers/water bodies so the same may be replaced by enzyme which enhances the detergents ability to remove tough stains and making the detergent environmentally safe. The targeted benefit of enzyme inclusion in detergents is due to much milder conditions; 90% of all liquid detergents contain α -amylase. These enzymes degrade the residues of starchy foods	Bacillus or Aspergillus (Amylases with activity at lower temperatures, alkaline pH, and oxidative conditions)
Starch liquefaction	Dispersion of insoluble starch granules in aqueous solution and decreasing viscosity followed by partial hydrolysis and	This is the most widespread applications of α -amylases (thermostable) which are used for starch hydrolysis into starch hydrolysates such as glucose and fructose. Because of their high sweetening property, these are used in huge quantities in the beverage industry as sweeteners for soft drinks. Liquefaction process causes reduction in viscosity and partial hydrolysis of starch thus avoided during subsequent cooling.	Bacillus species (thermostable) amyloliquefaciens; Bacillus stearothermophilus or Bacillus licheniformis
Fuel alcohol production	Starch liquefaction and saccharification	Ethanol has been used as fuel a since the early days of the automobile. Ethanol is a significant product of 21st century with its versatile usages and widely consumption across the globe. For the ethanol production, starch is the most used substrate due to its low price and easily available raw material. In this production, starch has to be solubilized and then submitted to two enzymatic steps in order to obtain fermentable sugars. The bioconversion of starch into ethanol involves liquefaction and saccharification, where starch is converted into sugar using an amylolytic microorganism or enzymes such as α -amylase, followed by fermentation, where sugar is converted into ethanol	As yeast Saccharomyces cerevisiae
Food industry	Bread softness and volume, flour adjustment; Juice treatment, low calorie beer	Wide applications of α -amylases in food industry include baking, brewing, starch liquefaction as well as a digestive aid (5). They are widely used in baking industry as flavour enhancement and antistaling agent to improve bread quality. α -amylases converts starch to smaller dextrins, which are subsequently fermented by the yeast. improves the taste, crust colour and toasting qualities of the bread; Amylases are also used for the clarification of beer or fruit juices, or for the pretreatment of animal feed to improve the digestibility of fiber	Thermostable maltogenic amylase of <i>Bacillus</i> stearothermophilus
Textile industry	De-sizing	Modern production processes for textiles introduce a considerable strain on the warp during weaving. The yarn must, therefore, be prevented from breaking. For this purpose a removable protective layer is applied to the threads. Sizing agents like starch are applied to yarn before fabric production to ensure a fast and secure weaving process. Starch is later removed from the woven fabric in a wet-process in the textile finishing industry. The α -amylases remove selectively the size and do not attack the fibers	<i>Bacillus</i> stain
Pulp and paper	De-sizing; Starch- coating, de-inking, drainage improvement	As for textiles, sizing of paper is performed to protect the paper against mechanical damage during processing. It also improves the quality of the finished paper. The size enhances the stiffness and strength in paper. It also improves the erasibility and is a good coating for the paper.	-

modified starch, and amylose-lipid complex.37

Production method

Isolated microbial strains used in starch processing must be able to produce the enzyme on the industrial scale. To produce alpha-amylase on the industrial scale, submerged fermentation (SMF), and solid state fermentation (SSF) are frequently utilized. SMF is used to produce bioproducts from broth medium such as molasses. This method requires remarkable moisture that is crucial for the growth of microorganisms (mostly bacteria) in the medium to produce alpha-amylase.¹⁴ This high moisture also provides easily applicable processes for sterilization, production, purification, controllable temperature, nutrient, pH, and etc; for instance, amylase production as a microbial source using *Bacillus* sp.⁴⁰

SSF is used for the production of alpha-amylase from easily recycled waste materials such as paper. This method needs low moisture, which can be regarded as an advantage. Further advantages of SSF include more straightforward equipment, more production, and less effluent production. However, SSF is slower than SMF in utilizing of substrates by microorganisms. Therefore, SSF is the most common method for the production of alphaamylase.¹⁴ *Bacillus thuringiensis* and *Bacillus cereus* have been frequently used in co-culture condition to produce alpha-amylase through SSF.⁴¹

Enzyme activity assay

The digestive activity of alpha-amylases is measured via several colorimetric methods, including dinitrosalicylic acid method (DNS), Nelson–Somogyi, and Iodine method.⁴² DNS is an alkaline reagent that attaches to the reducing sugars and then color changes can be detected by UV absorbance at 540 nm⁴³ (Figure 1). Amylases, xyloglucanases, pectinases, and β -mannanases are assessed by DNS method. A Drawback of this method is the lack of information on its stoichiometric properties with oligosaccharides⁴⁴ (Figure 1).

In iodine method, Lugol interacts with starch and forms complexes and alpha-amylase degrades starches and reduces UV absorbance at 580 nm.³¹ Figure 2 shows measurement of the alpha-amylase activity by iodine method. Also, While Nelson – Somogyi method is used for measuring of α -amylase activity. This method is 10 times more sensitive than DNS method.⁴⁴ DNS method



Figure 1. Assaying of alpha-amylase activity by DNS method.



Figure 2. Assaying of alpha-amylase activity by iodine method.



Figure 3. Assaying of alpha-amylase activity by Nelson-Somogyi method.

process is described in Figure 3.

Medium composition factors

The condition of the culture medium must be regarded for enzyme production. Medium composition and physical conditions can directly affect the production of alpha-amylase. Different factors have been shown to affect enzyme production, here these factors are presented.

Carbon source

Some of the most known substrates as a carbon source for microorganisms to produce alpha-amylase include maltose, glucose, and sucrose. A study on *Aspergillus oryzae* S2 showed that the proper concentration of starch (10%) are the best carbon source to produce alphaamylase.⁴⁵ *Penicillium notatum* IBGE 03 is another fungus that is used for alpha-amylase production; this fungus uses molasses as its favorable carbon source.⁴² *Bacillus subtilis* as another microorganism is used for the production of alpha-amylase in SSF and it needs glucose for optimized production of alpha-amylase is 0.02 g/g.⁴⁶ Another study on *Bacillus* family showed that the glucose concentration for optimum production of alpha-amylase by *Bacillus amyloliquefaciens* was 10.50 g/L.⁴⁷

pH optimization

pH plays a significant role in the production and secretion of alpha-amylase. Microorganisms need an appropriate growth condition for production of amylase, for example, most fungi grow in the light acidic condition; however, bacteria need a neutral pH (around pH 7).⁴⁸

Kluyveromyces marxianus IF0 0288 at pH 6.13 produces the highest quantity of alpha-amylase.⁴⁹ The optimized pH for the production of alpha-amylase by *Penicillium notatum* IBGE 03 has been reported to be 5.5.⁴² Also, *Bacillus sp.* MB6 produces a remarkable quantity of alphaamylase at pH 6.⁵⁰

Nitrogen source

As nitrogen contents in culture medium have a significant role in the growth of microorganisms. Different nitrogen sources have been widely studied for the optimization of alpha-amylase production, including the organics such as yeast extract, soybean, and peptone, which are the most applicable nitrogen sources in culture medium; other sources are the inorganics, such as ammonium hydrogen phosphate, ammonium sulfate, and ammonium chloride.¹⁴

Bacillus amyloliquefaciens KCP2 produces a significant amount of alpha-amylase using ammonium sulfate (0.2 g) as an inorganic nitrogen source under SSF.⁵¹ Also, *Bacillus amyloliquefaciens* has been shown to use yeast extract (2 g/L), as an organic nitrogen source to produce alphaamylase.⁵² A study on *Penicillium notatum* IBGE 03 used corn steep as the organic nitrogen source to optimize alpha-amylase production.⁴² Soybean has been used as a source of nitrogen by *Aspergillus oryzae* CBS 819.72, which produced alpha-amylase on optimized condition.³⁶

Metal ions

Ca²⁺ ions due to presence in alpha-amylase structure play an important role in alpha-amylase production; In most culture media, calcium chloride (CaCl₂) is added to produce alpha-amylase.⁴⁸

Bacillus amyloliquefaciens has been shown to use CaCl₂ (0.0275 M) as a crucial factor to produce alpha-amylase.⁵³ Also, further study by Zhao et al showed that CaCl₂ (2 g/L) plays an important role in the production of alpha-amylase by *Bacillus amyloliquefaciens*.⁵² *Penicillium* sp., another microbial source is remarkably dependent on CaCl₂ to produce high levels of alpha-amylase.⁵⁴

Enzyme optimization methods

Hydrolytic enzymes contribute to global business so there is an essential need for optimization of these enzymes.

Some methods have been developed for optimization of these enzymes, including response surface methodology⁵⁵ and Taguchi methods.

Response surface methodology

Due to the vital role for the optimization of factors and developing a novel experiment, RSM is considered as an important part of the experimental design.⁵² RSM is comprised of statistical and mathematical techniques consisting of two methods for optimization, including Box–Behnken designs and Central Composite Design (CCD).⁵⁶ Minitab[®] 16.1.0 is the most commonly used software for the optimization of amylase production by *Enterococcus faecium* DMF78.⁵⁷ On the other hand, design expert (version 8.0) is frequently utilized to improve the alpha-amylase production from thermostable and alklophilic alpha-amylase from *Bacillus amyloliquefaciens* KCP2.⁵¹

Box–Behnken designs: The Box–Behnken designs have been used for RSM experiment to enhance alpha-amylase production from *Aspergillus oryzae* S2, *Aspergillus oryzae* CBS 819.72, *Bacillus laterosporus*, and *Bacillus amyloliquefaciens*^{36,45,52,58}(Figure 4).

CCD: Figure 5 shows CCD. Bacillus amyloliquefaciens, Streptomyces erumpens MTCC 731, Bacillus amyloliquefaciens KCP2, and Aspergillus oryzae S2 synthesize alpha-amylase by CCD method.^{51,59-61}

Purification

Purification of the produced enzyme may include the main part of the enzyme production cost, in particular when there is a need for stringent purification. There



Figure 4. Schemes of Box–Behnken design.



Figure 5. Schemes of Central Composite Design (CCD).

are different methods of enzyme purification; however, selecting the final method is dependent on the market, cost, final quality, and available technology.⁶²

Mostly, after primary isolation via precipitation or membrane separations, enzymes are purified by chromatographic methods.^{11,63,64} Regarding the needs for large-scale production of enzyme, the purification methods have been improved to provide higher efficiency, cost-effectiveness, and faster purification in less processing steps.⁶⁵ Table 3 shows several purification methods for amylase in bacteria.

Stability of alpha-amylase

Industrial enzymes should be stable in harsh conditions. One of the industrial enzymes is alpha-amylase, which needs to be stable against high temperatures, pH, oxidation, and other conditions that are inevitable during starch hydrolysis. So far, several methods have been utilized to achieve a stable amylase; the first, isolation and screening enzyme for extremophile microorganisms such as thermophile bacteria and genetically manipulating of other microorganisms; the second, improving enzyme stability via protein engineering, enzyme immobilization and adding of additives.⁶⁶

Enzyme recycling maybe regarded as a strategy to save the enzyme. The limitation of recycling of the enzyme was solved by immobilization method.⁶⁷ Also, immobilization provides higher stability, and continuous operation, and it is suitable for several various engineering designs and improves control of reaction.⁶⁸ Several methods have been used for Immobilization, including entrapment, crosslinking, covalent attachment, encapsulation, and adsorption.⁶⁶

Due to fungal alpha-amylases recognized as safe status, the alpha-amylases are more preferred in the starch industry but it needs to be more stable and costeffective. A study by He et al showed increased stability and longer service life of fungal alpha-amylases (AmyA1) via immobilization on magnetic nanoparticles.⁶⁹

Another method for enhancing enzyme stability is protein engineering, which is classified into two categories; the site-directed mutagenesis and the random mutagenesis. Improving stability by random mutagenesis within the whole length of a gene uses several methods such as UV irradiation, DNA shuffling, chemical mutation, and error-prone PCR.⁶⁶ Site-directed mutagenesis is an approach to produce new biocatalyst with a higher stability, activity, expiration, and solubility compared with wild protein. Appropriate amino acid residues are selected and manipulated, is the leading characteristic of the sitedirected mutagenesis.⁷⁰

A useful effect of mutagenesis on the stability of amylase was shown by Wang et al. Most of the Alpha-amylases are active in natural pH. Due to acidic pH of native starch slurry (3.2-45) deactivates the enzyme during starch processing, starch slurry must be adjusted to natural pH. Table 3. Purification methods for alpha amylase

Microorganism	Method of purification Partial purification by ethanol precipitation	
Thermostable, thermophilic, alkaline and chelator resistant amylase from an alkaliphilic <i>Bacillus sp.</i> Isolate ANT-6		
Marine Vibrio sp.	Substitute affinity method by insoluble corn stratch	78
Bacillus subtilis	High-speed counter-current chromatography	73
Bacillus amyloliquefaciens	Primary precipitation by ammonium sulphate then final purification by ion exchange chromatography	
Thermostable α -amylase from <i>Bacillus sp</i> . PS-7	Partial purification with ammonium sulphate and further purification by sp. PS-7 Sephadex G-75 gelfiltration followed by phenyl agarose 4XL hydrophobic interaction chromatography.	
Bacillus amyloliquefaciens	Affinity precipitation by alginate as the affinity matrix.	96
-	Affinity adsorption chromatography	95
-	Expanded bed chromatography	69

A recent report on alpha-amylase production by *Bacillus subtilis* showed that the mutated alpha-amylase (in the active site) is more active in acidic condition.⁷¹

Another method for improving the alpha-amylase stability is chemical modification.⁷² In the starch industry, high temperatures are required for synthesis of the product, which inactivates alpha-amylase. Siddiqui et al reported that the modification of carboxyl groups of TAA by L-arginine methyl ester dihydrochloride may improve the hydrolytic activity of alpha-amylase at 60°C.⁷³

Water is crucial for alpha-amylase activity; on the other hand, water can lead to denaturation of the enzyme. some stabilizing agents such as sugars or polyols can be added to alpha-amylase in order to change water structure and improve hydrophobic interaction strength in the enzyme structure.^{66,74} Samborska et al reported that sucrose, trehalose, and polyols can have a remarkable stabilizing effect on alpha-amylase at thermophilic condition.⁷⁵

Conclusion

The production of alpha-amylase at a large scale are crucial for industrial goals. Therefore, finding microorganisms with such potential to produce a high quantity of amylase is an important goal of the scientists. Some microorganisms from special environments such as saline soil can be used to achieve such goal. Also according to the microorganism sources, some types of amylases can be used to save energy via the development of the cold-active alpha-amylase. Also, protein engineering methods can be applied to produce enzymes with remarkable stability.

Ethical Issues

Not applicable.

Conflict of Interest

Authors declare that there is no conflict of interest.

Acknowledgments

This project (Ph.D.thesis) is financially supported by Tabriz University of Medical Sciences, Tabriz, Iran (Grant Number: 58056).

References

- Tarhriz V, Akbari Z, Dilmaghani A, Hamidi A, Hejazi MA, Hejazi MS. Bioreduction of iron and biosorption of heavy metals (Ni²⁺, Co²⁺, Pb²⁺) by a novel environmental bacterium, *Tabrizicola aquatica* RCRI19 t. *Asian J Water Environ Pollut* 2019;16(3):73-81. doi: 10.3233/AJW190035
- Chapeland-Leclerc F, Dilmaghani A, Ez-Zaki L, Boisnard S, Da Silva B, Gaslonde T, et al. Systematic gene deletion and functional characterization of histidine kinase phosphorelay receptors (HKRs) in the human pathogenic fungus Aspergillus fumigatus. Fungal Genet Biol 2015;84:1-11. doi: 10.1016/j.fgb.2015.09.005
- Dafe A, Etemadi H, Dilmaghani A, Mahdavinia GR. Investigation of pectin/starch hydrogel as a carrier for oral delivery of probiotic bacteria. *Int J Biol Macromol* 2017;97:536-43. doi: 10.1016/j.ijbiomac.2017.01.060
- Tarhriz V, Eyvazi S, Shakeri E, Hejazi MS, Dilmaghani A. Antibacterial and antifungal activity of novel freshwater bacterium *Tabrizicola aquatica* as a prominent natural antibiotic available in Qurugöl Lake. *Pharm Sci* 2020;26(1):88-92. doi: 10.34172/ps.2019.56
- 5. Prasad NK. *Enzyme Technology: Pacemaker of Biotechnology*. New Delhi: PHI Learning Pvt Ltd; 2011.
- Tarhriz V, Hamidi A, Rahimi E, Eramabadi M, Eramabadi P, Yahaghi E, et al. Isolation and characterization of naphthalene-degradation bacteria from Qurugöl Lake located at Azerbaijan. *Biosci Biotech Res Asia* 2014;11(2):715-22. doi: 10.13005/bbra/1326
- Avwioroko OJ, Anigboro AA, Unachukwu NN, Tonukari NJ. Isolation, identification and in silico analysis of alphaamylase gene of *Aspergillus niger* strain CSA35 obtained from cassava undergoing spoilage. *Biochem Biophys Rep* 2018;14:35-42. doi: 10.1016/j.bbrep.2018.03.006
- Delkash-Roudsari S, Zibaee A, Abbasi Mozhdehi MR. Digestive alpha-amylase of Bacterocera oleae Gmelin (Diptera: Tephritidae): biochemical characterization and effect of proteinaceous inhibitor. *J King Saud Univ Sci* 2014;26(1):53-8. doi: 10.1016/j.jksus.2013.05.003
- Ficko-Blean E, Stuart CP, Boraston AB. Structural analysis of CPF_2247, a novel alpha-amylase from *Clostridium perfringens. Proteins* 2011;79(10):2771-7. doi: 10.1002/ prot.23116
- 10. Tanyildizi MS, Özer D, Elibol M. Optimization of alpha-

amylase production by *Bacillus* sp. using response surface methodology. *Process Biochem* 2005;40(7):2291-6. doi: 10.1016/j.procbio.2004.06.018

- Burhan A, Nisa U, Gökhan C, Ömer C, Ashabil A, Osman G. Enzymatic properties of a novel thermostable, thermophilic, alkaline and chelator resistant amylase from an alkaliphilic *Bacillus* sp. isolate ANT-6. *Process Biochem* 2003;38(10):1397-403. doi: 10.1016/S0032-9592(03)00037-2
- 12. Tarhriz V, Mohammadzadeh F, Hejazi MS, Nematzadeh G, Rahimi E. Isolation and characterization of some aquatic bacteria from Qurugöl Lake in Azerbaijan under aerobic conditions. *Adv Environ Biol* 2011;5(10):3173-8.
- 13. Oboh G. Isolation and characterization of amylase from fermented cassava (*Manihot esculenta* Crantz) wastewater. *Afr J Biotechnol* 2005;4(10):1117-23.
- 14. Sundarram A, Murthy TP. Alpha-amylase production and applications: a review. *J Appl Environ Microbiol* 2014;2(4):166-75. doi: 10.12691/jaem-2-4-10
- 15. Abou-Elela G, El-Sersy NA, Wefky SH. Statistical optimization of cold adapted alpha-amylase production by free and immobilized cells of *Nocardiopsis aegyptia*. *J Appl Sci Res* 2009;5(3):286-92.
- Konsoula Z, Liakopoulou-Kyriakides M. Co-production of alpha-amylase and beta-galactosidase by Bacillus subtilis in complex organic substrates. *Bioresour Technol* 2007;98(1):150-7. doi: 10.1016/j.biortech.2005.11.001
- Kizhakedathil MPJ, Chandrasekaran SD. Media optimization for extracellular amylase production by *Pseudomonas balearica* vitps19 using response surface methodology. *Front Biol* 2018;13(2):123-9. doi: 10.1007/ s11515-018-1485-3
- Gazali FM, Suwastika IN. Thermostable α-amylase activity from thermophilic bacteria isolated from Bora Hot Spring, Central Sulawesi. *Journal of Physics: Conference Series* 2018: 979:012001. doi :10.1088/1742-6596/979/1/012001
- 19. Yuuki T, Nomura T, Tezuka H, Tsuboi A, Yamagata H, Tsukagoshi N, et al. Complete nucleotide sequence of a gene coding for heat- and pH-stable alpha-amylase of *Bacillus licheniformis*: comparison of the amino acid sequences of three bacterial liquefying alpha-amylases deduced from the DNA sequences. *J Biochem* 1985;98(5):1147-56. doi: 10.1093/oxfordjournals.jbchem.a135381
- 20. Sudan SK, Kumar N, Kaur I, Sahni G. Production, purification and characterization of raw starch hydrolyzing thermostable acidic alpha-amylase from hot springs, India. *Int J Biol Macromol* 2018;117:831-9. doi: 10.1016/j. ijbiomac.2018.05.231
- 21. Prakash O, Jaiswal N. Alpha-amylase: an ideal representative of thermostable enzymes. *Appl Biochem Biotechnol* 2010;160(8):2401-14. doi: 10.1007/s12010-009-8735-4
- Shafiei M, Ziaee AA, Amoozegar MA. Purification and characterization of an organic-solvent-tolerant halophilic alpha-amylase from the moderately halophilic *Nesterenkonia* sp. strain F. J Ind Microbiol Biotechnol 2011;38(2):275-81. doi: 10.1007/s10295-010-0770-1
- Madern D, Ebel C, Zaccai G. Halophilic adaptation of enzymes. *Extremophiles* 2000;4(2):91-8. doi: 10.1007/ s007920050142
- 24. Fukuchi S, Yoshimune K, Wakayama M, Moriguchi M, Nishikawa K. Unique amino acid composition of proteins

in halophilic bacteria. J Mol Biol 2003;327(2):347-57. doi: 10.1016/s0022-2836(03)00150-5

- Bieger B, Essen LO, Oesterhelt D. Crystal structure of halophilic dodecin: a novel, dodecameric flavin binding protein from *Halobacterium salinarum*. Structure 2003;11(4):375-85. doi: 10.1016/s0969-2126(03)00048-0
- Dou S, Chi N, Zhou X, Zhang Q, Pang F, Xiu Z. Molecular cloning, expression, and biochemical characterization of a novel cold-active alpha-amylase from *Bacillus* sp. dsh19-1. *Extremophiles* 2018;22(5):739-49. doi: 10.1007/s00792-018-1034-7
- 27. Wang X, Kan G, Ren X, Yu G, Shi C, Xie Q, et al. Molecular cloning and characterization of a novel alpha-amylase from antarctic sea ice bacterium *Pseudoalteromonas* sp. M175 and its primary application in detergent. *Biomed Res Int* 2018;2018:3258383. doi: 10.1155/2018/3258383
- Kuddus M, Roohi, Arif JM, Ramteke PW. An overview of cold-active microbial alpha-amylase: adaptation strategies and biotechnological potentials. *Biotechnology* 2011;10(3):246-58. doi: 10.3923/biotech.2011.246.258
- 29. Nithya K, Muthukumar C, Kadaikunnan S, Alharbi NS, Khaled JM, Dhanasekaran D. Purification, characterization, and statistical optimization of a thermostable alpha-amylase from desert actinobacterium *Streptomyces fragilis* DA7-7. *3 Biotech* 2017;7(5):350. doi: 10.1007/s13205-017-0981-5
- Gupta A, Gautam N, Modi DR. Optimization of alphaamylase production from free and immobilized cells of *Aspergillus niger. J Biotechnol Pharm Res* 2010;1(1):1-8.
- Karim KMR, Husaini A, Sing NN, Sinang FM, Roslan HA, Hussain H. Purification of an alpha amylase from *Aspergillus flavus* NSH9 and molecular characterization of its nucleotide gene sequence. *3 Biotech* 2018;8(4):204. doi: 10.1007/s13205-018-1225-z
- de Souza PM, de Oliveira Magalhaes P. Application of microbial alpha-amylase in industry - a review. *Braz J Microbiol* 2010;41(4):850-61. doi: 10.1590/s1517-83822010000400004
- El-Fallal A, Abou-Dobara MA, El-Sayed A, Omar N. Starch and microbial alpha-amylase: from concepts to biotechnological applications. In: Chang CF, ed. *Carbohydrates: Comprehensive Studies on Glycobiology* and Glycotechnology. Croatia: InTech; 2012. p. 459-89. doi: 10.5772/51571
- Rana N, Walia A, Gaur A. Alpha-amylases from microbial sources and its potential applications in various industries. *Natl Acad Sci Lett* 2013;36(1):9-17. doi: 10.1007/s40009-012-0104-0
- Demirci A, Izmirlioglu G, Ercan D. Fermentation and enzyme technologies in food processing. In: Clark S, Jung S, Lamsal B, eds. *Food Processing: Principles and Applications*. 2nd ed. Chichester, West Sussex, UK: Wiley-Blackwell; 2014. p. 107-36. doi: 10.1002/9781118846315.ch6
- 36. Kammoun R, Naili B, Bejar S. Application of a statistical design to the optimization of parameters and culture medium for alpha-amylase production by *Aspergillus* oryzae CBS 819.72 grown on gruel (wheat grinding byproduct). *Bioresour Technol* 2008;99(13):5602-9. doi: 10.1016/j.biortech.2007.10.045
- Zhao X, Andersson M, Andersson R. Resistant starch and other dietary fiber components in tubers from a highamylose potato. *Food Chem* 2018;251:58-63. doi: 10.1016/j.

foodchem.2018.01.028

- Lee BH, Hamaker BR. Number of branch points in alpha-limit dextrins impact glucose generation rates by mammalian mucosal alpha-glucosidases. *Carbohydr Polym* 2017;157:207-13. doi: 10.1016/j.carbpol.2016.09.088
- Richardson TH, Tan X, Frey G, Callen W, Cabell M, Lam D, et al. A novel, high performance enzyme for starch liquefaction. Discovery and optimization of a low pH, thermostable alpha-amylase. J Biol Chem 2002;277(29):26501-7. doi: 10.1074/jbc.M203183200
- 40. Vidyalakshmi R, Paranthaman R, Indhumathi J. Amylase production on submerged fermentation by *Bacillus* spp. *World J Chem* 2009;4(1):89-91.
- 41. Abdullah R, Naeem N, Aftab M, Kaleem A, Iqtedar M, Iftikhar T, et al. Enhanced production of alpha amylase by exploiting novel bacterial co-culture technique employing solid state fermentation. *Iran J Sci Technol Trans A Sci* 2018;42(2):305-12. doi: 10.1007/s40995-016-0015-x
- Ahmed K, Munawar S, Khan MA. Cultural conditions for maximum alpha-amylase production by *Penicillium notatum* IBGE 03 using shaken flask technique of submerged fermentation. *Pure Appl Biol* 2015;4(3):306-12. doi: 10.19045/bspab.2015.43005
- Raul D, Biswas T, Mukhopadhyay S, Kumar Das S, Gupta S. Production and partial purification of alpha amylase from Bacillus subtilis (MTCC 121) using solid state fermentation. *Biochem Res Int* 2014;2014:568141. doi: 10.1155/2014/568141
- 44. Gusakov AV, Kondratyeva EG, Sinitsyn AP. Comparison of two methods for assaying reducing sugars in the determination of carbohydrase activities. *Int J Anal Chem* 2011;2011:283658. doi: 10.1155/2011/283658
- Naili B, Sahnoun M, Bejar S, Kammoun R. Optimization of submerged *Aspergillus oryzae* S2 alpha-amylase production. *Food Sci Biotechnol* 2016;25(1):185-92. doi: 10.1007/ s10068-016-0028-4
- Hassan H, Karim KA. Optimization of alpha amylase production from rice straw using solid-state fermentation of Bacillus subtilis. *Int J Sci Environ Technol* 2015;4(1):1-16.
- 47. Uygut MA, Tanyildizi MŞ. Optimization of alpha-amylase production by *Bacillus amyloliquefaciens* grown on orange peels. *Iran J Sci Technol Trans A Sci* 2018;42(2):443-9. doi: 10.1007/s40995-016-0077-9
- Sivaramakrishnan S, Gangadharan D, Nampoothiri KM, Soccol CR, Pandey A. Alpha-amylases from microbial sources–an overview on recent developments. *Food Technol Biotechnol* 2006;44(2):173-84.
- Stergiou P-Y, Foukis A, Theodorou L, Papagianni M, Papamichael E. Optimization of the production of extracellular alpha-amylase by *Kluyveromyces marxianus* IF0 0288 by response surface methodology. *Braz Arch Biol Technol* 2014;57(3):421-6. doi: 10.1590/S1516-8913201401485
- Paul JS, Lall BM, Jadhav SK, Tiwari KL. Parameter's optimization and kinetics study of alpha-amylase enzyme of *Bacillus* sp. MB6 isolated from vegetable waste. *Process Biochem* 2017;52:123-9. doi: 10.1016/j.procbio.2016.10.005
- Prajapati VS, Trivedi UB, Patel KC. A statistical approach for the production of thermostable and alklophilic alphaamylase from *Bacillus amyloliquefaciens* KCP2 under solid-state fermentation. *3 Biotech* 2015;5(2):211-20. doi:

 $10.1007/s13205\hbox{-}014\hbox{-}0213\hbox{-}1$

- 52. Zhao W, Zheng J, Wang YG, Zhou HB. A marked enhancement in production of amylase by *Bacillus amyloliquefaciens* in flask fermentation using statistical methods. J Cent South Univ Technol 2011;18(4):1054-62. doi: 10.1007/s11771-011-0803-6
- Gangadharan D, Sivaramakrishnan S, Nampoothiri KM, Sukumaran RK, Pandey A. Response surface methodology for the optimization of alpha amylase production by *Bacillus amyloliquefaciens*. *Bioresour Technol* 2008;99(11):4597-602. doi: 10.1016/j.biortech.2007.07.028
- Abdullah R, Nadeem S, Iqtedar M, Kaleem A, Iftikhar T, Naz S. Influence of growth conditions on enhanced production of alpha amylase from *Penicillium* species in solid state fermentation. *Indian J Biotechnol* 2017;16(3):426-32.
- 55. Magro M, Nauta S, Simsek C, Onuma Y, Garg S, van der Heide E, et al. Value of the SYNTAX score in patients treated by primary percutaneous coronary intervention for acute ST-elevation myocardial infarction: the MI SYNTAXscore study. *Am Heart J* 2011;161(4):771-81. doi: 10.1016/j.ahj.2011.01.004
- Khuri AI, Mukhopadhyay S. Response surface methodology. Wiley Interdiscip Rev Comput Stat 2010;2(2):128-49. doi: 10.1002/wics.73
- 57. Duque SM, Dizon EI, Merca FE, Flores DM. Optimization of raw-starch-digesting amylase (RSDA) production medium for *Enterococcus faecium* DMF78. *Int Food Res J* 2016;23(3):1280-8.
- Kumar NM, Karthikeyan S, Jayaraman G. Thermostable alpha-amylase enzyme production from *Bacillus laterosporus:* statistical optimization, purification and characterization. *Biocatal Agric Biotechnol* 2013;2(1):38-44. doi: 10.1016/j.bcab.2012.10.005
- Sahnoun M, Kriaa M, Elgharbi F, Ayadi DZ, Bejar S, Kammoun R. Aspergillus oryzae S2 alpha-amylase production under solid state fermentation: optimization of culture conditions. *Int J Biol Macromol* 2015;75:73-80. doi: 10.1016/j.ijbiomac.2015.01.026
- Tanyildizi MS, Elibol M, Özer D. Optimization of growth medium for the production of alpha-amylase from *Bacillus amyloliquefaciens* using response surface methodology. J Chem Technol Biotechnol 2006;81(4):618-22. doi: 10.1002/ jctb.1445
- 61. Kar S, Datta TK, Ray RC. Optimization of thermostable alpha-amylase production by *Streptomyces erumpens* MTCC 7317 in solid-state fermentation using cassava fibrous residue. *Braz Arch Biol Technol* 2010;53(2):301-9. doi: 10.1590/S1516-89132010000200008
- 62. Madhavan Nampoothiri K, Nair NR, John RP. An overview of the recent developments in polylactide (PLA) research. *Bioresour Technol* 2010;101(22):8493-501. doi: 10.1016/j. biortech.2010.05.092
- 63. Triveni R. Production, *Characterization and Utilization of Gelling Polysaccharide of Bacteria* [thesis]. University of Mysore; 2000.
- 64. Coronado M, Vargas C, Hofemeister J, Ventosa A, Nieto JJ. Production and biochemical characterization of an alpha-amylase from the moderate halophile *Halomonas meridiana*. *FEMS Microbiol Lett* 2000;183(1):67-71. doi: 10.1111/j.1574-6968.2000.tb08935.x
- 65. Amritkar N, Kamat M, Lali A. Expanded bed affinity

purification of bacterial alpha-amylase and cellulase on composite substrate analogue-cellulose matrices. *Process Biochem* 2004;39(5):565-70. doi: 10.1016/S0032-9592(03)00123-7

- 66. Dey TB, Kumar A, Banerjee R, Chandna P, Kuhad RC. Improvement of microbial α-amylase stability: strategic approaches. *Process Biochem* 2016;51(10):1380-90. doi: 10.1016/j.procbio.2016.06.021
- 67. Talekar S, Pandharbale A, Ladole M, Nadar S, Mulla M, Japhalekar K, et al. Carrier free co-immobilization of alpha amylase, glucoamylase and pullulanase as combined cross-linked enzyme aggregates (combi-CLEAs): a tri-enzyme biocatalyst with one pot starch hydrolytic activity. *Bioresour Technol* 2013;147:269-75. doi: 10.1016/j. biortech.2013.08.035
- Cordeiro AL, Lenk T, Werner C. Immobilization of Bacillus licheniformis alpha-amylase onto reactive polymer films. J Biotechnol 2011;154(4):216-21. doi: 10.1016/j. jbiotec.2011.04.008
- 69. He L, Mao Y, Zhang L, Wang H, Alias SA, Gao B, et al. Functional expression of a novel alpha-amylase from Antarctic psychrotolerant fungus for baking industry and its magnetic immobilization. *BMC Biotechnol* 2017;17(1):22. doi: 10.1186/s12896-017-0343-8
- 70. Sindhu R, Binod P, Madhavan A, Beevi US, Mathew AK,

Abraham A, et al. Molecular improvements in microbial alpha-amylases for enhanced stability and catalytic efficiency. *Bioresour Technol* 2017;245(Pt B):1740-8. doi: 10.1016/j.biortech.2017.04.098

- 71. Wang CH, Liu XL, Huang RB, He BF, Zhao MM. Enhanced acidic adaptation of *Bacillus subtilis* Ca-independent alphaamylase by rational engineering of pKa values. *Biochem Eng* J 2018;139:146-53. doi: 10.1016/j.bej.2018.08.015
- 72. Habibi AE, Khajeh K, Nemat-Gorgani M. Chemical modification of lysine residues in *Bacillus licheniformis* alpha-amylase: conversion of an endo- to an exo-type enzyme. *J Biochem Mol Biol* 2004;37(6):642-7. doi: 10.5483/ bmbrep.2004.37.6.642
- 73. Siddiqui KS, Poljak A, De Francisci D, Guerriero G, Pilak O, Burg D, et al. A chemically modified alpha-amylase with a molten-globule state has entropically driven enhanced thermal stability. *Protein Eng Des Sel* 2010;23(10):769-80. doi: 10.1093/protein/gzq051
- Khajeh K, Nemat-Gorgani M. Comparative studies on a mesophilic and a thermophilic alpha-amylase. *Appl Biochem Biotechnol* 2001;90(1):47-55. doi: 10.1385/ABAB:90:1:47
- 75. Samborska K, Guiavarc'h Y, Van Loey A, Hendrickx M. The thermal stability of *Aspergillus oryzae* alpha-amylase in presence of sugars and polyols. *J Food Process Eng* 2006;29(3):287-303. doi: 10.1111/j.1745-4530.2006.00062.x